Linking Genes to Microbial Biogeochemical Cycling Lessons from Arsenic

Zhu, Y.-G., Xue, X.-M., Kappler, A., Rosen, B. P., & Meharg, A. A. (2017). Linking Genes to Microbial Biogeochemical Cycling Lessons from Arsenic. *Environmental science & technology*, *51*(13), 7326-7339. https://doi.org/10.1021/acs.est.7b00689

Published in:
Environmental science & technology

Document Version:
Peer reviewed version

Queen's University Belfast - Research Portal:
Link to publication record in Queen's University Belfast Research Portal

Linking genes to microbial biogeochemical cycling: lessons from arsenic

Yong-Guan Zhu, [†, ‡, #, *] Xi-Mei Xue, [†, #] Andreas Kappler, [‖] Barry P Rosen, [§] Andrew A Meharg [⊥]

[†] Key Laboratory of Urban Environment and Health, Institute of Urban Environment, Chinese Academy of Sciences, Xiamen, China

[‡] State Key Laboratory of Urban and Regional Ecology, Research Center for Eco-Environmental Sciences, Chinese Academy of Sciences, Beijing, China

[‖] Geomicrobiology, Center for Applied Geosciences, University of Tübingen, Tübingen 72076, Germany

[§] Department of Cellular Biology and Pharmacology, Herbert Wertheim College of Medicine, Florida International University, Miami, FL 33199, U.S.A.

[⊥] Institute for Global Food Security, Queen's University Belfast, Belfast BT9 5HN, United Kingdom

[#]These authors contributed equally to the article.

*Address correspondence to Yong-Guan Zhu

Address: Institute of Urban Environment, Chinese Academy of Sciences, 1799 Jimei Road, Xiamen 361021, China, Phone number: +86(0)592 6190997, Fax number: +86(0)592 6190977, Email address: ygzhu@rcees.ac.cn

Abstract

The biotransformation of arsenic is highly relevant to the arsenic biogeochemical cycle. Identification of the molecular details of microbial pathways of arsenic biotransformation coupled with analyses of microbial communities by meta-omics can provide insights into detailed aspects of the complexities of this biocycle. Arsenic transformations couple to other biogeochemical cycles, and to the fate of both nutrients and other toxic environmental contaminants. Microbial redox metabolism of iron, carbon, sulfur and nitrogen affects the redox and bioavailability of arsenic species. In this critical review we illustrate the biogeochemical processes and genes involved in arsenic biotransformations. We discuss how current and future metagenomic-, metatranscriptomic-, metaproteomic-, and metabolomic-based methods will help to decipher individual microbial arsenic transformation processes, and their connections to other biogeochemical cycle. These insights will allow future use of microbial metabolic capabilities for new biotechnological solutions to environmental problems. To understand the complex nature of inorganic and organic arsenic species and the fate of environmental arsenic will require integrating systematic approaches with biogeochemical modeling. Finally, from the lessons learned from these studies of arsenic biogeochemistry, we will be able to predict how the environment changes arsenic, and, in response, how arsenic biotransformations change the environment.

1. Introduction

Biogeochemical cycles are interconnected through redox reactions and other biotransformations. [1] Biogeochemical processes such as the cycling of a particular element are likely to be mediated by multiple microbes and are often linked to other biogeochemical processes. For example, redox changes of arsenic are mediated by diverse arsenate (As(V))-reducing and arsenite (As(III))-oxidizing microbes. Arsenic biogeochemical cycling is often coupled to the cycling of iron (Fe), [2] carbon (C) [3] and nitrogen (N), [4] and to the dynamics of elements/ions associated with the arsenic redox cycle, such as sulfur (S). [5] Coupling of biogeochemical cycles has recently received attention. The study of coupled biogeochemical cycles offers a scientific basis for major current environmental problems. [6]

The proteins catalyzing physiological processes in living organisms are influenced by geological, physical and chemical forces and therefore continuously evolve and redistribute chemical species involved in biogeochemical cycles. Genetic analysis is the key to understand the arsenic biogeochemical cycle. Once the genes associated with the reactions and the environmental signals that affect gene expression are understood, we will be able to predict how microbial metabolism influences arsenic biogeochemical cycling. In this review, we focus on the known genes involved in arsenic biotransformations and the effect of other elements on arsenic biogeochemistry. We highlight the effects of other elements on arsenic metabolism and the current state of meta-omics research in microbial arsenic metabolism. Finally, we discuss how integration of meta-omics information into biogeochemical models can allow us to predict the possible biotransformation of other elements.

2. Arsenic metabolism: from genes to biogeochemical processes

Organisms have evolved various strategies to transform arsenic for detoxification or energy metabolism. [7] An overview about the enzymatic pathways for arsenic biotransformations is presented in Figure 1, and related microbial genes are summarized in Table 1.

2.1 The arsenic redox cycle

The earliest microorganisms evolved in an anoxic environment, where the predominant arsenic species was most probably reduced As(III), with little oxidized As(V). The physiological activities of the earliest microorganisms were, therefore, largely driven by anaerobic metabolic processes, [8] and we propose that As(III) bioavailability was a driving force for the evolution or acquisition of genes encoding anaerobic respiratory pathways. [33] For example, the photosynthetic purple sulfur bacterium *Ectothiorhodospira* PHS-1 carries out anoxygenic photosynthesis using As(III) as an electron donor in the light [34, 35, 36] and uses As(V) as an electron acceptor in the dark. [37] The chemolithoautotrophic As(III)-oxidizer *Alkalilimnicola ehrlichii* MLHE-1 utilizes As(III) as an electron donor and nitrate as an electron acceptor in energy-generating respiratory chains. [38] Microorganisms with similar metabolic versatility probably evolved quite early. These microbes could cope with extreme growth conditions, such as high concentrations of As(III) or low oxygen, similar to those that existed in an primordial anoxic biosphere.

Since As(III) was probably the primary bioavailable arsenic species on the early anoxic Earth, it was the inevitable choice for organisms to utilize As(III) as an electron donor to produce energy. [39] As(III) oxidation is catalyzed by the enzyme As(III) oxidase, which is composed of two different subunits, a large subunit (α) having molybdopterin and a [3Fe-4S] cluster (AioA) and a smaller subunit (β)

incorporating a Rieske-type [2Fe-2S] cluster (AioB). [40] The cluster of *aioA* and *aioB* genes (*aio* operon) usually consists of *aioS* and *aioR* genes, encoding for a two-component signal transduction pair, AioS (sensor histidine kinase)/AioR (transcriptional regulator), which regulates expression of *aio* genes via recognizing As(III). [9] The operon sometimes has an *aioX* gene that encodes an As(III)-binding protein involved in As(III)-based signaling and regulation of As(III) oxidation, [12] a *cytC* gene encoding a cytochrome c that is required for efficient As(III) oxidation in *Ochrobactrum tritici* SCII24, [41] or a *moeA* gene encoding MoeA protein that synthesizes the molybdenum cofactor of AioAB oxidase. [9] Recently, a new type of As(III) oxidase, ArxA that exhibited both As(V) reductase and As(III) oxidase activities *in vitro*, [10] was identified in *A. ehrlichii* MLHE-1. [42] In *Ectothiorhodospira* sp. PHS-1 these genes code for As(III) oxidation coupled to photosynthesis .[35, 11] In addition to *arxA*, the MLHE-1 and PHS-1 *arx* operons, contain four other genes *arxB2*, *arxB*, *arxC*, and *arxD*, that encode two proteins with [4Fe-4S] centers, a membrane anchoring and quinol oxidoreductase subunit and a TorD-like molybdoenzyme chaperone respectively. [11] An adjacent and divergent gene cluster, *arxXSR,* encodes putative regulatory proteins, a periplasmic substrate-binding protein specific for phosphate (ArxX), a two-component histidine kinase sensor (ArxS), and a response regulator (ArxR). [11] ArxA has higher sequence similarity to the ArrA subunit than to AioA, and fills the phylogenetic gap between As(III) oxidases and As(V) reductases. [42, 11]

Note that As(III) oxidation by anaerobes would have produced As(V) in the absence of an oxygen-containing atmosphere, which opened a niche for As(V)-respiring microbes prior to the Great Oxidation Event (GOE). [33] Dissimilatory As(V)-respiring prokaryotes (DARPs) evolved pathways to take advantage of the

116 appearance of As(V) as a terminal electron acceptor. This new energy-generating
117 respiratory chain utilized the respiratory As(V) reductase, ArrAB, that reduce the less
118 toxic As(V) to the more toxic and potentially more mobile As(III). [40, 43, 44] ArrAB is a
119 heterodimer consisting of a large catalytic subunit (ArrA) and a small subunit (ArrB).
120 [15, 16] The *arr* operon also includes *arrC*, *arrD*, *arrS*, and *arrR*. Their gene products are
121 ArrC, a membrane-bound As(V) reductase subunit, ArrD, a As(V) reductase chaperon,
122 ArrS, a sensor histidine kinase and ArrR, a transcriptional regulator respectively. [17] A
123 phylogenetic analysis was conducted to search for molybdenum-b*is* (pyranopterin
124 guanine dinucleotide)-containing catalytic subunits of representative enzymes. This
125 complex iron sulfur molybdoenzyme family includes Arr, Aio, Arx, polysulfide
126 reductase, and nitrate reductase. The results indicate that Arr clusters most likely
127 evolved from polysulfide reductases. [17]

128 After the GOE, As(III) in oceans mostly oxidized to As(V), a new environmental
129 toxin. As(V) enters the cells of most organisms adventitiously via phosphate uptake
130 systems. [45] As a consequence, early life had to evolve novel strategies for coping with
131 new (potentially toxic) arsenic species. As described in more detail below, nearly
132 every extant microbe has ArsB or Acr3 efflux permeases for As(III) detoxification, so
133 it is reasonable to assume that organisms that arose before the GOE already had an
134 As(III) efflux system. When As(V) became the predominant soluble species, all cells
135 had to do was to reduce As(V) to As(III), the substrate of ArsB or Acr3, and they
136 would become resistant to As(V). A number of independently-evolved As(V)
137 reductases arose in a variety of organisms using a small molecular mass protein As(V)
138 reductases (one of several types of ArsC or Acr2 reductases). The ArsC system
139 conferred by the *ars* operon is the most well studied mechanism of arsenic
140 detoxification and resistance (for details see the previous review). [18] Most recently, a

glutathione S-transferase B (GstB) was found to mediate an alternate pathway which conferred As(V) resistance to *E. coli* mutant cells lacking *arsC* by directly reducing As(V) to As(III). [21] These enzymes all use small molecule proteins such as glutaredoxin (Grx) or reduced glutathione (GSH) coupled to thioredoxin (Trx) as electron donor. The Acr2 reductases evolved from proteins that incorporated the phosphate binding loop of phosphorprotein tyrosine phosphatases related to the cell phosphatase CDC25. [46] These phosphatase can be converted into As(V) reductases by just a few mutations, [47] indicating a facile evolutionary path.

2.2 The arsenic methylation cycle

In addition to oxidation and reduction of inorganic arsenic species, pathways for biotransformation of arsenic, including methylation and demethylation, organoarsenical degradation, evolved in early organisms. Interest in arsenic biomethylation began in 1800's with the observation that inorganic arsenic compounds used as wallpaper pigments were converted into Gosio gas (trimethylarsine) by fungi. More recent reports of methylated arsenical showed that arsenic methylation was widespread in the environment and detected in bacteria, [48] cyanobacteria, [49] algae, [23] protozoa. [50] Arsenic methylation is a common stratagem to detoxify arsenic. The highly toxic trivalent products are rapidly oxidized nonenzymatically in air to the less toxic pentavalent methylated arsenic species. Also, gaseous end-products such as trimethylarsine will emit to air, thus removing the product. Methylation is catalyzed by the enzyme As(III) S-adenosylmethionine (SAM) methyltransferase (EC 2.1.1.137), designated as AS3MT in animals and as ArsM in microorganisms. Expression of typical prokaryotic and archaeal *arsM* genes are regulated by the As(III)-responsive transcriptional repressor ArsR, [20] consistent with arsenic methylation being a detoxification pathway in the microbes. Expression of

arsM in some cyanobacteria appears to be constitutive, [51] indicating that alternate detoxification pathways are used by microorganisms in which the expression of *arsM* is not regulated. [52]

The degradation of environmental organoarsenicals has been documented for some time, [53, 54] while few molecular mechanisms for these reactions have been demonstrated. Recently, a two-step pathway of MSMA reduction and demethylation was elucidated. [55] Although no reductases of pentavalent organoarsenicals have been identified as yet, the enzyme, ArsI, which catalyzes demethylation of trivalent organoarsenicals, was identified and characterized from the environmental isolate bacterium *Bacillus* sp. MD1 [24] and from the cyanobacterium *Nostoc* sp. 7120. [56] ArsI, a non-heme iron-dependent dioxygenase with C-As lyase activity, cleaves the C-As bond in MAs(III), trivalent roxarsone, and other trivalent aromatic arsenicals. Putative ArsI orthologs were found only in bacterial species, suggesting that alternate pathways of organoarsenical demethylation might exist in other organisms. [24]

2.3 The organoarsenical cycle

The arsenic concentration in seawater is around 1 to 2 μg per liter, mainly inorganic arsenic that is usually transformed into complex organoarsenical compounds by marine organisms. [57] Arsenosugars, first identified in 1981, [58] are commonly detected water-soluble arsenic species present in marine algae; arsenobetaine is the most abundant arsenic species in the majority of marine animals. [57] More complex organoarsenicals have been identified with the improvement of analytical techniques in recent years. Since the structure of an arsenosugar phospholipid (AsPL) from a brown alga *Undaria pinnatifida* was first identified, [59] AsPL has been found in algae [60] and cyanobacteria. [61] Arsenic-containing fatty acids (AsFA) that were first identified in cod liver oil [62] have now been found in algae [63] and various fish species.

[64, 65] AsHC that were first reported in capelin [66] have been detected in fish [64, 65] and algae. [60] A new class of arsenolipids, trimethylarsenio fatty alcohols (TMAsFOH), was reported in Capelin oil. [67] Two new groups of arsenolipids, arsenic-containing phosphatidylcholines (AsPC) and arsenic-containing phosphatidylethanolamine (AsPE) from herring caviar, were characterized. [68] In total, more than 20 arsenosugars and 70 arsenolipids have been identified in marine organisms that live in low phosphate and high salt environments. These organoarsenicals are not toxic for the marine plants and animals, but their function in these marine organisms is not known. Phytoplankton can substitute sulfur- and nitrogen-containing membrane lipids for membrane phospholipids, [69] arsenolipids might be used in membranes in place of phospholipids due to the more similarity of As(V), than sulfate and nitrate, to inorganic phosphate. Thus As(V) could be used as a phosphate-sparing substitute in phosphate-limiting environments. A recent study on *Ectocarpus siliculosus* that was found to produce more arsenosugar phospholipids under low-phosphate than under normal phosphate conditions [70] supports this hypothesis.

These organoarsenicals may be toxic to organisms that cannot biosynthesize them. *In vitro* toxicological characterization of three arsenic-containing hydrocarbons showed that cytotoxicity of the arsenic-containing hydrocarbons was comparable to that of As(III) for cultured human bladder and liver cells, [71] and arsenolipids were metabolized by humans to dimethylated arsenical species (DMAs) and other small molecular arsenic compounds, then excreted in the urine. [72, 73]

Although several potential pathways have been proposed for the synthesis of complex organoarsenicals, [8] few genes involved in these biotransformation have been identified. Even less is known about the degradation of these organoarsenicals, and more studies are needed on this front.

2.4 The arsenic thiolation cycle

Thioarsenates ($H_3AsS_nO_{4-n}$) are the dominant arsenic species in alkaline, arsenic-rich, sulfidic environments. These play a significant role in the arsenic biochemical cycle in sulfidic geothermal environments. [74, 75, 76] Thioarsenates are transformed to As(V) and/or As(III) via exposing to oxidizing agents or increased pH, [77] by biological conversion by sulfur-oxidizing bacteria,[75, 78] or by abiotic decomposition (desulfidation) with subsequent biological oxidation. [77, 79] Thioarsenates contain reduced S^{2-} and oxidized As(V). They can serve both as electron donors and electron acceptors. For example, monothioarsenate can be used as an electron donor by *Thermocrinis ruber* OC 14/7/2, [80] and for anoxygenic photosynthesis by phototrophic purple sulfur bacteria growing in an alkaline environment.[81] Recently, the haloalkaliphilic bacterium MLMS-1 can grow chemolithotrophically by oxidizing the S^{2-} of monothioarsenate to S^0 or SO_4^{2-}, while concurrently reducing As(V) to As(III). [78] In summary, various microbes have evolved to utilize thioarsenates that are widespread in sulfidic environments.

2.5 Arsenic efflux pathways

The best way to deal with toxic arsenicals in cells is acquisition of an efficient efflux system. As(III) efflux systems have been intensively studied in both microbes and higher organisms. [82, 83] As(III) efflux in most bacteria is mediated by ArsB in an energy-dependent process, driven in *Staphylococcus aureus* by the membrane potential [84] and in *E. coli* by ATP hydrolysis that ArsA binds to ArsB to an ATP-driven arsenic-specific pump. [26] In the legume symbiont *S. meliloti*, an aquaglyceroporin (AqpS), instead of ArsB, has been identified to extrude As(III) from cells. [27] Acr3 has been shown to be an As(III)-efflux transporter in both bacteria and yeast, and provides a pathway for As(III) extrusion from cells. [19] In fact, genes for

Acr3 are more wide-spread in bacteria and archaea than are *arsB* genes. The cytosolic As(III)/glutathione complex sequestered into vacuoles by an ABC-type transporter, Ycf1p (yeast cadmium factor protein), is the second pathway for As(III) detoxification in yeast *S. cerevisiae*. [28]

Moreover, a novel mechanism for As(V) resistance was identified in a variety of microbes including *Pseudomonas aeruginosa*. [29] In these bacteria there are two genes that always go together, one encoding a typical glyceraldehyde-3-phosphate dehydrogenase (GAPDH) and the second one, called *arsJ*, that encodes an organoarsenical efflux permease (ArsJ). GAPDH uses As(V) and glyceraldehyde 3-phosphate to form the extremely unstable organoarsenical 1-arsenso-3-phosphoglycerate, which is extruded from cells by ArsJ and immediately breaks down into As(V) and 3-phosphoglycerate. The net reaction is effectively As(V) extrusion, and the coupled reaction confers As(V) resistance to these microbes, the only known efflux pathway for As(V). Meanwhile the bacterial permease, ArsP, from *Campylobacter jejuni*, was demonstrated to be an efflux system specific for trivalent organoarsenicals. [30] It is more selective for the ancient organoarsenical MAs(III) than for the recently anthropogenically-developed antimicrobial aromatic arsenical growth promoters such as trivalent roxarsone. More and more arsenic reductases and trivalent arsenic-specific transporters identified show that arsenic reduction and efflux play an important role in arsenic biogeochemical cycling.

3. Coupling of arsenic biogeochemical cycling to other elements

Any biogeochemical process, such as the cycling of a particular element, is likely to be mediated by more than one organism, and often linked to other fundamental biogeochemical processes. Arsenic biotransformations are often coupled to the

cycling of C, Fe, S and N. [85, 2, 86] The effect of turnover of these elements on microbes involved in arsenic biogeochemical cycling is summarized in Figure 2 which contains data based on previous studies [8, 87, 88, 89] and as described below.

3.1 The effects of iron on arsenic biogeochemical cycling

The chemical speciation of arsenic and arsenic mobility in natural environments are strongly dependent on redox potential and pH. Under oxic conditions, As(V) is the predominated arsenic species, present mainly as $H_2AsO_4^-$ at acid pH or $HAsO_4^{2-}$ at alkaline pH. In anoxic environments, arsenic occurs primarily as reduced As(III) ($As(OH)_3$ at neutral pH or $H_2AsO_3^-$ at alkaline pH), and more mobile than As(V). [90] Moreover, pH will impact arsenic interactions with Fe, the sorption of As(V) onto amorphous iron oxide and goethite is higher than that of As(III) below pH 5-6, As(V) and As(III) sorption onto iron oxide are both relatively high at neutral pH, As(III) is more easily adsorbed to iron oxide than As(V) above pH 7-8. [91]

Transformation of arsenic-bearing Fe mineral phases strongly affects the bioavailability of arsenic within soils and aquifers due to direct and indirect interactions between the arsenic and Fe cycles including mineral formation, transformation, dissolution and redox reactions. [89, 92, 93, 94] Previous studies from our laboratory showed that Fe plaque consisting of Fe(III) (oxyhydr)oxides, which was induced artificially through adding ferrous iron in solution to paddy soils, has high affinity for As(V), and reduced arsenic uptake by rice. [95, 96, 97]

Fe(III)-reducing bacteria modulate arsenic mobility in the rhizosphere. [98, 99, 100] Dissimilatory reduction of Fe(III) (oxyhydr)oxides to Fe(II) by dissimilatory iron-reducing bacteria (DIRB) can result either in the release of As(V) from poorly crystalline or more crystalline ferric minerals as well as from sorption sites within sediments, [101] or in the binding of arsenic to the formed Fe(II) minerals. [102, 103, 104, 105,

[106] DIRB are commonly present in rice paddy soil, and mediate dissimilatory reduction of Fe(III) on the rice root-plaque. [107] A study on the role of DIRB in arsenic release under a range of biogeochemical regimes indicated that Fe(III) reduction was stimulated by addition of acetate as a potential electron donor that resulted in a marked increase in the number of DIRB, reduction of As(V) to As(III), and arsenic release after Fe(III) reduction. [100] If DARPs were used as Fe(III)-reducers, *Shewanella* sp. ANA-3 [108] or *Sulfurospirillum barnesii* [109] could release both As(III) and Fe(II) from ferrihydrite containing As(V) by reducing solid-phase As(V) and Fe(III). Eventually, most of the ferrihydrite matrix was liberated as Fe(II) and As(III) if sufficient organic electron donor was present.[110] There is more aluminum in the crust than iron. However, *S. barnesii* does not reductively dissolve the As(V)-aluminum hydroxide precipitate, [111] so we did not include a detailed description of the effect of aluminum on arsenic biogeochemical cycling.

In addition to Fe(III) reduction that has the potential to mobilize or immobilize arsenic depending on geochemical conditions that lead to the formation of either dissolved Fe^{2+} or Fe(II) minerals, the formation of Fe(III) minerals under Fe(II)-oxidizing conditions has the potential to significantly immobilize arsenic and thus to lower its bioavailability. [112, 113] In particular for nitrate-reducing Fe(II)-oxidizing bacteria it has been shown that they form poorly soluble Fe(III) minerals and efficiently co-precipitate arsenic (Figure 2). [112, 113, 114] Besides nitrate-dependent Fe(II) oxidation, also microaerophilic Fe(II) oxidation has the potential to influence arsenic mobility and could even be used in biotechnical applications for arsenic removal, e.g. in drinking water filters [115] although it has been shown that in commercial drinking water filters the formation of iron biominerals by Fe(II)-oxidizing bacteria lowers arsenic removal from the water. [116] Moreover, a

thermo-acidophilic iron-oxidizing archaeon *Acidianus brierleyi*, has been used to immobilize As(III) in the copper refinery process by producing thermodynamically stable crystalline scorodite ($FeAsO_4 \cdot 2H_2O$).[117]

3.2 The effects of sulfur on arsenic biogeochemical cycling

More than 200 different arsenic-containing minerals have been found in the Earth's crust, and 20% are sulfides and sulfosalts. [118] The behavior of arsenic is affected by abiotic or biological redox of sulfur, which can either release or immobilize arsenic. [114, 119] Abiotic sulfide, a strong reductant under sulfate-reduced conditions, plays a critical role in arsenic solubility by forming pyrite (FeS_2), realgar (AsS), orpiment (As_2S_3), [120] arsenopyrite (FeAsS), [121] or by reducing As(V). [122]

In addition to abiotic factors, sulfate-reducing bacteria (SRB) also cause dramatic changes in Fe, sulfide and arsenic species by generating hydrogen sulfide [123] or elemental sulfur from sulfate, [5] or by localized reprecipitation of released arsenic as As_2S_3 [124] or FeAsS, which have low solubility. [125] Sulfur-oxidizing bacteria have the potential to immobilize arsenic by using free or arsenic-bound sulfur as an electron donor to directly or indirectly transform As(III) and thioarsenates to As(V),[75] or reduce As(V) [126, 127] In brief, transformations involving sulfur have the potential to significantly impact the fate of environmental arsenic.

3.3 The effects of N on arsenic biogeochemical cycling

Nitrate is an ecologically significant oxidant that can support microbial oxidation of As(III) in the absence of oxygen. The first evidence that microbes are capable of linking anoxic As(III) oxidation to denitrification came from a field study in anoxic lake water columns. [4] The absence or presence of nitrate affected the redox state of arsenic. As(III) was present where nitrate was depleted, but As(V) was the dominant species during anoxic nitrate-rich periods. Subsequently, a nitrate-dependent As(III)

oxidation bacterium *A. ehrlichi* strain MLHE-1 was found to be capable of coupling As(III) oxidation with partial denitrification of nitrate to nitrite. [38, 128] Two other anoxic chemolithoautotrophic strains, *Azoarcus* strain DAO1 and *Sinorhizobium* strain DAO10, were able to oxidize As(III) and fix CO_2 via complete denitrification of nitrate to dinitrogen gas. [129] Biological nitrate-dependent As(III) oxidation is widespread in the environment, and potentially plays a significant role in arsenic biogeochemical cycling. [130]

As mentioned above, nitrate also influences the bioavailability and mobility of arsenic indirectly by linking nitrate reduction to Fe(II) oxidation. Previous studies showed tight coupling between N, Fe, and arsenic in paddy soil. [85] Addition of nitrate reduced arsenic uptake by rice probably because i) the nitrate inhibited/reduced Fe(III) reduction leading to less arsenic mobilization and ii) nitrate-dependent Fe(II)-oxidizing bacteria stimulated Fe(II) oxidation, which led to arsenic co-precipitation with Fe(III) minerals in soil. Nitrate strongly affects arsenic cycling under anoxic conditions in nitrate-rich Upper Mystic Lake by microbially catalyzing As(III) to more particle-reactive As(V) and oxidizing Fe(II) to arsenic-sorbing particulate ferric oxides. [4] Microbial nitrate-dependent Fe(II) oxidation in groundwater, [131] freshwater sediments [132] and marine sediments [133] has the potential to contribute to the reduction of arsenic mobility in various ecosystems.

3.4 The effects of organic matter on arsenic biogeochemical cycling

Natural organic matter (NOM) is widely distributed in the environment. NOM consists of heterogeneous mixtures of organic compounds with various structural and functional properties, [134] that influence the fate of arsenic by competitive adsorption and redox reactions, [135, 136, 137] and by formation of arsenic-bearing organic-metal-complexes and mineral colloids. [138, 139] NOM molecules have

366 combinations of carboxylic, amino, sulfhydryl, hydroxyl, esteric, phenolic, nitroso,
367 and other functional groups. [140] They are considered to be an efficient geochemical
368 trap for arsenic both under oxic and reducing conditions. Whereas As(V) is
369 immobilized by binding to protonated amino groups of NOM [3] or a nucleophile
370 substitution reaction between As(V) and phenolic OH groups of NOM, [141] As(III) is
371 associated with NOM via phenolic OH or carboxyl groups of NOM by H-bonding,
372 hydrophobic As(III)-NOM interactions, [141] or via ternary As(III)-Fe(III)-NOM
373 complexes that form bridges between Fe(III), arsenic oxyanions and the functional
374 entities of NOM. [142] In contrast, under sulfate-reducing conditions, the formation of a
375 trigonal-pyramidal complex between As(III) and sulfhydryl groups of NOM could
376 potentially be a sequestration mechanism for arsenic. [143]

377 Organic matter could also change the fate of arsenic by influencing microbial
378 communities or activities. [144, 145, 146] NOM is thought to drive the reductive dissolution
379 of Fe(III) (oxyhydr)oxides, thereby causing redox transformations of As(V) to As(III)
380 and facilitate arsenic release. [147] In addition to the reductive dissolution of
381 arsenic-bearing Fe(III) (oxyhydr)oxide phases, studies have revealed that addition of
382 organic matter to paddy soil significantly increased arsenic methylation and
383 volatilization. [148] Application of rice straw into soil increased arsenic accumulation in
384 rice by influencing microbial processes involved in arsenic redox. [149]

385 Humic acids, which are forms of NOM, can reduce As(V). [150, 151, 152] Small
386 organic molecules, including lactate, pyruvate, fumarate, malate, succinate, butyrate,
387 citrate, acetate, glycerol, ethanol, and formate, can be used an electron donor by
388 DARPs for As(V) reduction. [153, 154, 155, 156] *Desulfosporosinus* sp. Y5 has been found
389 to couple even complex aromatic substrates such as phenol, syringic acid, benzoate,
390 ferulic acid, and toluene, to As(V) reduction. [157] In fact, some DARPs can also respire

391 sulfate, selenite, Fe(III), thiosulfate, nitrate, or nitrite. [153, 154] This diversity of electron
392 donors and acceptors may be of benefit to microbes grown in environments where Fe,
393 N, S, or C coexist with arsenic.

394 Apart from iron oxides and organic materials, silicon significantly decreased
395 As(III) concentration, but increased the concentration of DMAs in both the vegetative
396 and reproductive tissues of rice. [158] In brief, in addition to biological transformations,
397 other inorganic elements, compounds/minerals and physico-chemical properties
398 interact with arsenic in the environment. Coupling of arsenic with other elements
399 makes it necessary to consider genes involved in cycling of other elements, such as Fe,
400 N, S, and C when studying the arsenic biogeochemical cycling. [2, 87, 88]

401 Known genes involved in arsenic biotransformation are readily determined in
402 pure cultures. However, in the field, these genes cannot easily be quantified in
403 bacterial communities with a limited number of primers, even though As(V)-reducing
404 and As(III)-oxidizing bacteria are widely distributed in the environment. [159, 160, 161] In
405 addition, the speciation, fate and biogeochemical transformation processes of arsenic
406 in the environment are much more complex than under laboratory conditions. [162, 163] It
407 is therefore necessary to apply more systematic and more comprehensive approaches
408 such as metagenomics, metatranscriptomics, metaproteomics, and metabolomics to
409 understand interactions between environmental microbes. These approaches will take
410 into account local geochemical surroundings and neighboring organisms by analyzing
411 DNA, RNA, proteins, and metabolites extracted directly from environmental samples.

412

413 **4 Understanding biogeochemical arsenic cycling by application of "omics"**
414 **methods and integrated modeling**

415 **4.1 Metagenomics**

Metagenomics provides an inestimable window into the microbial world by characterizing microorganisms involved in difficult-to-elucidate but important biochemical pathways, as the overwhelming majority of microbes in the environment cannot be cultured in the laboratory. From a metagenomic library two arsenic-resistant bacteria and one novel As(V) resistance gene (*arsN*), which encodes a protein similar to acetyltransferases, were identified. [22] Xiao et al applied metagenomic techniques to analyze genes associated with arsenic transformation. They analyzed five low-arsenic paddy soils using high-throughput sequencing and constructed a protein database of arsenic metabolizing genes. Their analysis shows that arsenic metabolism genes are ubiquitous and abundant, even in low-arsenic environments. [164] Metagenomics was also used to unravel the correlations between the microbes and arsenic transformation in different niches. [165, 166, 167, 168, 169] Although metagenomics provides taxonomic and functional profiles of a microbial community, it does not demonstrate the levels of expression of the genes nor their physiological activity. [170] Therefore, metatranscriptomics is needed to delineate the active functional genes and communities.

4.2 Metatranscriptomics

Metatranscriptomics offers novel insights into the expression of functional genes and microbial activities of complex microbial communities at a specific moment or under specific environmental conditions by sequencing the total mRNAs extracted from natural microbial communities. Functional metatranscriptomics has potential for isolation and characterization of novel genes involved in heavy metal transformation. [171] Metatranscriptomics enhances our understanding of microbial responses to their environment [172] and the functional profile of a microbial community. [173] Recently, a transcriptomics meta-analysis was used to unravel the effect of As(III) on the

symbiotic interaction between the model legume *Medicago truncatula* and its symbiont *Ensifer* (syn. *Sinorbizobium*) *medicae* MA11. [174] This study identified the adaptive responses of the bacterial symbiont to arsenic exposure. This metatranscriptomic approach will be useful to study how microbes regulate their genes to adapt to the changes in environmental conditions, particularly arsenic concentrations. [175] The correlation between mRNA and protein inventories in environmental microbial communities is low when environmental conditions change rapidly. mRNA inventories respond rapidly and sensitively to the shift, while changes in protein inventories are slow. [176] As a consequence, microbial metaproteomics and metabolomics had to be used to identify the repertoire of proteins and small molecular metabolites that microbes use to adapt to complex and dynamic environments. In this way, the metabolic activities of a microbial community in specific environments at the moment of sampling could be elucidated. [170]

4.3 Metaproteomics and metabolomics

Metaproteomics and metabolomics are the comprehensive methods by which proteins produced by microbes and metabolites released by microorganisms into the environment are characterized and quantified using a combination of liquid or gas chromatography-based separations and mass spectrometry-based identification techniques. [177, 178] The study of microbial proteome and metabolome can provide valuable information about the function of microbial communities and the interactions of the microbial communities with the environment. [179, 180] When the diversity of arsenic-adapted prokaryotic communities in mildly arsenic-contaminated sediments was analyzed using meta-proteomic and 16S rRNA amplification, the results indicated that the data sizes provided by metaproteomics are less than those provided by metagenomics and metatranscriptomics. [181] So far, metabolomics has been used

mainly to analyze low molecular mass metabolites within a tissue, biofluid, a cell or cell compartment of an organism including plants, animals, bacteria, and humans exposed to an environmental stressor. [180] High-throughput metabolomics has been applied to the analysis of metabolites in the liver of mice when co-exposed to high fat and cholesterol diets and arsenic-contaminated drinking water. [182] However, application of metaproteomic and metabolomic techniques to real environment is limited due to difficulties with amplification and the low quantities of extractable proteins and metabolites because of the interferences with many components present in complex environmental systems, such as soil. [180, 183]

4.4 Integrating meta-omic techniques

From the above, meta-omics are key techniques in elucidating the dynamic and complex interactions between microbial communities and the environment. Integrating multiple meta-omic datasets will provide a complete exhibition from genes to biogeochemical cycles. Metatranscriptomic and metagenomic techniques were combined to detect large numbers of novel genes from complex marine microbial communities. [173] Datasets of meta-genomics were integrated with metabolomics to reveal how a microbial community interacts with the environment and responds to environmental parameters. [184] Taken together, meta-omic technologies offer an unprecedented opportunity to elucidate the functions of microbes that are not readily cultured under normal laboratory conditions in biogeochemical cycles.

Recently, "Arsenomics" was termed as an approach to focusing on the analysis of alterations in transcriptome, proteome and metabolome occurring in microbes exposed to arsenic. [185, 186] With the application of meta-omics to environmental

490 science, we believe that Arsenomics will evolve to include the analysis of
491 metagenomic, metatranscriptomic, metaproteomic, and metabolomic changes in
492 microbial communities from the real environment where they are exposed to arsenic.

493 **4.5 Integrating environmental meta-omics into biogeochemical models for**
494 **arsenic**

495 Microbes are ubiquitous in diverse environmental niches including soil, oceans,
496 sediments, freshwater environments, and inside the body of animals or plants, and
497 exert great influence on biogeochemical cycles in these habitats. [187] For example,
498 microbes involved in arsenic transformation are ubiquitously distributed in paddy
499 soils, resulting in various concentrations and percentage of inorganic and methylated
500 arsenic species among different rice plants. [188, 189, 190, 191] Since higher plants appear
501 not to methylate arsenic, [192] microbial methylation is probably the primary source of
502 methylated arsenic in plants, which occurs in soil prior to plant uptake.

503 *In situ* measurements or prediction of arsenic transformations contribute to
504 analysis of the dynamics of arsenic and prediction of arsenic bioavailability. *In situ*
505 measurements of As(V) reduction in Mono Lake, California (dissolved inorganic
506 arsenic ~ 200 μM), made with radiotracers (^{73}As and ^{35}S) of mass balance
507 considerations, revealed that As(V) reduction occurred in the monimolimnion waters
508 with the highest rates between 18 and 19 meters (rate, ~ 5.9 μM/day) and sulfate
509 reduction rates increased with depth at depths of 21 meters and below with the highest
510 rates at 28 meters (rate, ~ 2.3 μM/day). [193] The radioisotope method was further
511 employed to examine the As(V) and sulfate reduction processes in sediments of two
512 arsenic-rich soda lakes, Mono Lake (moderately salt, ~ 90 g/L) and Searles Lake
513 (saturated salt, ~ 340 g/L). [194] The rate constant [k] of As(V) reduction was 0.103 to
514 0.04 h^{-1} in Mono Lake and 0.012 to 0.002 h^{-1} in Searles Lake, and sulfate reduction

was only detected in Mono Lake ($k=7.6\times10^{4}$ to 3.2×10^{-6} h^{-1}). Denatured gradient gel electrophoresis (DGGE) of 16S rRNA genes amplified from Mono Lake and Searles Lake sediment DNA indicated that microbial communities from two sediments were distinct from each other. More *arrA* gene signal was found in Mono Lake than in Searles Lake, where higher As(V) reduction activity was observed, due to PCR biases, the presence of novel *arrA* genes, or higher expression of low-abundance *arrA* genes. [194]

Characterization of genes involved in arsenic biotransformation and application of multiple meta-omics in environment analysis will lead to insights into the microbial world, as limited information on functional genes cannot predict the status of arsenic in the environment. The information of field/*in situ* characterization of functional genes/functional microbial communities and biogeochemical fluxes should be integrated into biogeochemical models to complete the transition from lab to the field, from biochemistry to biogeochemistry, and from genes-genomics to microbial communities. This integration will help to predict the dynamics of arsenic in the environment, and to improve the effectiveness of mitigation technologies. Many strategies have been developed to model low complexity environments. For example, recent work integrating environmental genomics and qPCR in biogeochemical models explored the nexus between microbial community and geochemisty in the Arabian Sea oxygen minimum zone. [195] Metabolic processes coupling C, N, and S transformations in the Saanich Inlet oxygen-starved zone were integrated using a biogeochemical model that integrates multi-omic information and geochemistry. [196] These studies indicated that such integrated modeling approaches can provide a novel insight into microbial metabolic networks in water bodies, and pave a road for prediction of elemental cycling.

5 Perspectives

In summary, as arsenic biotransformations are catalyzed by a suite of enzymes from diverse environmental organisms, and these are coupled to biogeochemical cycles of other elements such as Fe, S, and N. As more and more genomes are being sequenced, more genes directly or indirectly involved in arsenic metabolism will be discovered and characterized. With the development of new technologies, we anticipate rapid advances in analytical chemistry, microbiology and genomics that will improve our understanding of how microbial metabolic pathways contribute to and govern complex environmental processes. In the future, integrating meta-omic datasets into biogeochemical models will improve the ability of prediction and offer a deeper insight into arsenic biogeochemical processes in diverse niches.

In future studies, it will be necessary to analyze the interaction between organisms and the environment using additional meta-omics approaches at different spatio-temporal scales. Geochemical analyses in combination with genetic analyses will provide insights into the specific roles of the complex biochemical pathways in the global arsenic biogeochemical cycle. More importantly, integrating modeling approaches linking arsenic biogeochemical cycle with meta-omics data should be developed to predict the dynamic of arsenic species in water, sediments and soils and provide our society and authorities with the tools necessary for limiting arsenic pollution, improving remediation and providing safe drinking water and food.

Acknowledgments

Our research was supported by National Natural Science Foundation of China (41571130063), Program of Chinese Academy of Sciences (XDB15020402), National Natural Science Foundation of China (21507125), and International Science & technology Cooperation Program of China (2011DFB91710). BPR was supported by NIH grants GM55425 and ES023779.

Reference

(1) Bashkin V N. *Modern biogeochemistry*; Springer Science & Business Media, 2002.

(2) Oremland, R. S.; Stolz, J. F. The ecology of arsenic. *Science (80).* **2003**, *300*, 939–944 DOI: 10.1126/science.1081903.

(3) Thanabalasingam, P.; Pickering, W. F. Arsenic sorption by humic acids. *Environ. Pollut. (Series B)* **1986**, *12* (3), 233–246 DOI: 10.1016/0143-148X(86)90012-1.

(4) Senn, D. B.; Hemond, H. F. Nitrate controls on iron and arsenic in an urban lake. *Science (80).* **2002**, *296*, 2373–2376 DOI: 10.1126/science.1072402.

(5) Saalfield, S.; Bostick, B. Changes in iron, sulfur, and arsenic speciation associated with bacterial sulfate reduction in ferrihydrate-rich systems. *Environ. Sci. Technol.* **2009**, *43*, 8787–8793 DOI: 10.1021/es901651k.

(6) Finzi, A. C.; Cole, J. J.; Doney, S. C.; Holland, E. A.; Jackson, R. B. Research frontiers in the analysis of coupled biogeochemical cycles. *Front Ecol Env.* **2011**, *9* (1), 74–80 DOI: 10.1890/100137.

(7) Mukhopadhyay, R.; Rosen, B. P.; Phung, L. T.; Silver, S. Microbial arsenic: from geocycles to genes and enzymes. *FEMS Microbiol Rev* **2002**, *26*, 311–325 DOI: 10.1111/j.1574-6976.2002.tb00617.x.

(8) Zhu, Y.-G.; Yoshinaga, M.; Zhao, F.-J.; Rosen, B. P. Earth Abides Arsenic Biotransformations. *Annu. Rev. Earth Planet. Sci.* **2014**, *42* (1), 443–467 DOI: 10.1146/annurev-earth-060313-054942.

(9) Sardiwal, S.; Santini, J. M.; Osborne, T. H.; Djordjevic, S. Characterization of a two-component signal transduction system that controls arsenite oxidation in the chemolithoautotroph NT-26. *FEMS Microbiol. Lett.* **2010**, *313* (1), 20–28 DOI: 10.1111/j.1574-6968.2010.02121.x.

(10) Richey, C.; Chovanec, P.; Hoeft, S. E.; Oremland, R. S.; Basu, P.; Stolz, J. F. Respiratory arsenate reductase as a bidirectional enzyme. *Biochem. Biophys. Res. Commun.* **2009**, *382* (2), 298–302 DOI: 10.1016/j.bbrc.2009.03.045.

(11) Zargar, K.; Conrad, A.; Bernick, D. L.; Lowe, T. M.; Stolc, V.; Hoeft, S.; Oremland, R. S.; Stolz, J.; Saltikov, C. W. ArxA, a new clade of arsenite oxidase within the DMSO reductase family of molybdenum oxidoreductases. *Environ. Microbiol.* **2012**, *14* (7), 1635–1645 DOI:

10.1111/j.1462-2920.2012.02722.x.

(12) Liu, G.; Liu, M.; Kim, E. H.; Maaty, W. S.; Bothner, B.; Lei, B.; Rensing, C.; Wang, G.; McDermott, T. R. A periplasmic arsenite-binding protein involved in regulating arsenite oxidation. *Environ. Microbiol.* **2012**, *14* (7), 1624–1634 DOI: 10.1111/j.1462-2920.2011.02672.x.

(13) Chen, J.; Bhattacharjee, H.; Rosen, B. P. ArsH is an organoarsenical oxidase that confers resistance to trivalent forms of the herbicide monosodium methylarsenate and the poultry growth promoter roxarsone. *Mol. Microbiol.* **2015**, *96* (5), 1042–1052 DOI: 10.1111/mmi.12988.

(14) Xue, X. M.; Yan, Y.; Xu, H. J.; Wang, N.; Zhang, X.; Ye, J. ArsH from *Synechocystis* sp. PCC 6803 reduces chromate and ferric iron. *FEMS Microbiol. Lett.* **2014**, *356* (1), 105–112 DOI: 10.1111/1574-6968.12481.

(15) Afkar, E.; Lisak, J.; Saltikov, C.; Basu, P.; Oremland, R. S.; Stolz, J. F. The respiratory arsenate reductase from *Bacillus selenitireducens* strain MLS10. *FEMS Microbiol. Lett.* **2003**, *226* (1), 107–112 DOI: 10.1016/S0378-1097(03)00609-8.

(16) Krafft, T.; Macy, J. M. Purification and characterization of the respiratory arsenate reductase of *Chrysiogenes arsenatis*. *Eur. J. Biochem.* **1998**, *255* (3), 647–653 DOI: 10.1046/j.1432-1327.1998.2550647.x.

(17) Van Lis, R.; Nitschke, W.; Duval, S.; Schoepp-Cothenet, B. Arsenics as bioenergetic substrates. *Biochim. Biophys. Acta - Bioenerg.* **2013**, *1827* (2), 176–188 DOI: 10.1016/j.bbabio.2012.08.007.

(18) Mukhopadhyay, R.; Rosen, B. P. Arsenate reductases in prokaryotes and eukaryotes. *Environ. Health Perspect.* **2002**, *110*, 745–748 DOI: 10.1289/ehp.02110s5745.

(19) Wysocki, R.; Goffeau, A. Isolation of three contiguous genes, ACR1, ACR2 and ACR3, involved in resistance to arsenic compounds in the yeast *Saccharomyces cerevisiae*. *Yeast* **1997**, *13*, 819–828 DOI: 10.1002/(SICI)1097-0061(199707)13.

(20) David J. Thomas; Barry P. Rosen. Arsenic Methyltransferase. In *Encyclopedia of Metalloproteins*; Uversky, V. N., Kretsinger, R. H., Permyakov, Ei. E. a., Eds.; Science+Business Media: New York, 2013; pp 138–143 DOI: 10.1007/978-1-4614-1533-6.

(21) Chrysostomou, C.; Quandt, E. M.; Marshall, N. M.; Stone, E.; Georgiou, G. An alternate pathway of arsenate resistance in *E. coli* mediated by the glutathione S-transferase GstB. *ACS Chem. Biol.* **2015**, *10* (3), 875–882 DOI: 10.1021/cb500755j.

(22) Chauhan, N. S.; Ranjan, R.; Purohit, H. J.; Kalia, V. C.; Sharma, R. Identifcation of genes conferring arsenic resistance to *Escherichia coli* from an effluent treatment plant sludge metagenomic library. *FEMS Microbiol Ecol* **2009**, *67*, 130–139 DOI: 10.1111/j.1574-6941.2008.00613.x.

(23) Qin, J.; Rosen, B. P.; Zhang, Y.; Wang, G.; Franke, S.; Rensing, C. Arsenic detoxification and evolution of trimethylarsine gas by a microbial arsenite S-adenosylmethionine methyltransferase. *Proc. Natl. Acad. Sci.* **2006**, *103* (7), 2075–2080 DOI: 10.1073/pnas.0506836103.

(24) Yoshinaga, M.; Rosen, B. P. A C · As lyase for degradation of environmental organoarsenical herbicides and animal husbandry growth promoters. *Proc. Natl Acad. Sci.* **2014**, *111* (21), 7701–7706 DOI: 10.1073/pnas.1403057111.

(25) Louis S. Tisa; Rose, B. P. Molecular characterization of an anion pump-The ArsB protein is the membrane anchor for the ArsA protein. *J. Biol. Chem.* **1990**, *265*, 190–194.

(26) Lin, Y.; Walmsley, A. R.; Rosen, B. P. An arsenic metallochaperone for an arsenic detoxification pump. *Proc. Natl Acad. Sci.* **2006**, *103* (42), 15617–15622 DOI: 10.1073/pnas.0603974103.

(27) Yang, H.; Cheng, J.; Finan, T. M.; Rosen, B. P. Novel pathway for arsenic detoxification in the legume symbiont *Sinorhizobium meliloti*. *J. Bacteriol.* **2005**, *187* (20), 6991–6997 DOI: 10.1128/JB.187.20.6991.

(28) Ghosh, M.; Shen, J.; Rosen, B. P. Pathways of As (III) detoxification in *Saccharomyces cerevisiae*. *Proc. Natl Acad. Sci.* **1999**, *96*, 5001–5006 DOI: 10.1073/pnas.96.9.5001.

(29) Chen, J.; Yoshinaga, M.; Garbinski, L. D.; Rosen, B. P. Synergistic interaction of glyceraldehydes-3-phosphate dehydrogenase and ArsJ, a novel organoarsenical efflux permease, confers arsenate resistance. *Mol. Microbiol.* **2016**, *100* (6), 945–953 DOI: 10.1111/mmi.13371.

(30) Chen, J.; Madegowda, M.; Bhattacharjee, H.; Rosen, B. P. ArsP: A methylarsenite efflux permease. *Mol. Microbiol.* **2015**, *98* (4), 625–635 DOI: 10.1111/mmi.13145.

(31) Grondin, K.; Haimeur, A.; Mukhopadhyay, R.; Rosen, B. P.; Ouellette, M. Co-amplification of the gamma-glutamylcysteine synthetase gene gsh1 and of the ABC transporter gene pgpA in arsenite-resistant *Leishmania tarentolae*. *EMBO J.* **1997**, *16* (11), 3057–3065 DOI: 10.1093/emboj/16.11.3057.

(32) Wang, L.; Chen, S.; Xiao, X.; Huang, X.; You, D.; Zhou, X.; Deng, Z. arsRBOCT arsenic resistance system encoded by linear plasmid pHZ227 in *Streptomyces* sp. strain FR-008. *Appl Env. Microbiol* **2006**, *72* (5), 3738–3742 DOI: 10.1128/AEM.72.5.3738.

(33) Oremland, R. S.; Saltikov, C. W.; Wolfe-Simon, F.; Stolz, J. F. Arsenic in the Evolution of Earth and Extraterrestrial Ecosystems. *Geomicrobiol. J.* **2009**, *26* (V), 522–536 DOI: 10.1080/01490450903102525.

(34) McCann, S. H.; Boren, A.; Hernandez-maldonado, J.; Stoneburner, B.; Saltikov, C. W.; Stolz, J. F.; Oremland, R. S. Arsenite as an Electron Donor for Anoxygenic Photosynthesis: Description of Three Strains of *Ectothiorhodospira* from Mono Lake, California and Big Soda Lake , Nevada. *Life* **2016**, *7* (1), 1–14 DOI: 10.3390/life7010001.

(35) Hernandez-maldonado, J.; Sanchez-sedillo, B.; Stoneburner, B.; Boren, A.; Miller, L.; Mccann, S.; Rosen, M.; Oremland, R. S.; Saltikov, C. W. The genetic basis of anoxygenic photosynthetic arsenite oxidation. *Env. Microbiol* **2017**, *19*, 130–141 DOI: 10.1111/1462-2920.13509.

(36) Kulp, T. R.; Hoeft, S. E.; Asao, M.; Madigan, M. T.; Hollibaugh, J. T.; Fisher, J. C.; Stolz, J. F.; Culbertson, C. W.; Miller, L. G.; Oremland, R. S. Arsenic(III) fuels anoxygenic photosynthesis in hot spring biofilms from Mono Lake, California. *Science* **2008**, *321* (5891), 967–970 DOI:

10.1126/science.1160799.
(37) Hoeft, S. E.; Kulp, T. R.; Han, S.; Lanoil, B.; Oremland, R. S. Coupled arsenotrophy in a hot spring photosynthetic biofilm at mono lake, California. *Appl. Environ. Microbiol.* **2010**, *76* (14), 4633–4639 DOI: 10.1128/AEM.00545-10.
(38) Oremland, R. S.; Hoeft, S. E.; Santini, J. M.; Bano, N.; Hollibaugh, R. a; Hollibaugh, J. T. Anaerobic Oxidation of Arsenite in Mono Lake Water and by a Facultative, Arsenite-Oxidizing Chemoautotroph, Strain MLHE-1. *Appl Env. Microbiol* **2002**, *68* (10), 4795–4802 DOI: 10.1128/AEM.68.10.4795.
(39) Kulp, T. R. Arsenic and primordial life. *Nat. Geosci.* **2014**, *7* (11), 785–786 DOI: 10.1038/ngeo2275.
(40) Stolz, J. F.; Basu, P.; Oremland, R. S. Microbial Arsenic Metabolism: New Twists on an Old Poison. *Microbe* **2010**, *5* (2), 53–59 DOI: 10.1128/microbe.5.53.1.
(41) Branco, R.; Francisco, R.; Chung, A. P.; Morais, P. V. Identification of an aox system that requires cytochrome c in the highly arsenic-resistant bacterium *Ochrobactrum tritici* SCII24. *Appl. Environ. Microbiol.* **2009**, *75* (15), 5141–5147 DOI: 10.1128/AEM.02798-08.
(42) Zargar, K.; Hoeft, S.; Oremland, R.; Saltikov, C. W. Identification of a novel arsenite oxidase gene, arxA, in the haloalkaliphilic, arsenite-oxidizing bacterium *Alkalilimnicola ehrlichii* strain MLHE-1. *J. Bacteriol.* **2010**, *192* (14), 3755–3762 DOI: 10.1128/JB.00244-10.
(43) Hery, M.; Gault, A. G.; Rowland, H. A. L.; Lear, G.; Polya, D. A.; Lloyd, J. R. Molecular and cultivation-dependent analysis of metal-reducing bacteria implicated in arsenic mobilisation in south-east asian aquifers. *Appl. Geochemistry* **2008**, *23* (11), 3215–3223 DOI: 10.1016/j.apgeochem.2008.07.003.
(44) Pederick, R. L.; Gault, A. G.; Charnock, J. M.; Polya, D. A.; Lloyd, J. R. Probing the biogeochemistry of arsenic: Response of two contrasting aquifer sediments from Cambodia to stimulation by arsenate and ferric iron. *J. Environ. Sci. Heal. Part A* **2007**, *42*, 1763–1774 DOI: 10.1080/10934520701564269.
(45) Yan, Y.; Ding, K.; Yu, X.; Ye, J.; Xue, X. Ability of Periplasmic Phosphate Binding Proteins from *Synechocystis* sp . PCC 6803 to Discriminate Phosphate Against Arsenate. *Water. Air. Soil Pollut.* **2017**, *228*, 148 DOI: 10.1007/s11270-017-3334-4.
(46) Bhattacharjee, H.; Sheng, J.; Ajees, A. A.; Mukhopadhyay, R.; Rosen, B. P. Adventitious arsenate reductase activity of the catalytic domain of the human Cdc25B and Cdc25C phosphatases. *Biochemistry* **2010**, *49* (4), 802–809 DOI: 10.1021/bi9019127.
(47) Zhou, R.; Royyuru, A.; Athma, P.; Suits, F.; Silverman, B. D. Magnitude and spatial orientation of the hydrophobic moments of multi-domain proteins. *Int. J. Bioinform. Res. Appl.* **2006**, *2* (2), 161–176 DOI: 10.1016/j.molbiopara.2006.03.009.
(48) Wang, P.; Sun, G.; Jia, Y.; Meharg, A. A.; Zhu, Y. A review on completing arsenic biogeochemical cycle: Microbial volatilization of arsines in environment. *J. Environ. Sci.* **2014**, *26*, 371–381 DOI: 10.1016/S1001-0742(13)60432-5.
(49) Yin, X.-X.; Chen, J.; Qin, J.; Sun, G.-X.; Rosen, B. P.; Zhu, Y.-G. Biotransformation and volatilization of arsenic by three photosynthetic cyanobacteria. *Plant Physiol.* **2011**, *156* (3),

719 1631–1638 DOI: 10.1104/pp.111.178947.
720 (50) Yin, X. X.; Zhang, Y. Y.; Yang, J.; Zhu, Y. G. Rapid biotransformation of arsenic by a model
721 protozoan *Tetrahymena thermophila*. *Environ. Pollut.* **2011**, *159* (4), 837–840 DOI:
722 10.1016/j.envpol.2010.12.033.
723 (51) Ye, J.; Rensing, C.; Rosen, B. P.; Zhu, Y. G. Arsenic biomethylation by photosynthetic
724 organisms. *Trends Plant Sci.* **2012**, *17* (3), 155–162 DOI: 10.1016/j.tplants.2011.12.003.
725 (52) Xue, X.-M.; Ye, J.; Raber, G.; Francesconi, K. A.; Li, G.; Gao, H.; Yan, Y.; Rensing, C.; Zhu,
726 Y.-G. Arsenic Methyltransferase is Involved in Arsenosugar Biosynthesis by Providing DMA.
727 *Environ. Sci. Technol.* **2017**, *51* (3), 1224–1230 DOI: 10.1021/acs.est.6b04952.
728 (53) Stolz, J. F.; Perera, E.; Kilonzo, B.; Kail, B.; Crable, B.; Fisher, E.; Ranganathan, M.; Wormer,
729 L.; Basu, P. Biotransformation of 3-nitro-4-hydroxybenzene arsonic acid (Roxarsone) and
730 release of inorganic arsenic by clostridium species. *Environ. Sci. Technol.* **2007**, *41* (3),
731 818–823 DOI: 10.1021/es061802i.
732 (54) Lehr, C. R.; Polishchuk, E.; Radoja, U.; Cullen, W. R. Demethylation of methylarsenic species
733 by *Mycobacterium neoaurum*. *Appl. Organomet. Chem.* **2003**, *17* (11), 831–834 DOI:
734 10.1002/aoc.544.
735 (55) Yoshinaga, M.; Cai, Y.; Rosen, B. P. Demethylation of methylarsonic acid by a microbial
736 community. *Environ. Microbiol.* **2011**, *13* (5), 1205–1215 DOI:
737 10.1111/j.1462-2920.2010.02420.x.
738 (56) Yan, Y.; Ye, J.; Xue, X.-M.; Zhu, Y.-G. Arsenic Demethylation by a C · As Lyase in
739 Cyanobacterium *Nostoc* sp. PCC 7120. *Environ. Sci. Technol.* **2015**, *49*, 14350–14358 DOI:
740 10.1021/acs.est.5b03357.
741 (57) Edmonds, J. S.; Francesconi, K. A. Organoarsenic compounds in the marine environment. In
742 *Organometallic compounds in the environment*; Craig, P. J., Ed.; Wiley: Chichester, 2013; pp
743 195–222 DOI: 10.1017/CBO9781107415324.004
744 (58) Edmonds, J. S.; Francesconi, K. A. Arseno-sugars from brown kelp (*Ecklonia radiata*) as
745 intermediates in cycling of arsenic in a marine ecosystem. *Nature* **1981**, *289*, 602–604 DOI:
746 10.1038/289602a0
747 (59) Morita, M.; Shibata, Y. Isolation and identification of arseno-lipid from a brown alga, *Undaria*
748 *pinnatifida* (Wakame). *Chemosphere* **1988**, *17* (6), 1147–1152 DOI:
749 10.1016/0045-6535(88)90180-4.
750 (60) Garcıı´a-Salgadoı, S.; Raber, G.; Raml, R.; Magnes, C.; Francesconi, K. A. Arsenosugar
751 phospholipids and arsenic hydrocarbons in two species of brown macroalgae. *Environ. Chem.*
752 **2012**, *9*, 63–66 DOI: 10.1071/EN11164.
753 (61) Xue, X.-M.; Raber, G.; Foster, S.; Chen, S.-C.; Francesconi, K. A.; Zhu, Y.-G. Biosynthesis of
754 arsenolipids by the cyanobacterium *Synechocystis* sp. PCC 6803. *Environ. Chem.* **2014**, *11* (5),
755 506–513 DOI: 10.1071/EN14069.
756 (62) Rumpler, A.; Edmonds, J. S.; Katsu, M.; Jensen, K. B.; Goessler, W.; Raber, G.;
757 Gunnlaugsdottir, H.; Francesconi, K. A. Arsenic-containing long-chain fatty acids in cod-liver
758 oil: A result of biosynthetic infidelity? *Angew. Chemie - Int. Ed.* **2008**, *47* (14), 2665–2667

DOI: 10.1002/anie.200705405.

(63) Raab, A.; Newcombe, C.; Pitton, D.; Ebel, R.; Feldmann, J. Comprehensive analysis of lipophilic arsenic species in a brown alga (*Saccharina latissima*). *Anal. Chem.* **2013**, *85* (5), 2817–2824 DOI: 10.1021/ac303340t.

(64) Taleshi, M. S.; Raber, G.; Edmonds, J. S.; Jensen, K. B.; Francesconi, K. A. Arsenolipids in oil from blue whiting *Micromesistius poutassou*--evidence for arsenic-containing esters. *Sci. Rep.* **2014**, *4*, 7492 DOI: 10.1038/srep07492.

(65) Taleshi, M. S.; Edmonds, J. S.; Goessler, W.; Ruiz-Chancho, M. J.; Raber, G.; Jensen, K. B.; Francesconi, K. A. Arsenic-containing lipids are natural constituents of sashimi tuna. *Environ. Sci. Technol.* **2010**, *44* (4), 1478–1483 DOI: 10.1021/es9030358.

(66) Taleshi, M. S.; Jensen, K. B.; Raber, G.; Edmonds, J. S.; Gunnlaugsdottir, H.; Francesconi, K. A. Arsenic-containing hydrocarbons: natural compounds in oil from the fish capelin, *Mallotus villosus*. *Chem. Commun.* **2008**, *39* (39), 4706–4707 DOI: 10.1039/b808049f.

(67) Amayo, K. O.; Raab, A.; Krupp, E. M.; Gunnlaugsdottir, H.; Feldmann, J. Novel identification of arsenolipids using chemical derivatizations in conjunction with RP-HPLC-ICPMS/ESMS. *Anal. Chem.* **2013**, *85* (19), 9321–9327 DOI: 10.1021/ac4020935.

(68) Viczek, S. A.; Jensen, K. B.; Francesconi, K. A. Arsenic-containing Phosphatidylcholines: a New Group of Arsenolipids Discovered in Herring Caviar. *Angew. Chemie Int. Ed.* **2016**, *55*, 5259–5262 DOI: 10.1002/anie.201512031.

(69) Van Mooy, B. a S.; Fredricks, H. F.; Pedler, B. E.; Dyhrman, S. T.; Karl, D. M.; Koblízek, M.; Lomas, M. W.; Mincer, T. J.; Moore, L. R.; Moutin, T.; et al. Phytoplankton in the ocean use non-phosphorus lipids in response to phosphorus scarcity. *Nature* **2009**, *458*, 69–72 DOI: 10.1038/nature07659.

(70) Petursdottir, A. H.; Fletcher, K.; Gunnlaugsdottir, H.; Krupp, E.; Kupper, F. C.; Feldmann, J. Environmental effects on arsenosugars and arsenolipids in Ectocarpus (Phaeophyta). *Environ. Chem.* **2016**, *13* (1), 21–33 DOI: 10.1071/EN14229.

(71) Meyer, S.; Matissek, M.; Müller, S. M.; Taleshi, M. S.; Ebert, F.; Francesconi, K. A.; Schwerdtle, T. *In vitro* toxicological characterisation of three arsenic-containing hydrocarbons. *Metallomics* **2014**, *6*, 1023–1033 DOI: 10.1039/c4mt00061g.

(72) Schmeisser, E.; Goessler, W.; Francesconi, K. A. Human metabolism of arsenolipids present in cod liver. *Anal. Bioanal. Chem.* **2006**, *385* (2), 367–376 DOI: 10.1007/s00216-006-0401-x.

(73) Taylor, V.; Goodale, B.; Raab, A.; Schwerdtle, T.; Reimer, K.; Conklin, S.; Karagas, M. R.; Francesconi, K. A. Human exposure to organic arsenic species from seafood. *Sci. Total Environ.* **2017**, *580*, 266–282 DOI: 10.1016/j.scitotenv.2016.12.113.

(74) Planer-friedrich, B.; Suess, E.; Scheinost, A. C.; Wallschla, D. Arsenic speciation in sulfidic waters: Reconciling contradictory spectroscopic and chromatographic evidence. *Anal. Chem.* **2010**, *82* (24), 10228–10235 DOI: 10.1021/ac1024717.

(75) Fisher, J. C.; Wallschlager, D.; Planer-Friendrich B., H. J. T. A new role for sulfur in arsenic cycling. *Environ. Sci. Technol.* **2008**, *42* (1), 81–85 DOI: 10.1021/es0713936.

(76) Planer-Friedrich, B.; London, J.; Mccleskey, R. B.; Nordstrom, D. K.; Wallschlager, D.

Thioarsenates in geothermal waters of yellowstone National Park: Determination, preservation, and geochemical importance. *Environ. Sci. Technol.* **2007**, *41* (15), 5245–5251 DOI: 10.1021/es070273v.

(77) Taylor, P.; Planer-friedrich, B.; Fisher, J. C.; Hollibaugh, J. T.; Süß, E.; Planer-friedrich, B.; Fisher, J. C.; Hollibaugh, J. T.; Elke, S.; Wallschl, D. Oxidative Transformation of Trithioarsenate Along Alkaline Geothermal Drainages — Abiotic versus Microbially Mediated Processes. *Geomicrobiol J* **2009**, *26*, 339–350 DOI: 10.1080/01490450902755364.

(78) Planer-friedrich, B.; Hartig, C.; Lohmayer, R.; Suess, E.; Mccann, S. H.; Oremland, R. Anaerobic chemolithotrophic growth of the haloalkaliphilic bacterium strain MLMS-1 by disproportionation of monothioarsenate. *Env. Sci Technol* **2015**, *49*, 6554–6563 DOI: 10.1021/acs.est.5b01165.

(79) Härtig, C.; Planer-friedrich, B. Thioarsenate transformation by filamentous microbial mats thriving in an alkaline, sulfidic hot Spring. *Environ. Sci. Technol.* **2012**, *46* (8), 4348–4356 DOI: 10.1021/es204277j.

(80) Hartig, C.; Lohmayer, R.; Kolb, S.; Horn, M. A.; Inskeep, W. P.; Planer-friedrich, B. Chemolithotrophic growth of the aerobic hyperthermophilic bacterium *Thermocrinis ruber* OC 14/7/2 on monothioarsenate and arsenite. *FEMS Microbiol Ecol* **2014**, *90*, 747–760 DOI: 10.1111/1574-6941.12431.

(81) Edwardson, C. F.; Planer-friedrich, B.; Hollibaugh, J. T. Transformation of monothioarsenate by haloalkaliphilic, anoxygenic photosynthetic purple sulfur bacteria. *FEMS Microbiol Ecol* **2014**, *90*, 858–868 DOI: 10.1111/1574-6941.12440.

(82) Tamaki, S.; Frankenberger, W. T. J. Environmental biochemistry of arsenic. In *Reviews of environmental contamination and toxicology*; Ware, G. W., Ed.; Springer New York: United States, 1992; Vol. 124, pp 79–110 DOI: 10.1007/978-1-4612-2864-6_4

(83) Ali, W.; Isayenkov, S. V.; Zhao, F. J.; Maathuis, F. J. M. Arsenite transport in plants. *Cell. Mol. Life Sci.* **2009**, *66* (14), 2329–2339 DOI: 10.1007/s00018-009-0021-7.

(84) Broer, S.; Ji, G.; Broer, A.; Silver, S. Arsenic efflux governed by the arsenic resistance determinant of *Staphylococcus aureus* plasmid pI258. *J. Bacteriol.* **1993**, *175* (11), 3480–3485 DOI: 10.1128/jb.175.11.3480-3485.1993.

(85) Chen, X.-P.; Zhu, Y.-G.; Hong, M.-N.; Kappler, A.; Xu, Y.-X. Effects of different forms of nitrogen fertilizers on arsenic uptake by rice plants. *Environ. Toxicol. Chem.* **2008**, *27* (4), 881–887 DOI: 10.1897/07-368.1.

(86) Handley, K. M.; Mcbeth, J. M.; Charnock, J. M.; Vaughan, D. J.; Wincott, P. L.; Polya, D. A.; Lloyd, J. R. Effect of iron redox transformations on arsenic solid-phase associations in an arsenic-rich, ferruginous hydrothermal sediment. *Geochim. Cosmochim. Acta* **2013**, *102*, 124–142 DOI: 10.1016/j.gca.2012.10.024.

(87) Carlson, H. K.; Clark, I. C.; Blazewicz, S. J.; Iavarone, A. T.; Coates, J. D. Fe(II) oxidation is an innate capability of nitrate-reducing bacteria that involves abiotic and biotic reactions. *J. Bacteriol.* **2013**, *195* (14), 3260–3268 DOI: 10.1128/JB.00058-13.

(88) Bertin, P. N.; Heinrich-Salmeron, A.; Pelletier, E.; Goulhen-Chollet, F.; Arsène-Ploetze, F.;

Gallien, S.; Lauga, B.; Casiot, C.; Calteau, A.; Vallenet, D.; et al. Metabolic diversity among main microorganisms inside an arsenic-rich ecosystem revealed by meta- and proteo-genomics. *ISME J.* **2011**, *5* (11), 1735–1747 DOI: 10.1038/ismej.2011.51.

(89) Melton, E. D.; Swanner, E. D.; Behrens, S.; Schmidt, C.; Kappler, A. The interplay of microbially mediated and abiotic reactions in the biogeochemical Fe cycle. *Nat. Rev. Microbiol.* **2014**, *12* (12), 797–809 DOI: 10.1038/nrmicro3347.

(90) Masscheleyn, P. H.; Delaune, R. D.; Patrick, Jr., W. H. Effect of redox potential and pH on arsenic speciation and solubility in a contaminated soil. *Environ. Sci. Technol.* **1991**, *25* (8), 1414–1419 DOI: 10.1021/es00020a008.

(91) Dixit, S.; Hering, J. G. Comparison of arsenic(V) and arsenic(III) sorption onto iron oxide minerals: Implications for arsenic mobility. *Environ. Sci. Technol.* **2003**, *37* (18), 4182–4189 DOI: 10.1021/es030309t.

(92) Amstaetter, K.; Borch, T.; Larese-Casanova, P.; Kappler, A. Redox transformation of arsenic by Fe(II)-activated goethite (-FeOOH). *Environ. Sci. Technol.* **2010**, *44* (1), 102–108 DOI: 10.1021/es901274s.

(93) Borch, T.; Kretzschmar, R.; Kappler, A.; Van Cappellen, P.; Ginder-Vogel, M.; Voegelin, A.; Campbell, K. Biogeochemical redox processes and their impact on contaminant dynamics. *Environ. Sci. Technol.* **2010**, *44* (1), 15–23 DOI: 10.1021/es9026248.

(94) Liu, X.; Fu, W.; Da, E.; Shi, X.; Cao, Y.; Rathinasabapathi, B.; Chen, Y.; Ma, L. Q. Microbial siderophores and root exudates enhanced goethite dissolution and Fe/As uptake by As-hyperaccumulator *Pteris vittata*. *Environ. Pollut.* **2017**, *223*, 230–237 DOI: 10.1016/j.envpol.2017.01.016.

(95) Liu, W. J.; Zhu, Y. G.; Smith, F. A.; Smith, S. E. Do iron plaque and genotypes affect arsenate uptake and translocation by rice seedlings (*Oryza sativa* L.) grown in solution culture? *J. Exp. Bot.* **2004**, *55* (403), 1707–1713 DOI: 10.1093/jxb/erh205.

(96) Liu, W. J.; Zhu, Y. G.; Hu, Y.; Williams, P. N.; Gault, A. G.; Meharg, A. A.; Charnock, J. M.; Smith, F. A. Arsenic sequestration in iron plaque, its accumulation and speciation in mature rice plants (*Oryza sativa* L.). *Environ. Sci. Technol.* **2006**, *40* (18), 5730–5736 DOI: 10.1021/es060800v.

(97) Chen, Z.; Zhu, Y. G.; Liu, W. J.; Meharg, A. A. Direct evidence showing the effect of root surface iron plaque on arsenite and aresnate uptake in rice (*Oryza sativa*) roots. *New Phytol.* **2005**, *165* (1), 91–97 DOI: 10.1111/j.1469-8137.2004.01241.x.

(98) Reyes, C.; Lloyd, J. R.; Saltikov, C. W. Geomicrobiology of Iron and Arsenic in Anoxic Sediments. In *Arsenic Contamination of Groundwater*; Ahuja, S., Ed.; John Wiley & Sons, Inc.: Hoboken, NJ, USA, 2008; pp 123–146 DOI: 10.1002/9780470371046.ch6

(99) Lloyd, J. R.; Oremland, R. Microbial Transformations of Arsenic in the Environment: From Soda Lakes to Aquifers. *Elements* **2006**, *2*, 85–90 DOI: 10.2113/gselements.2.2.85.

(100) Islam, F. S.; Gault, A. G.; Boothman, C.; Polya, D. A.; Charnock, J. M.; Chatterjee, D.; Lloyd, J. R. Role of metal-reducing bacteria in arsenic release from Bengal delta sediments. *Nature* **2004**, *430* (6995), 68–71 DOI: 10.1038/nature02638.

(101) Cummings, D. E.; Caccavo, F.; Fendorf, S.; Rosenzweig, R. F. Arsenic mobilization by the dissimilatory Fe(III)-reducing bacterium *Shewanella alga* BrY. *Environ. Sci. Technol.* **1999**, *33* (5), 723–729 DOI: 10.1021/es980541c.

(102) Wang, X.-J.; Chen, X.-P.; Kappler, A.; Sun, G.-X.; Zhu, Y.-G. Arsenic binding to iron (II) minerals produced by an iron (III)-reducing aeromonas strain isolated from paddy soil. *Environ. Toxicol. Chem.* **2009**, *28* (11), 2255–2262 DOI: 10.1897/09-085.1.

(103) Muehe, E. M.; Scheer, L.; Daus, B.; Kappler, A. Fate of arsenic during microbial reduction of biogenic versus abiogenic As-Fe(III)-mineral coprecipitates. *Environ. Sci. Technol.* **2013**, *47* (15), 8297–8307 DOI: 10.1021/es400801z.

(104) Muehe, E. M.; Morin, G.; Scheer, L.; Le Pape, P.; Esteve, I.; Daus, B.; Kappler, A. Arsenic(V) Incorporation in Vivianite during Microbial Reduction of Arsenic(V)-Bearing Biogenic Fe(III) (Oxyhydr)oxides. *Environ. Sci. Technol.* **2016**, *50* (5), 2281–2291 DOI: 10.1021/acs.est.5b04625.

(105) Islam, F. S.; Pederick, R. L.; Gault, A. G.; Adams, L. K.; Polya, D. A.; Charnock, J. M.; Lloyd, J. R. Interactions between the Fe(III) -Reducing Bacterium *Geobacter sulfurreducens* and Arsenate, and Capture of the Metalloid by Biogenic Fe(II). *Appl Env. Microbiolli* **2005**, *71* (12), 8642–8648 DOI: 10.1128/AEM.71.12.8642.

(106) Islam, F. S.; Boothman, C.; Gault, A. G.; Polya, D. A.; Lloyd, J. R. Potential role of the Fe(III)-reducing bacteria *Geobacter* and *Geothrix* in controlling arsenic solubility in Bengal delta sediments. *Mineral. Mag.* **2005**, *69* (5), 865–875 DOI: 10.1180/0026461056950294.

(107) Wang, X.; Chen, X.; Yang, J.; Wang, Z.; Sun, G. Effect of microbial mediated iron plaque reduction on arsenic mobility in paddy soil. *J. Environ. Sci.* **2009**, *21* (11), 1562–1568 DOI: 10.1016/S1001-0742(08)62456-0.

(108) Campbell, K. M.; Malasarn, D. Simultaneous Microbial Reduction of Iron(III) and Arsenic(V) in Suspensions of Hydrous Ferric Oxide. *Env. Sci Technol* **2006**, *40* (19), 5950–5955 DOI: 10.1021/es0600476.

(109) Kocar, B. D.; Herbel, M. J.; Tufano, K. J.; Fendorf, S. Contrasting effects of dissimilatory iron(III) and arsenic(V) reduction on arsenic retention and transport. *Environ. Sci. Technol.* **2006**, *40* (21), 6715–6721 DOI: 10.1021/es061540k.

(110) Oremland, R. S.; Stolz, J. F. Arsenic, microbes and contaminated aquifers. *Trends Microbiol.* **2005**, *13* (2), 45–49 DOI: 10.1155/2011/863623.

(111) Zobrist, J.; Dowdle, P. R.; Davis, J. A.; Oremland, R. S. Mobilization of arsenite by dissimilatory reduction of adsorbed arsenate. *Environ. Sci. Technol.* **2000**, *34* (22), 4747–4753 DOI: 10.1021/es001068h.

(112) Hohmann, C.; Winkler, E.; Morin, G.; Kappler, A. Anaerobic Fe(II)-oxidizing bacteria show as resistance and immobilize as during Fe(III) mineral precipitation. *Environ. Sci. Technol.* **2010**, *44* (1), 94–101 DOI: 10.1021/es900708s.

(113) Hohmann, C.; Morin, G.; Ona-Nguema, G.; Guigner, J. M.; Brown, G. E.; Kappler, A. Molecular-level modes of As binding to Fe(III) (oxyhydr)oxides precipitated by the anaerobic nitrate-reducing Fe(II)-oxidizing Acidovorax sp. strain BoFeN1. *Geochim. Cosmochim. Acta*

2011, *75* (17), 4699–4712 DOI: 10.1016/j.gca.2011.02.044.

(114) Omoregie, E. O.; Couture, R.; Cappellen, P. Van; Corkhill, C. L.; Charnock, J. M.; Polya, D. A.; Vaughan, D.; Vanbroekhoven, K.; Lloyd, R. Arsenic Bioremediation by Biogenic Iron Oxides and Sulfides. *Appl Env. Microbiol* **2013**, *79* (14), 4325–4335 DOI: 10.1128/AEM.00683-13.

(115) Nitzsche, K. S.; Weigold, P.; Lösekann-Behrens, T.; Kappler, A.; Behrens, S. Microbial community composition of a household sand filter used for arsenic, iron, and manganese removal from groundwater in Vietnam. *Chemosphere* **2015**, *138*, 47–59 DOI: 10.1016/j.chemosphere.2015.05.032.

(116) Kleinert, S.; Muehe, E. M.; Posth, N. R.; Dippon, U.; Daus, B.; Kappler, A. Biogenic Fe(III) minerals lower the efficiency of iron-mineral-based commercial filter systems for arsenic removal. *Environ. Sci. Technol.* **2011**, *45* (17), 7533–7541 DOI: 10.1021/es201522n.

(117) Okibe, N.; Koga, M.; Morishita, S.; Tanaka, M.; Heguri, S.; Asano, S.; Sasaki, K.; Hirajima, T. Microbial formation of crystalline scorodite for treatment of As(III) -bearing copper refinery process solution using *Acidianus brierleyi*. *Hydrometallurgy* **2014**, *143*, 34–41 DOI: 10.1016/j.hydromet.2014.01.008.

(118) Mandal, B. K.; Suzuki, K. T. Arsenic round the world : a review. *Talanta* **2002**, *58*, 201–235 DOI: 10.1016/S0039-9140(02)00268-0.

(119) Corkhill, C. L.; Wincott, P. L.; Lloyd, J. R.; Vaughan, D. J. The oxidative dissolution of arsenopyrite (FeAsS) and enargite (Cu3AsS4) by *Leptospirillum ferrooxidans*. *Geochim. Cosmochim. Acta* **2008**, *72* (23), 5616–5633 DOI: 10.1016/j.gca.2008.09.008.

(120) O'Day, P. a; Vlassopoulos, D.; Root, R.; Rivera, N. The influence of sulfur and iron on dissolved arsenic concentrations in the shallow subsurface under changing redox conditions. *Proc. Natl. Acad. Sci. U. S. A.* **2004**, *101* (38), 13703–13708 DOI: 10.1073/pnas.0402775101.

(121) Kim, M.; Nriagu, J.; Haack, S. Arsenic species and chemistry in groundwater of southeast Michigan. *Environ. Pollut.* **2002**, *120*, 379–390 DOI: 10.1016/S0269-7491(02)00114-8.

(122) Rochette, E. A.; Bostick, B. C.; Li, G.; Fendorf, S. Kinetics of arsenate reduction by dissolved sulfide. *Environ. Sci. Technol.* **2000**, *34* (22), 4714–4720 DOI: 10.1021/es000963y.

(123) Kuhn, A.; Sigg, L.; Jul, N. Arsenic Cycling in Eutrophic Lake Greifen, Switzerland: Influence of Seasonal Redox Processes. *Limnol. Ocean.* **1993**, *38* (5), 1052–1059 DOI: 10.4319/lo.1993.38.5.1052.

(124) Newman, D. K.; Beveridge, T. J. Precipitation of Arsenic Trisulfide by *Desulfotomaculum auripigmentum*. *Appl. Environ. Microbiol.* **1997**, *63* (5), 2022–2028.

(125) Rittle, K. A.; Drever, J. I.; Colberg, P. J. S. Precipitation of Arsenic during Bacterial Sulfate Reduction. *Geomicrobiol. J.* **1995**, *13* (1), 1–11 DOI: 10.1080/01490459509378000.

(126) Hoeft, S. E.; Kulp, T. R.; Stolz, J. F.; Hollibaugh, J. T.; Oremland, R. S. Dissimilatory Arsenate Reduction with Sulfide as Electron Donor: Experiments with Mono Lake Water and Isolation of Strain MLMS-1, a Chemoautotrophic Arsenate Respirer. *Appl. Environ. Microbiol.* **2004**, *70* (5), 2741–2747 DOI: 10.1128/AEM.70.5.2741.

(127) Hollibaugh, J. T.; Budinoff, C.; Hollibaugh, R. A.; Ransom, B.; Bano, N. Sulfide Oxidation

Coupled to Arsenate Reduction by a Diverse Microbial Community in a Soda Lake. *Appl Env. Microbiol* **2006**, *72* (3), 2043–2049 DOI: 10.1128/AEM.72.3.2043.

(128) Hoeft, S. E.; Blum, J. S.; Stolz, J. F.; Tabita, F. R.; Witte, B.; King, G. M.; Santini, J. M.; Oremland, R. S. *Alkalilimnicola ehrlichii* sp. nov., a novel, arsenite-oxidizing haloalkaliphilic gammaproteobacterium capable of chemoautotrophic or heterotrophic growth with nitrate or oxygen as the electron acceptor. *Int. J. Syst. Evol. Microbiol.* **2007**, *57* (3), 504–512 DOI: 10.1099/ijs.0.64576-0.

(129) Rhine, E. D.; Phelps, C. D.; Young, L. Y. Anaerobic arsenite oxidation by novel denitrifying isolates. *Environ. Microbiol.* **2006**, *8* (5), 899–908 DOI: 10.1111/j.1462-2920.2005.00977.x.

(130) Sun, W.; Sierra, R.; Fernandez, N.; Sanz, J. L.; Amils, R.; Legatzki, A.; Maier, R. M.; Amils, R.; Field, J. A. Molecular Characterization and *In Situ* Quantification of Anoxic Arsenite Oxidizing Denitrifying Enrichment cultures. *FEMS Microbiol Ecol.* **2009**, *68* (1), 72–85 DOI: 10.1007/s12671-013-0269-8.Moving.

(131) Senko, J. M.; Dewers, T. A.; Krumholz, L. R. Effect of Oxidation Rate and Fe(II) State on Microbial Nitrate-Dependent Fe(III) Mineral Formation. *Appl. Environ. Microbiol.* **2005**, *71* (11), 7172–7177 DOI: 10.1128/AEM.71.11.7172.

(132) Weber, K. A.; Urrutia, M. M.; Churchill, P. F.; Kukkadapu, R. K.; Roden, E. E. Anaerobic redox cycling of iron by freshwater sediment microorganisms. *Environ. Microbiol.* **2006**, *8*, 100–113 DOI: 10.1111/j.1462-2920.2005.00873.x.

(133) Schippers, A.; Jorgensen, B. B. Biogeochemistry of pyrite and iron sulfide oxidation in marine sediments. *Geochim. Cosmochim. Acta* **2002**, *66* (1), 85–92 DOI: 10.1016/S0016-7037(01)00745-1.

(134) Stevenson, F. J. *Humus chemistry: genesis, composition, reactions*, 2 nd.; John Wiley & Sons, 1994.

(135) Jiang, J.; Bauer, I.; Paul, A.; Kappler, A. Arsenic redox changes by microbially and chemically formed semiquinone radicals and hydroquinones in a humic substance model quinone. *Environ. Sci. Technol.* **2009**, *43* (10), 3639–3645 DOI: 10.1021/es803112a.

(136) Sharma, P.; Rolle, M.; Kocar, B. D.; Fendorf, S.; Kapppler, A. Influence of Natural Organic Matter on As Transport and Retention. *Environ. Sci. Technol.* **2010**, *45* (2), 546–553 DOI: 10.2134/jeq2003.1393.

(137) Sharma, P.; Kappler, A. Desorption of arsenic from clay and humic acid-coated clay by dissolved phosphate and silicate. *J. Contam. Hydrol.* **2011**, *126* (3–4), 216–225 DOI: 10.1016/j.jconhyd.2011.08.005.

(138) Bauer, M.; Blodau, C. Arsenic distribution in the dissolved, colloidal and particulate size fraction of experimental solutions rich in dissolved organic matter and ferric iron. *Geochim. Cosmochim. Acta* **2009**, *73* (3), 529–542 DOI: 10.1016/j.gca.2008.10.030.

(139) Thomasarrigo, L. K.; Mikutta, C.; Byrne, J.; Barmettler, K.; Kappler, A.; Kretzschmar, R. Iron and Arsenic Speciation and Distribution in Organic Flocs from Streambeds of an Arsenic-Enriched Peatland. *Environ. Sci. Technol.* **2014**, *48*, 13218–13228 DOI: 10.1021/es503550g.

(140) Macalady, D. L.; Ranville, J. F. The chemistry and geochemistry of natural organic matter (NOM). In *Perspectives in environmental chemistry*; Macalady, D. L., Ed.; Oxford University Press: New York, 1988; pp 94–137.

(141) Buschmann, J.; Kappeler, A.; Lindauer, U.; Kistler, D.; Berg, M.; Sigg, L. Arsenite and arsenate binding to dissolved humic acids: Influence of pH, type of humic acid, and aluminum. *Environ. Sci. Technol.* **2006**, *40* (19), 6015–6020 DOI: 10.1021/es061057+.

(142) Sharma, P.; Ofner, J.; Kappler, A. Formation of binary and ternary colloids and dissolved complexes of organic matter, Fe and As. *Environ. Sci. Technol.* **2010**, *44* (12), 4479–4485 DOI: 10.1021/es100066s.

(143) Langner, P.; Mikutta, C.; Kretzschmar, R. Arsenic sequestration by organic sulphur in peat. *Nat. Geosci.* **2011**, *5* (1), 66–73 DOI: 10.1038/ngeo1329.

(144) Rowland, H. A. L.; Polya, D. A.; Lloyd, J. R.; Pancost, R. D. Characterisation of organic matter in a shallow, reducing, arsenic-rich aquifer, West Bengal. *Org. Geochem.* **2006**, *37*, 1101–1114 DOI: 10.1016/j.orggeochem.2006.04.011.

(145) Rowland, H. A. L.; Pederick, R. L.; Polya, D. A.; Pancost, R. D.; Dongen, B. E. Van; Gault, A. G.; Vaughan, D. J.; Bryant, C.; Anderson, B.; Lloyd, J. R. The control of organic matter on microbially mediated iron reduction and arsenic release in shallow alluvial aquifers, Cambodia. *Geobiology* **2007**, *5*, 281–292 DOI: 10.1111/j.1472-4669.2007.00100.x.

(146) Rowland, H. A. L.; Boothman, C.; Pancost, R.; Gault, A. G.; Polya, D. A.; Lloyd, J. R. The Role of Indigenous Microorganisms in the Biodegradation of Naturally Occurring Petroleum, the Reduction of Iron, and the Mobilization of Arsenite from West Bengal. *J. Environ. Qual.* **2009**, *38*, 1598–1607 DOI: 10.2134/jeq2008.0223.

(147) Bauer, M.; Blodau, C. Mobilization of arsenic by dissolved organic matter from iron oxides, soils and sediments. *Sci. Total Environ.* **2006**, *354* (2–3), 179–190 DOI: 10.1016/j.scitotenv.2005.01.027.

(148) Huang, H.; Jia, Y.; Sun, G.-X.; Zhu, Y.-G. Arsenic speciation and volatilization from flooded paddy soils amended with different organic matters-supporting information. *Environ. Sci. Technol.* **2012**, *46* (4), 2163–2168 DOI: 10.1021/es203635s.

(149) Jia, Y.; Huang, H.; Chen, Z.; Zhu, Y. G. Arsenic uptake by rice is influenced by microbe-mediated arsenic redox changes in the rhizosphere. *Environ. Sci. Technol.* **2014**, *48* (2), 1001–1007 DOI: 10.1021/es403877s.

(150) Palmer, N. E.; Freudenthal, J. H.; Wandruszka, R. Von. Reduction of Arsenates by Humic Materials. *Environ. Chem.* **2006**, *3*, 131–136 DOI: 10.1071/EN05081.

(151) Palmer, N. E.; Wandruszka, R. Von. Humic acids as reducing agents: the involvement of quinoid moieties in arsenate reduction. *Environ. Sci. Pollut. Res.* **2010**, *17*, 1362–1370 DOI: 10.1007/s11356-010-0322-2.

(152) Redman, A. D.; Macalady, D.; Ahmann, D. Natural organic matter affects arsenic speciation and sorption onto hematite. *Environ. Sci. Technol.* **2002**, *36* (13), 2889–2896 DOI: 10.1021/es0112801.

(153) Niggemyer, A.; Spring, S.; Stackebrandt, E. Isolation and Characterization of a Novel As (V)

-Reducing Bacterium: Implications for Arsenic Mobilization and the Genus *Desulfitobacterium*. *Appl. Environ. Microbiol.* **2001**, *67* (12), 5568–5580 DOI: 10.1128/AEM.67.12.5568.

(154) Stolz, J. F.; Oremland, R. S. Bacterial respiration of arsenic and selenium. *FEMS Microbiol Rev* **1999**, *23*, 615–627 DOI: 10.1111/j.1574-6976.1999.tb00416.x.

(155) Handley, K. M.; Hery, M.; Lloyd, J. R. Redox cycling of arsenic by the hydrothermal marine bacterium *Marinobacter santoriniensis*. *Env. Microbiol* **2009**, *11* (6), 1601–1611 DOI: 10.1111/j.1462-2920.2009.01890.x.

(156) Lear, G.; Song, B.; Gault, A. G.; Polya, D. A.; Lloyd, J. R. Molecular analysis of arsenate-reducing bacteria within Cambodian sediments following amendment with acetate. *Appl. Environ. Microbiol.* **2007**, *73* (4), 1041–1048 DOI: 10.1128/AEM.01654-06.

(157) Liu, A.; Garcia-dominguez, E.; Rhine, E. D.; Young, L. Y. A novel arsenate respiring isolate that can utilize aromatic substrates. *FEMS Microbiol Ecol* **2004**, *48*, 323–332 DOI: 10.1016/j.femsec.2004.02.008.

(158) Liu, W. J.; McGrath, S. P.; Zhao, F. J. Silicon has opposite effects on the accumulation of inorganic and methylated arsenic species in rice. *Plant Soil* **2014**, *376* (1–2), 423–431 DOI: 10.1007/s11104-013-1991-7.

(159) Escudero, L. V; Casamayor, E. O.; Chong, G.; Pedro, C.; Demergasso, C. Distribution of Microbial Arsenic Reduction, Oxidation and Extrusion Genes along a Wide Range of Environmental Arsenic Concentrations. *PLoS One* **2013**, *8* (10), e78890 DOI: 10.1371/journal.pone.0078890.

(160) Han, Y.; Fu, J.; Xiang, P.; Cao, Y.; Rathinasabapathi, B.; Chen, Y.; Ma, L. Q. Arsenic and phosphate rock impacted the abundance and diversity of bacterial arsenic oxidase and reductase genes in rhizosphere of As-hyperaccumulator *Pteris vittata*. *J. Hazard. Mater.* **2017**, *321*, 146–153 DOI: 10.1016/j.jhazmat.2016.08.079.

(161) Wang, X.; Rathinasabapathi, B.; Oliveira, L. M. de; Guilherme, L. R. G.; Ma, L. Q. Bacteria-Mediated Arsenic Oxidation and Reduction in the Growth Media of Arsenic Hyperaccumulator *Pteris vittata*. *Env. Sci Technol* **2012**, *46*, 11259–11266 DOI: 10.1021/es300454b.

(162) Zhang, S. Y.; Zhao, F. J.; Sun, G. X.; Su, J. Q.; Yang, X. R.; Li, H.; Zhu, Y. G. Diversity and abundance of arsenic biotransformation genes in paddy soils from southern china. *Environ. Sci. Technol.* **2015**, *49* (7), 4138–4146 DOI: 10.1021/acs.est.5b00028.

(163) Jia, Y.; Huang, H.; Sun, G. X.; Zhao, F. J.; Zhu, Y. G. Pathways and relative contributions to arsenic volatilization from rice plants and paddy soil. *Environ. Sci. Technol.* **2012**, *46* (15), 8090–8096 DOI: 10.1021/es300499a.

(164) Xiao, K. Q.; Li, L. G.; Ma, L. P.; Zhang, S. Y.; Bao, P.; Zhang, T.; Zhu, Y. G. Metagenomic analysis revealed highly diverse microbial arsenic metabolism genes in paddy soils with low-arsenic contents. *Environ. Pollut.* **2016**, *211*, 1–8 DOI: 10.1016/j.envpol.2015.12.023.

(165) Lu, K.; Mahbub, R.; Cable, P. H.; Ru, H.; Parry, N. M. A.; Bodnar, W. M.; Wishnok, J. S.; Styblo, M.; Swenberg, J. A.; Fox, J. G.; et al. Gut Microbiome Phenotypes Driven by Host Genetics Affect Arsenic Metabolism. *Chem. Res. Toxicol.* **2014**, *27*, 172–174 DOI:

10.1021/tx400454z.

(166) Cai, L.; Yu, K.; Yang, Y.; Chen, B. W.; Li, X. D.; Zhang, T. Metagenomic exploration reveals high levels of microbial arsenic metabolism genes in activated sludge and coastal sediments. *Appl. Microbiol. Biotechnol.* **2013**, *97* (21), 9579–9588 DOI: 10.1007/s00253-012-4678-8.

(167) Rascovan, N.; Javier, M.; Martín P, V.; María, E. F. Metagenomic study of red biofilms from Diamante Lake reveals ancient arsenic bioenergetics in haloarchaea Metagenomic study of red biofilms from Diamante Lake reveals ancient arsenic bioenergetics in haloarchaea. *ISME J.* **2016**, *10*, 299–309 DOI: 10.1038/ismej.2015.109.

(168) Luo, J.; Bai, Y.; Liang, J.; Qu, J. Metagenomic Approach Reveals Variation of Microbes with Arsenic and Antimony Metabolism Genes from Highly Contaminated Soil. *PLoS One* **2014**, *9* (10), e108185 DOI: 10.1371/journal.pone.0108185.

(169) Sangwan, N.; Lambert, C.; Sharma, A.; Gupta, V.; Khurana, P.; Khurana, J. P.; Sockett, R. E.; Gilbert, J. A.; Lal, R. Arsenic rich Himalayan hot spring metagenomics reveal genetically novel predator – prey genotypes. *Environ. Microbiol. Rep.* **2015**, *7*, 812–823 DOI: 10.1111/1758-2229.12297.

(170) Aguiar-pulido, V.; Huang, W.; Suarez-ulloa, V.; Cickovski, T.; Mathee, K.; Narasimhan, G. Metagenomics, metatranscriptomics, and metabolomics approaches for microbiome analysis. *Evol. Bioinforma.* **2016**, *12* (S1), 5–16 DOI: 10.4137/EBO.S36436.

(171) Kour, M. Isolation and Characterization of Metal Resistance Genes by using Metatranscriptomic Approach. Master Dissertation, Thapar University, Patiala, Punjab-147004, 2014.

(172) Carvalhais, L. C.; Dennis, P. G.; Tyson, G. W.; Schenk, P. M. Application of metatranscriptomics to soil environments. *J. Microbiol. Methods* **2012**, *91* (2), 246–251 DOI: 10.1016/j.mimet.2012.08.011.

(173) Gilbert, J. A.; Field, D.; Huang, Y.; Edwards, R.; Li, W.; Gilna, P.; Joint, I. Detection of Large Numbers of Novel Sequences in the Metatranscriptomes of Complex Marine Microbial Communities. *PLoS One* **2008**, *3* (8), e3042 DOI: 10.1371/journal.pone.0003042.

(174) Lafuente, A.; Patricia, P.; Molina-s, D.; Caviedes, M. A.; Rodr, I. D. Unraveling the effect of arsenic on the model *Medicago – Ensifer* interaction: a transcriptomic meta-analysis. *ISME J.* **2015**, *205*, 255–272 DOI: 10.1111/nph.13009.

(175) Zhang, Y.; Chen, S.; Hao, X.; Su, J. Q.; Xue, X.; Yan, Y.; Zhu, Y. G.; Ye, J. Transcriptomic analysis reveals adaptive responses of an enterobacteriaceae strain LSJC7 to arsenic exposure. *Front. Microbiol.* **2016**, *7*, 1–12 DOI: 10.3389/fmicb.2016.00636.

(176) Moran, M. A.; Satinsky, B.; Gifford, S. M.; Luo, H.; Rivers, A.; Chan, L.; Meng, J.; Durham, B. P.; Shen, C.; Varaljay, V. A.; et al. Sizing up metatranscriptomics. **2012**, *7* (2), 237–243 DOI: 10.1038/ismej.2012.94.

(177) VerBerkmoes, N. C.; Denef, V. J.; Hettich, R. L.; Banfield, J. F. Functional analysis of natural microbial consortia using community proteomics. *Nat Rev Microbiol* **2009**, *7* (3), 196–205 DOI: 10.1038/nrmicro2080.

(178) Fiehn, O. Metabolomics - The link between genotypes and phenotypes. *Plant Mol. Biol.* **2002**,

48 (1–2), 155–171 DOI: 10.1023/A:1013713905833.

(179) Schneider, T.; Riedel, K. Environmental proteomics: Analysis of structure and function of microbial communities. *Proteomics* **2010**, *10* (4), 785–798 DOI: 10.1002/pmic.200900450.

(180) Lankadurai, B. P.; Nagato, E. G.; Simpson, M. J. Environmental metabolomics: an emerging approach to study organism responses to environmental stressors. *Environ. Rev.* **2013**, *21*, 180–205 DOI: 10.1139/er-2013-0011.

(181) Halter, D.; Cordi, A.; Gribaldo, S.; Gallien, S.; Goulhen-Chollet, F.; Heinrich-Salmeron, A.; Carapito, C.; Pagnout, C.; Montaut, D.; Seby, F.; et al. Taxonomic and functional prokaryote diversity in mildly arsenic-contaminated sediments. *Res. Microbiol.* **2011**, *162* (9), 878–887 DOI: 10.1016/j.resmic.2011.06.001.

(182) Shi, X.; Wei, X.; Koo, I.; Schmidt, R. H.; Yin, X.; Kim, S. H.; Vaughn, A.; Mcclain, C. J.; Arteel, G. E.; Zhang, X.; et al. Metabolomic Analysis of the Effects of Chronic Arsenic Exposure in a Mouse Model of Diet-Induced Fatty Liver Disease. *J. Proteome Res.* **2014**, *13*, 547–554 DOI: 10.1021/pr400719u.

(183) Bastida, F.; Moreno, J. L.; Nicolás, C.; Hernández, T.; García, C. Soil metaproteomics: a review of an emerging environmental science. Significance, methodology and perspectives. *Eur. J. Soil Sci.* **2009**, *60* (6), 845–859 DOI: 10.1111/j.1365-2389.2009.01184.x.

(184) Lu, K.; Abo, R. P.; Schlieper, K. A.; Michelle, E.; Levine, S.; Wishnok, J. S.; Swenberg, J. A.; Tannenbaum, S. R.; Fox, J. G. Arsenic Exposure Perturbs the Gut Microbiome and Its Metabolic Profile in Mice: An Integrated Metagenomics and Metabolomics Analysis. *Environ. Health Perspect.* **2014**, *122*, 284–291 DOI: 10.1289/ehp.1307429.

(185) Tripathi, R. D.; Tripathi, P.; Dwivedi, S.; Dubey, S.; Chatterjee, S.; Chakrabarty, D.; Trivedi, P. K. Arsenomics: Omics of arsenic metabolism in plants. *Front. Physiol.* **2012**, *3*, 1–14 DOI: 10.3389/fphys.2012.00275.

(186) Sacheti, P.; Bhonsle, H.; Patil, R.; Kulkarni, M. J.; Srikanth, R.; Gade, W. Arsenomics of *Exiguobacterium* sp. PS (NCIM 5463). *RSC Adv.* **2013**, *3*, 9705–9713 DOI: 10.1039/c3ra40897c.

(187) Zhang, S.; Williams, P. N.; Luo, J.; Zhu, Y. Microbial mediated arsenic biotransformation in wetlands. *Front. Environ. Sci. eng.* **2017**, *11* (1), 1–11 DOI: 10.1007/s11783-017-0893-y.

(188) Meharg, A. A.; Williams, P. N.; Adomako, E.; Lawgali, Y. Y.; Deacon, C.; Villada, A.; Cambell, R. C. J.; Sun, G.; Zhu, Y. G.; Feldmann, J.; et al. Geographical variation in total and inorganic arsenic content of polished (white) rice. *Environ. Sci. Technol.* **2009**, *43* (5), 1612–1617 DOI: 10.1021/es802612a.

(189) Sun, G. X.; Williams, P. N.; Carey, A. M.; Zhu, Y. G.; Deacon, C.; Raab, A.; Feldmann, J.; Islam, R. M.; Meharg, A. A. Inorganic arsenic in rice bran and its products are an order of magnitude higher than in bulk grain. *Environ. Sci. Technol.* **2008**, *42* (19), 7542–7546 DOI: 10.1021/es801238p.

(190) Sun, G. X.; Williams, P. N.; Zhu, Y. G.; Deacon, C.; Carey, A. M.; Raab, A.; Feldmann, J.; Meharg, A. A. Survey of arsenic and its speciation in rice products such as breakfast cereals, rice crackers and Japanese rice condiments. *Environ. Int.* **2009**, *35* (3), 473–475 DOI:

10.1016/j.envint.2008.07.020.
(191) Norton, G. J.; Pinson, S. R. M.; Alexander, J.; Mckay, S.; Hansen, H.; Duan, G.; Islam, M. R.; Islam, S.; Stroud, J. L.; Zhao, F.; et al. Variation in grain arsenic assessed in a diverse panel of rice (*Oryza sativa*) grown in multiple sites. *New Phytol.* **2011**, *193*, 650–664 DOI: 10.1111/j.1469-8137.2011.03983.x.
(192) Lomax, C.; Liu, W. J.; Wu, L.; Xue, K.; Xiong, J.; Zhou, J.; McGrath, S. P.; Meharg, A. A.; Miller, A. J.; Zhao, F. J. Methylated arsenic species in plants originate from soil microorganisms. *New Phytol.* **2012**, *193* (3), 665–672 DOI: 10.1111/j.1469-8137.2011.03956.x.
(193) Oremland, R. S.; Dowdle, P. R.; Hoeft, S.; Sharp, J. O.; Schaefer, J. K.; Miller, L. G.; Blum, J. S.; Smith, R. L.; Bloom, N. S.; Wallschlager, D. Bacterial dissimilatory reduction of arsenate and sulfate in meromictic Mono Lake, California. *Geochim. Cosmochim. Acta* **2000**, *64* (18), 3073–3084 DOI: 10.1016/S0016-7037(00)00422-1.
(194) Kulp, T. R.; Hoeft, S. E.; Miller, L. G.; Saltikov, C.; Murphy, J. N.; Han, S.; Lanoil, B.; Oremland, R. S. Dissimilatory Arsenate and Sulfate Reduction in Sediments of Two Hypersaline, Arsenic-Rich Soda Lakes: Mono and Searles Lakes, California. *Appl. Environ. Microbiol.* **2006**, *72* (10), 6514–6526 DOI: 10.1128/AEM.01066-06.
(195) Reed, D. C.; Algar, C. K.; Huber, J. A.; Dick, G. J. Gene-centric approach to integrating environmental genomics and biogeochemical models. *Proc. Natl Acad. Sci.* **2014**, *111* (5), 1879–1884 DOI: 10.1073/pnas.1313713111.
(196) Louca, S.; Hawley, A. K.; Katsev, S.; Torres-Beltran, M.; Bhatia, M. P.; Kheirandish, S.; Michiels, C. C.; Capelle, D.; Lavik, G.; Doebeli, M.; et al. Integrating biogeochemistry with multiomic sequence information in a model oxygen minimum zone. *Proc. Natl. Acad. Sci.* **2016**, *113* (40), E5925–E5933 DOI: 10.1073/pnas.1602897113.

Legends

Figure 1. Proposed pathways for arsenic redox reactions and synthesis of novel organoarsenicals.

Figure 2. The model of effect of Fe, N, S and natural organic matter (NOM) on microbes involved in arsenic biogeochemical cycling highlights proteins associated with elemental metabolisms. Green ovals denote arsenic transporters, yellow ovals denote transmembrane enzymes. Red words are enzymes, blue words are related arsenic compounds. The full name of enzymes that were not mentioned in the text was provided in the follow, NarG, transmembrane nitrate reductase that drives the nitrate

reduction to nitrite; Nas, cytoplasmic-assimilatory nitrate reductase that drives the nitrate reduction to nitrite; NapA, periplasmic-dissimilatory nitrate reductase; Nir /Nrf (associated with NapA), nitrite reductase that drives the nitrite reduction to nitric oxide; NorB/C, nitric oxide reductase that drives the nitric oxide reduction to nitrous oxide; NosZ, nitrous oxide reductase that drives the nitrous oxide reduction to nitrogen; Nif, nitrogenase that catalyzes the nitrogen fixation to ammonia; Hs, hydrazine synthase that catalyzes the production of nitrogen from nitrous oxide and ammonia; cyt, cytochrome.

1203

1204 Table 1. The genes involved in arsenic metabolisms

Gene	Protein	Protein abbreviation	function	Reference
As(III) oxidation				
aioA and *aioB*	As(III) oxidase	AioAB	Oxidate As(III)	[9]
arxA	As(III) oxidase	ArxAB	Oxidate As(III)	[10]
arxB/arxB2	[4Fe-4S] containing protein	ArxB/ ArxB2	Unknown function	[11]
aioX/arxX	As(III)-binding protein	AioX/ArxX	Involved in As(III)-based signaling and regulation of As(III) oxidation	[12, 11]
aioS/arxS	sensor histidine kinase	AioS /ArxS	One part of two-component signal transduction system	[9, 11]
aioR/arxR	transcriptional regulator	AioR/ArxR	Regulate the expression of *aio/arx* operon	[9, 11]
moeA	molybdenum cofactor biosynthesis protein	MoeA	Synthesize the molybdenum cofactor of AioAB oxidase	[9]
arxC	membrane anchoring and quinol oxidoreductase subunit	ArxC	Involved in As(III) oxidation	[11]
arxD	TorD-like molybdoenzyme chaperone	ArxD	Involved in As(III) oxidation	[11]
arsH	organoarsenical oxidase	ArsH	Oxidate trivalent methylated and aromatic arsenicals, reduce chromium and iron	[13, 14]
As(V) reduction				
arrA and *arrB*	As(V) respiratory reductases	ArrAB	Reduce As(V)	[15, 16]
arrC	As(V) reductase membranous subunit	ArrC	Involved in As(V) reduction	[17]
arrD	As(V) reductase chaperon	ArrD	Involved in As(V) reduction	[17]
arrS	sensor histidine kinase	ArrS	Regulate the expression of *arr* operon	[17]
arsC	As(V) reductase	ArsC	Reduce As(V)	[18]
ACR2	As(V) reductase	ACR2	Reduce As(V)	[19]

ACR1	transcriptional regulatory protein	ACR1	Regulate the expression of *ACR* genes	[19]
arrR/arsR	arsenic-responsive repressor	ArrR /ArsR	Regulate the expression of *arr*/*ars* operon	[17, 20]
GstB	glutathione S-transferase B	GstB	Reduce As(V) to As(III) with reduced GSH	[21]
arsN	acetyltransferase	ArsN	Putative As(V) reductase	[22]
Arsenic methylation and demethylation				
arsM	As(III) S-adenosylmethionine (SAM) methyltransferases	ArsM	Methylate arsenic	[23]
arsI	A C · As lyase	ArsI	Catalyze demethylation of trivalent organoarsenicals	[24]
Arsenic transport				
arsB	As(III)-pump protein	ArsB	Extrude As(III) from the cell	[25]
arsA	As(III)-pump ATPase	ArsA	The catalytic subunit of an oxyanion-translocating ATPase	[26]
arsD	arsenical metallochaperone	ArsD	Transfer trivalent metalloids to ArsA	[26]
apqS	aquaglyceroporin	AqpS	Extrude As(III) from the cell	[27]
ACR3	As(III) permease	ACR3	Extrude As(III) from the cell	[19]
Ycf1p	yeast cadmium factor protein cytosolic	Ycf1p	Sequester cytosolic As(III)/glutathione complex into vacuoles	[28]
arsJ	organoarsenical efflux permease	ArsJ	Extrude organoarsenicals from the cell	[29]
arsP	efflux system specific for trivalent organoarsenicals	ArsP	Extrude trivalent organoarsenicals from the cell	[30]
pgpA	P-glycoprotein-related protein	PgpA	Recognize and transport thiol-metal conjugates	[31]
Unknown functions				
arsO	putative flavin-binding monooxygenase	ArsO	Unknown function	[32]
arsT	putative thioredoxin reductase	ArsT	Unknown function	[32]

1205