

# Biological control agent selection under environmental change using functional responses, abundances and fecundities; the Relative Control Potential (RCP) metric

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1	Biological control agent selection under environmental change using
2	functional responses, abundances and fecundities; the Relative
3	Control Potential (RCP) metric
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#### 21 ABSTRACT

We currently lack the capacity to rapidly and reliably predict the efficacy of biological 22 23 control agents due to inadequate consistency in derivations of functional and numerical responses and potential effects of context-dependencies. Here, we propose and apply a novel 24 metric, Relative Control Potential (RCP), which combines the functional response (FR, per 25 26 *capita* effect) with proxies for the numerical response (NR, agent population response) to 27 compare agent efficacies, where  $RCP = FR \times abundance$  (or other proxies e.g. fecundity). The RCP metric is a comparative ratio between potential biocontrol agents, where values > 128 29 indicate higher relative control efficacy. Further, RCP can compare the efficacy of agents under environmental contexts, such as temperature change. We thus derived the RCP for two 30 31 predatory cyclopoid copepods, Macrocyclops albidus (Cyclopoida: Cyclopidae) and Megacyclops viridis (Cyclopoida: Cyclopidae), towards larvae of the mosquito Culex pipiens 32 (Diptera: Culicidae) under temperatures representative of current and future climate. Both 33 34 copepods exhibited potentially population destabilising Type II FRs, with increasing temperatures inducing greater magnitude FRs through increased attack rates and decreased 35 handling times. Attack rates by *M. albidus* were higher than *M. viridis*, yet handling times 36 and maximum feeding rates were similar between the species across all temperatures. The 37 38 inclusion of abundance data drives an elevated RCP of M. albidus and the integration of fecundity drives greater RCP of *M. albidus* at peak temperatures.  $Q_{10}$  values are indicative of 39 increased feeding activity by both copepods with temperature increases, however relative 40 feeding level increases of *M. viridis* slowed towards the peak temperature. We present RCP 41 42 calculations and biplots that represent the comparative efficacies of the two biological control agents across temperatures. The Relative Control Potential (RCP) metric thus provides a new 43 tool for practitioners to better assess the potential efficacy of biocontrol agents before their 44 45 integration into management approaches for pests, vectors and invasive species.

46 Keywords: Functional response; Numerical response; Relative Control Potential; *Culex* 

47 pipiens; Macrocyclops albidus; Megacyclops viridis

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# 49 **1. Introduction**

50 Biological control has been applied to manage pests, vectors and invasive species in a variety of ecological systems (O'Neil, 1990; Marten and Reid, 2007; Van Driesche and 51 Bellows, 2011; Calvo et al. 2016). However, attempts to reveal agent efficacy through the 52 53 coupling of functional and numerical responses (FRs, NRs) are limited in practice, reducing our predictive capacity for population-level effects (but see Heisswolf et al. 2009; Costa et al. 54 2017). Further, natural systems are characterised by a number of abiotic and biotic context-55 dependencies that can alter species interaction strengths, including structural complexity 56 (Barrios-O'Neill et al. 2015), temperature (Wasserman et al. 2016; South et al. 2017), 57 58 dissolved oxygen (Laverty et al. 2015), parasitism (Bunke et al. 2015; Laverty et al. 2017a) and multiple/higher predators (Alexander et al. 2013; Barrios-O'Neill et al. 2014). Thus, 59 rapid and reliable FR and NR derivations under context-dependencies are critical for the 60 61 future of biocontrol strategies.

62 Effects associated with environmental change can be particularly profound in 63 modulating natural systems. Indeed, climate change, coupled with urbanisation, is stimulating an unprecedented change in the population dynamics and status of mosquito vectors and their 64 transmission of disease (Townroe and Callaghan, 2014; Medlock and Leach, 2015; Siraj et al. 65 66 2017), with mosquito invasions increasing with the transportation of goods and humans (e.g. Yee, 2016; Medlock et al. 2017). The *Culex pipiens* (Diptera: Culicidae; Linnaeus, 1758) 67 68 complex is widespread globally and act as one of the primary vectors of West Nile virus in the USA and continental Europe (Hubalek and Halouzka, 1999; Fonseca et al. 2004). 69

70 Freshwater cyclopoid copepods exhibit marked potential for the biological control of mosquitoes (Marten and Reid, 2007; Baldacchino et al. 2017), and have been operationalised 71 72 in large-scale field applications (Kay and Nam, 2005). However, we require rapid assessment of the relative biocontrol potential of such agents under changing climatic conditions. In this 73 study, we therefore present and apply a new metric, based on FRs and NRs, to compare the 74 efficacy of two native and widely-distributed copepods, Macrocyclops albidus (Cyclopoida: 75 76 Cyclopidae; Jurine, 1820) and *Megacyclops viridis* (Cyclopoida: Cyclopidae; Jurine, 1820) 77 under current and predicted temperature regimes.

Functional responses (FRs) quantify consumption under differing resource densities, 78 describing the key components of search, capture and handling time. Three broad FR types 79 80 have been described: the linear Type I, hyperbolic Type II and sigmoidal Type III (Solomon, 1949; Holling, 1959). Functional response form and magnitude are both powerful predictors 81 of the impacts of consumers on resource populations across taxonomic and trophic groups 82 83 (Dick et al. 2014; 2017). However, as FRs only assess per capita impacts, incorporation of the Numerical Response (NR), that is the consumer population response, is also required to 84 discern the Total Response (TR) of consumers, whereby: 85

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 $TR = FR \times NR$ 

In comparison to FRs, NRs are inherently more nebulous and difficult to quantify (Dick et al. 2017). Thus, simple consumer abundance (AB) has recently been proposed as a proxy for the numerical response, giving the 'Impact Potential' (IP) metric, which has proved robust in predicting ecological impact in the context of invasion biology (Dick et al. 2017; Laverty et al. 2017b):

$$IP = FR \times AB$$

(2)

(3)

where FR is the maximum feeding rate (reciprocal of handling time) and AB is a measure of
consumer field abundance. In the context of biocontrol, we can express this as 'Control
Potential', CP:

98  $CP = FR \times AB$ 

99

where FR is the maximum feeding rate as above. However, in addition, we propose that
attack rate be used as a second measure of FR. The attack rate parameter describes the slope
of the FR curve at low prey densities and high attack rates can thus be particularly
destabilising to prey populations. CP as an absolute measure is, however, rather meaningless,
and needs a comparator. Thus, where two or more biocontrol agents require assessment as to
their relative potential efficacies, CP can become 'Relative Control Potential' (RCP):

106 
$$\operatorname{RCP} = \left(\frac{\operatorname{FR}\operatorname{agent}A}{\operatorname{FR}\operatorname{agent}B}\right) \times \left(\frac{\operatorname{AB}\operatorname{agent}A}{\operatorname{AB}\operatorname{agent}B}\right)$$

107 (4)

where RCP = 1, we predict no difference between biocontrol agents; for RCP < 1, we predict agent A to have lesser efficacy than agent B; whereas when RCP > 1, agent A is predicted to have greater efficacy than agent B. Further, increasing values above 1 indicate increasing relative efficacy of agent A compared to agent B.

Furthermore, we propose the use of fecundity as a second proxy for NR, which enables the incorporation into RCP of how quickly biocontrol agents can proliferate. Error can also be incorporated into the RCP metric depending on data availability, using a probability density function (pdf) to generate confidence intervals (CIs) and probabilities that RCP > 1 or > 10 (see Dick et al. 2017). Moreover, as environmental gradients such as 117 temperature can have profound impacts on consumer-resource interactions (Englund et al. 118 2011; Rall et al. 2012), RCP can be used to compare the efficacy of each biocontrol agent 119 across environmental gradients. Here, we apply the RCP metric (Eqn. 4) to compare the 120 biological control potentials of the copepods *M. albidus* and *M. viridis* towards the mosquito 121 complex *C. pipiens* over a temperature gradient reflective of current and future UK climate 122 change scenarios. We also apply the  $Q_{10}$  coefficient to further illustrate feeding activity 123 responses of the two agents across temperatures (Bennett, 1990).

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125 2. Materials and methods

### 126 2.1. Animal collection and rearing

M. albidus and M. viridis were collected at Glastry Clay Pit Ponds, Northern Ireland 127 (54°29'18.5"N; 5°28'19.9"W) in January 2017 and kept in Queen's Marine Laboratory, 128 Portaferry, N. Ireland, at  $25 \pm 2$  °C under a 16:8 light:dark regime and 50 - 60% relative 129 humidity, since these conditions stimulated proliferation. Cultures were initiated using 130 ovigerous females, placed individually into 250 mL cups with dechlorinated tap water and 131 fed ad libitum with Chilomonas paramecium and Paramecium caudatum to obtain nauplii. 132 Starter cultures of these protozoans were available commercially (Sciento, Manchester, 133 134 England) and cultured under the same laboratory conditions in 2 L glass beakers using autoclaved wheat seeds, with C. paramecium providing nourishment for nauplii and early 135 copepodids and P. caudatum for late copepodids and adults. Adult copepods were identified 136 by Maria Holyńska, Museum and Institute of Zoology, Warsaw, Poland. Copepods were 137 mass-reared in 10 L tanks and fed *ad libitum* on the protozoan diet. At maturity, copepods 138 were maintained at  $12 \pm 2$  °C under a 12:12 light and dark regime and acclimatised for 7 d 139

prior to experimentation in 5 L holding arenas of 22 cm diameter fed *ad libitum* on theprotozoan prey.

142 *Culex pipiens* were obtained from a laboratory colony established at the University of Reading, England several years prior, originating from field-collected mosquitoes at the 143 Pirbright Institute, Surrey, England. The colony was sustained under the same conditions as 144 145 the copepod cultures, at  $25 \pm 2$  °C in  $32.5 \times 32.5 \times 32.5$  cm cages (Bugdorm, Watkins and 146 Doncaster, Leominster, England) and fed three times per week with defibrinated horse blood (TCS Biosciences, Buckingham, England) using a Hemotek® blood-feeding system 147 (Hemotek Ltd., Accrington, England) and additionally provided with cotton pads soaked in a 148 10% sucrose solution. Cages contained black cups filled with 200 mL dechlorinated tap water 149 150 for oviposition. Egg rafts were extracted three times per week and placed into larval bowls containing 3 L dechlorinated tap water, and fed *ad libitum* with ground guinea pig pellets 151 (Pets at Home, Newtownabbey, Northern Ireland) until pupation. 152

# 153 2.2. Experimental procedure

Non-ovigerous adult female M. albidus and M. viridis (1.6 - 1.8 mm and 2.0 - 2.3 mm)154 mm body length excluding caudal setae, respectively) were selected for experiments. We 155 selected non-ovigerous females to standardise predators as cyclopoids are sexually dimorphic 156 (Laybourn-Parry et al. 1988) and to eliminate cannibalism of hatching juveniles (Toscano et 157 al. 2016). First instar C. pipiens larvae (1.1 - 1.3 mm) were used as prey. Functional response 158 159 experiments were undertaken in transparent polypropylene cups (42 mm dia.) containing 20 mL dechlorinated tap water from a continuously aerated source in a 12:12 light and dark 160 regime over 24 h at temperatures representing reasonable current autumn/winter (12 °C), 161 spring/summer (16 °C) and future spring/summer (20 °C) conditions in the UK ( $\pm 0.1$  °C; 162 Clifton NEIB water baths; Hulme et al. 2002; Hammond and Pryce, 2007). Dissolved oxygen 163

was monitored using a YSI model 550A meter (Letchworth, England) to ensure levels 164 remained above 80% saturation. Both predators and prev were acclimatised to the two 165 166 elevated temperatures over a two hour period prior to experiments; temperatures were increased every 30 minutes by either 1 °C or 2 °C (i.e. to 16 °C or 20 °C). Following the 167 acclimatisation period, we added single adult females of either M. albidus or M. viridis to 168 169 containers with prey densities of 2, 4, 8, 15, 30 and 60 (n = 4 per experimental group). 170 Controls consisted of four replicates at each prey density and temperature in the absence of predators. Predators were individually starved for 24 h in containers of the same volume and 171 172 diameter as the experimental arenas before being transferred to containers holding the corresponding prey density. Predators were removed from experimental arenas after 24 173 hours, with the numbers of prey alive counted to derive the numbers killed in each replicate. 174

# 175 2.3. Data manipulation and statistical analyses

Statistical analyses were undertaken in R v3.3.1. (R Core Team, 2016). Logistic
regression of proportion of prey killed as a function of prey density was used to infer FR
types; Type II FRs are characterised by a significant negative first-order term and Type III
FRs by a significant positive first order term followed by a significant negative second order
term. To account for prey depletion, we fitted Rogers' random predator equation for
conditions without prey replacement (Trexler et al. 1988; Juliano, 2001):

182 
$$N_e = N_0 (1 - \exp(a(N_e h - T)))$$

where  $N_e$  is the number of prey eaten,  $N_0$  is the initial density of prey, *a* is the attack rate, *h* is the handling time and *T* is the total experimental period. The Lambert W function was applied due to the implicit nature of the random predator equation (Bolker, 2008). Attack rates and maximum feeding rates '1/*h*' were non-parametrically bootstrapped (*n* = 30) to

(5)

facilitate modelling of FR parameters with respect to 'predator' and 'temperature' factors and their interactions. Bootstrapped parameters were analysed using generalised linear models (GLMs) assuming a quasi-Poisson distribution. *F*-tests were used in a step-deletion process to compare deviance between models (Crawley, 2007). We applied Tukey's HSD method to infer specific pairwise differences using the 'multcomp' package in R (Hothorn et al. 2008).

Benthic survey data for *M. albidus* and *M. viridis* (as *Acanthocyclops viridis*) derived from Tinson and Laybourn-Parry (1986) were used to calculate RCP based on maximum field abundances using pooled bootstrapped mean maximum feeding and attack rates across all three temperatures. To calculate RCP using fecundity, we used results from Laybourn-Parry et al. (1988) to discern the proportion of total consumed energy devoted to reproduction across corresponding temperatures for the two copepods:

199 Fecundity = 
$$\left(\frac{P_r}{C}\right) \times 100$$

200

where  $P_r$  is the quantity of energy expended through the production of eggs and *C* is the total energy consumed at a given temperature (Table 1). Reproductive energy proportions at 16 °C were supplemented with those available for 15 °C. We generated 'RCP biplots' to present the RCP (see Laverty et al. 2017b) of the two predators using both the abundance (AB) and fecundity (FE) proxies for the numerical response.

206 We additionally calculated  $Q_{10}$  values to further quantify the effects of increased 207 temperature on feeding rates and compare how these varied between predatory cyclopoids:

208 
$$Q_{10} = \left(\frac{FR_2}{FR_1}\right)^{\left(\frac{10}{T_2 - T_1}\right)}$$

209

(7)

(6)

where  $Q_{10}$  is a coefficient without units,  $FR_1$  is the maximum feeding rate at temperature  $T_1$ and  $FR_2$  is the maximum feeding rate at temperature  $T_2$ . The  $Q_{10}$  coefficient assesses how temperature increases of 10 °C affect the rate of biological processes (Bennett, 1990); values of 1 – 1.5 are associated with a thermal plateau and values of 2 – 4 indicate substantive increases in activity as temperature increases (Huey, 1982; Bennett, 1990).

215

# 216 **3. Results**

Prey survival in control treatments was a minimum of 98.5% across temperatures, and 217 thus experimental deaths were attributed to predation by copepods, which was also directly 218 observed. Type II FRs were found in all predator and temperature combinations, as indicated 219 by significantly negative first order terms (Table 2; Figure 1). Overall, attack rates (initial FR 220 slopes; see Figure 1) for *M. albidus* were significantly higher than for *M. viridis* ( $F_{1, 178}$  = 221 7.25, p = 0.01) and increased significantly with temperature ( $F_{2,176} = 74.41$ , p < 0.001). There 222 were significant increases in attack rates between 12 °C and 16 °C (z = 5.61, p < 0.001), and 223 12 °C and 20 °C (z = 6.75, p < 0.001), but not between 16 °C and 20 °C (z = 1.20, p = 0.45). 224 There was a significant 'predator × temperature' interaction ( $F_{2, 174} = 3.09, p = 0.05$ ), 225 reflecting significantly greater attack rates by *M. albidus* only at 12 °C (z = 3.42, p = 0.01; 226 Table 2; Figure 1). Overall, maximum feeding rates (asymptotes of FR curves; see Figure 1) 227 did not differ significantly between the two predators ( $F_{1, 178} = 2.88, p = 0.09$ ), but increased 228 significantly with temperature ( $F_{2, 176} = 110.29$ , p < 0.001; Figure 1). There were significant 229 increases in maximum feeding rates between all temperature levels ( $12 \text{ }^{\circ}\text{C} - 16 \text{ }^{\circ}\text{C}, z = 4.23$ , 230 p < 0.001; 16 °C - 20 °C, z = 4.79, p < 0.001; 12 °C - 20 °C, z = 8.81, p < 0.001). There was a 231 significant 'predator × temperature' interaction ( $F_{2, 174} = 3.46$ , p = 0.03), reflecting a non-232 significant difference between maximum feeding rates of M. albidus at 20 °C and M. viridis 233

at 16 °C (z = 2.48, p = 0.13), compared to a significantly lower maximum feeding rate of *M*. *albidus* at 16 °C in comparison to *M*. *viridis* at 20 °C (z = 6.24, p < 0.001).</li>

236	The RCP calculations integrating field abundances with maximum feeding and attack
237	rates are presented in Table 3. M. albidus displays much higher abundance than M. viridis,
238	driving greater RCP using both FR parameters (Figure 2a and 2b). There is a general increase
239	in the proportion of total consumed energy devoted to reproduction as temperature increases
240	(Table 1). Anomalous to this is the response to warming of <i>M. viridis</i> at 20 °C, with fecundity
241	here falling markedly (Table 1). There are relatively similar levels of RCP for the two species
242	illustrated at both 12 °C and 16 °C, followed by a substantial decrease in efficacy of $M$ .
243	viridis at 20 °C (Table 4; Figures 2c and 2d). Indeed, the certainty of the RCP using fecundity
244	strengthens at peak temperatures (Table 4). Under both NR proxies, differential efficacies
245	between predators were more pronounced using the attack rate parameter (Figure 2). The $Q_{10}$
246	coefficients for the two predators across the temperature gradient indicate that between 12 $^{\circ}$ C
247	and 20 °C both the feeding rates of <i>M. albidus</i> and <i>M. viridis</i> were highly responsive to
248	temperature increases. There was a marked difference between the predators in the
249	incremental drivers of this response, with $M$ . viridis exhibiting a rapid increase between 12 °C
250	and 16 °C which slowed between 16 °C and 20 °C. Conversely, M. albidus was consistent in
251	its feeding response to increased temperatures (Table 5).

252

# 253 **4. Discussion**

Biological control of pests, disease vectors and invasive species can be effective (e.g. Hajek, 2007; Nam et al. 2012; Veronesi et al. 2015), but efforts to predict the efficacy of natural enemies are limited when *per capita* effects (e.g. Functional Responses; FRs) are solely considered (Lester and Harmsen, 2002; Fernández-arhex and Corley, 2003). The

complementary agent population response (e.g. Numerical Response; NR) is, however, 258 somewhat nebulous and more difficult to derive, with proxies for the NR required to allow 259 260 rapid assessment of the overall impact of a consumer (i.e. Total Response; TR; Dick et al. 2017). Recent developments that combine functional and numerical responses (or their 261 proxies) into the comparative Relative Impact Potential metric (RIP; Dick et al. 2017) yield 262 high explanatory and predictive power for the ecological impacts of invasive species. Indeed, 263 264 this RIP approach was 100% successful at identifying high impact invasive species, and the metric correlated tightly with degree of ecological impact (i.e. reduction of native species 265 266 populations). Hence, the present RCP metric is promising to assess the efficacy of biocontrol agents, which are chosen on the same basis regarding impact on populations of target species 267 (see Dick et al. 2017). Further value in the application of RCP surrounds the integration of 268 context-dependencies associated with environmental change, which can strongly affect 269 interactions between consumers and their resources (e.g. oxygen availability: Laverty et al. 270 271 2015; habitat complexity: Barrios-O'Neill et al. 2015). Thus, we present the new Relative Control Potential (RCP) metric that uses *per capita* and consumer population responses to 272 compare efficacy among biocontrol agents and can allow predictions of changes in such 273 efficacies under context-dependencies. 274

The risk of mosquito-borne disease at continental scales has reached unprecedented 275 276 levels in recent decades (Medlock and Leach, 2015). Arboviruses such as Zika, West Nile, dengue and chikungunya present enormous public health concerns, with disease dynamics 277 shifting rapidly under environmental change (Benelli and Mehlhorn, 2016; Siraj et al. 2017). 278 279 Agricultural systems will additionally be impacted (Chevalier, 2013). This risk necessitates the formation of techniques to assess and compare the potential efficacies of biological 280 control agents. Here, RCP revealed M. albidus as a more effective agent than M. viridis. 281 282 Temperature was shown to mediate changes to the FR parameters of *M. albidus* and *M.* 

*viridis*, driving higher magnitude FRs through increasing attack rates and decreasing handling 283 times. The temperature-dependence of attack rates reported here contrasts to suggestions that 284 285 this FR parameter is temperature-independent (Rall et al. 2012; Dell et al. 2014). We show that both predators exhibit high maximum feeding rates that exceed 30 of the West Nile virus 286 vector C. pipiens per day at 20 °C. Critically, the Type II FRs found are indicative of a 287 capacity to destabilise prey populations due to high proportional consumption at low prey 288 289 densities (Long and Whitefleet-Smith, 2013). M. albidus and M. viridis show strong 290 similarities in their *per capita* consumption, although the attack rates of *M. albidus* were 291 significantly greater overall, illustrated by steeper gradients in the FR curves at low densities. As a result, *M. albidus* may be more effective in controlling *C. pipiens* populations. 292

293 We demonstrate that integrating field abundances with RCP reveals far stronger control efficacies of *M. albidus* compared to *M. viridis*. The utility of abundance estimates 294 295 lies in the projection of how many conspecifics may engage in the predator-prey (or other 296 consumer-resource) interaction and it is thus a useful NR proxy. On the other hand, incorporating measures of fecundity estimates how rapidly biological control agents can 297 reproduce when introduced. Using fecundity, we demonstrate temperature-dependencies of 298 control efficacy. Large fecundity variabilities are illustrated in the RCP biplots, with the 299 reproductive allocation of *M. viridis* declining rapidly at 20 °C, whilst that of *M. albidus* 300 301 continues to rise. The reduction in fecundity shown by M. viridis at 20 °C is concurrent with a slowing feeding activity response discerned through  $Q_{10}$  analysis, whereas M. albidus 302 displays a consistent incremental increase in feeding with warming. Overall, M. albidus 303 304 displays greater potential for the control of West Nile virus vector C. pipiens than M. viridis, particularly under climate change projections where our certainty for differential efficacy 305 increases (e.g. Hulme et al. 2002). This differential efficacy is more pronounced when the 306 307 attack rate parameter is considered. An increased metabolic demand could enable M. albidus

to sustain efficiency in the consumption of *C. pipiens*, which proliferate more rapidly at
higher temperatures, particularly in domestic and peri-domestic habitats in urbanised
environments (Townroe and Callaghan, 2014).

In this study, temperature had a profound effect on predator-prev interactions. It has 311 been suggested that interaction strengths of ambush predators such as the benthic copepods 312 313 examined here are temperature independent (Awasthi et al. 2012; Novich et al. 2014). Thus, 314 marked thermal dependencies may result, rather, from individual prey foraging responses to temperature change, wherein motility and velocity may increase with warming. These 315 interactions make predator-prey systems highly specific to both the species (Englund et al. 316 2011) and environment (Broitman et al. 2011), with optimal foraging patterns of ectothermic 317 prey potentially peaking at intermediate temperatures (Englund et al. 2011; Kalinoski and 318 DeLong 2016). Further, for endotherms, such interactions are often dependent on the specific 319 feeding strategy (Dell et al. 2014). In this study, feeding rates of candidate biocontrol agents 320 321 peaked at the highest temperature, however the rate of increase slowed between 16 °C and 20  $^{\circ}$ C, as compared to between 12  $^{\circ}$ C and 16  $^{\circ}$ C; this trend was particularly marked for *M*. 322 viridis. Notably, these temperatures are well within the thermal tolerances of common 323 cyclopoid copepods (Marten and Reid, 2007). Temperature additionally has a substantial 324 influence on the development of the focal prey, C. pipiens, driving significant reductions in 325 326 development times under conditions of warming (Loetti et al. 2011; Ruybal et al. 2016), and necessitating increased foraging intensity. Yet, increases in C. pipiens mortality due to 327 drivers outside of predation are also evident as temperatures rise due to thermal stressors 328 329 (Ruybal et al. 2016). M. albidus has been proved particularly effective against the invasive arbovirus vector A. albopictus following field trials (Marten, 1990; Veronesi et al. 2015). 330 Previous research has suggested that copepods are more efficient consumers of Aedes spp. 331 332 than *Culex* spp. as a result of morphological variations between the genera (Marten and Reid, 2007). However, laboratory trials have shown similar levels of overall predation by *M. albidus* towards both *A. albopictus* and *C. pipiens* (Veronesi et al. 2015), as well as by *Mesocyclops annulatus* (Micieli et al. 2002). Further field trials are required to elucidate
whether their efficiencies towards *C. pipiens* translate empirically, particularly as it is the
major West Nile virus vector in the USA and Europe (Hubalek and Halouzka, 1999; Fonseca
et al. 2004) and part of one of the most widespread mosquito complexes in the world
(Harbach, 2012).

In conclusion, this is the first study to develop and apply the new RCP metric to 340 biological control agent selection. We demonstrate that the integration of abundance and 341 fecundity estimations can provide a means to differentiate between biocontrol agents that 342 343 display similar *per capita* efficacies across temperature gradients. We additionally illustrate the value in the use of maximum feeding and attack rates for instructing agent selection. 344 Adaptations of this metric have been applied successfully in the context of invasion biology 345 346 to explain and predict the ecological impact of invasive species (Dick et al. 2017; Laverty et al. 2017b), and similar fundamental principles enable its application to the selection of 347 biocontrol agents. Overall, temperature increases will induce greater per capita predation 348 pressure by predatory copepods towards C. pipiens and likely other mosquito species. 349 Copepod applications to waterbodies can form an integral part of mosquito control efforts 350 351 (Baldacchino et al. 2015), with large-scale field trials having proved successful (e.g. Kay and Nam, 2005), particularly given their ability to thrive in both natural and artificial waterbodies 352 (Marten and Reid, 2007). Importantly, copepods can be augmented synergistically using 353 354 existing control methods, such as the use of bacterial Bacillus thuringiensis var. israelensis (Bti; Kosiyachinda et al. 2003). Nanoparticles have additionally been found to heighten 355 predation (Murugan et al. 2015). The straightforward derivation of the RCP metric, and its 356 357 visual representation in biplots, will allow comparisons of biological control agents across

many ecological systems, and could increase cost-effectiveness of natural enemies in the
long-term. Further proxies for the numerical response, such as biocontrol agent longevity or
biomass, can be integrated into the RCP metric as per the requirements of the assessed system
or biocontrol approach, increasing the robustness and flexibility of the method. Moreover, the
additional integration of a qualifier to account for target organism responses under matched
environmental change scenarios could bolster the power of the RCP metric in reliably
selecting biocontrol agents, and this is worth further consideration in future research.

365

# 366 Authors' contributions

Relative Control Potential (RCP) concept and framework: RNC, JTAD and JWED. Design
and execution of experimental trials: RNC and JTAD. Fitting functional response models and
undertaking RCP calculations: RNC. First draft of the manuscript: RNC. Manuscript

370 revisions: all authors.

371

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377

# 378 **References**

Alexander, M.E., Dick, J.T.A., O'Connor, N.E., 2013. Trait-mediated indirect interactions in

380 <u>a marine intertidal system as quantified by functional responses. Oikos 122(11), 1521–</u>

381 <u>1531.</u>

- Awasthi, A.K., Wu, C-W., Tsai, K-H., King, C-C., Hwang, J-S., 2012. How does the ambush
   predatory copepod *Megacyclops formosanus* (Harada, 1931) capture mosquito larvae of
- 384 *Aedes aegypti*? Zool. Stud. 51(7), 927–936.
- 385 Baldacchino, F., Caputo, B., Chandre, F., Drago, A., della Torre, A., Montarsi, F., Rizzoli,
- 386 <u>A., 2015. Control methods against invasive Aedes mosquitoes in Europe: A review. Pest</u>
  387 Manage. Sci. 71(11), 1471–1485.
- 388 Baldacchino, F. Bruno, M.C., Visentin, P., Blondel, K., Arnolid, D., Hauffe, H.C., Rizzoli,
- 389 <u>A., 2017. Predation efficiency of copepods against the new invasive mosquito species</u>
- 390 *Aedes koreicus* (Diptera: Culicidae) in Italy. Eur. Zool. J. 84(1), 43–48.
- 391 Barrios-O'Neill, D., Dick, J.T.A., Emmerson, M.C., Ricciardi, A., MacIsaac, H.J., Alexander,
- 392 M.E., Bovy, H.C., 2014. Fortune favours the bold: A higher predator reduces the impact
- 393 of a native but not an invasive intermediate predator. J. Anim. Ecol. 83(3), 693–701.
- Barrios-O'Neill, D., Dick, J.T.A., Emmerson, M.C., Ricciardi, A., MacIsaac, H., 2015.
- 395 Predator-free space, functional responses and biological invasions. Funct. Ecol. 29(3),
- <u>396</u> <u>377–384.</u>
- Benelli, G., Mehlhorn, H., 2016. Declining malaria, rising of dengue and Zika virus: insights
  for mosquito vector control. Parasitol. Res. 115(5), 1747-1754.
- Bennett, A.F., 1990. Thermal dependence of locomotor capacity. Am. J. Physiol. 259(2),
  253–258.
- Bolker, B.M., 2008. Ecological Models and Data in R. Princeton University Press, Princeton.
- 402 Broitman, B.R., Szathmary, P.L., Mislan, K.A.S., Blanchette, C.A., Helmuth, B., 2009.
- 403 Predator-prey interactions under climate change: the importance of habitat vs body

- 404 <u>temperature. Oikos 118(2), 219–224.</u>
- 405 Bunke, M., Alexander, M.E., Dick, J.T.A., Hatcher, M.J., Paterson, R., Dunn, A.M., 2015.
- 406 Eaten alive: cannibalism is enhanced by parasites. Open Sci. 2, 140369.
- 407 Calvo, F.J., Torres-Ruiz., A., Velázquez-Gonzále, J.C., Rodríguez-Leyva, E., Lomeli-Flores,
- 408 J.R., 2016. Evaluation of *Dicyphus hesperus* for biological control of sweet potato
- 409 whitefly and potato psyllid on greenhouse tomato. Biocontrol 61(4), 415-424.
- 410 Chevalier, V., 2013. Relevance of Rift Valley fever to public health in the European Union.
- 411 <u>Clin. Microbiol. Infect. 19(8), 705-708.</u>
- 412 Costa, J.F., Matos, C.H.C., de Oliveira, C.R.F., da Silva, T.G.F., Lima Neto, I.F.A., 2017.
- 413 <u>Functional and numerical responses of *Stethorus tridens* Gordon (Coleoptera:</u>
- 414 <u>Coccinellidae) preying on *Tetranychus bastosi* Tuttle, Baker & Sales (Acari:</u>
- 415 <u>Tetranychidae) on physic nut (*Jatropha curcas*). Biol. Control 111, 1-5.</u>
- 416 Crawley, M.J., 2007. The R book. John Wiley & Sons, Chichester.
- 417 Dell, A.I., Pawar, S., Savage, V.M., 2014. Temperature dependence of trophic interactions
- 418 <u>are driven by asymmetry of species responses and foraging strategy. J. Anim. Ecol.</u>
- 419 <u>83(1), 70–84.</u>
- 420 Dick, J.T.A., Alexander, M.E., Jeschke, J.M., Ricciardi, A., MacIsaac, H.J., Robinson, T.B.,
- 421 Kumschick, S., Weyl, O.L.F., Dunn, A.M., Hatcher, M.J., Paterson, R.A., Farnsworth,
- 422 K.D., Richardson, D.M., 2014. Advancing impact prediction and hypothesis testing in
- 423 invasion ecology using a comparative functional response approach. Biol. Invasions
- 424 <u>16(4)</u>, 735–753.
- 425 Dick, J.T.A., Laverty, C., Lennon, J.J., Barrios-O'Neill., D., Mensink, P.J., Britton, R.,
- 426 Médoc, V., Boets, P., Alexander, M.E., Taylor, N.G., Dunn, A.M., Hatcher, M.J.,

- 427 Rosewarne, P.J., Crookes, S., MacIsaac, H.J., Xu, M., Ricciardi, A., Wasserman, R.J.,
- 428 Ellender, B.R., Weyl, O.L.F., Lucy, F.E., Banks, P.B., Dodd, J.A., MacNeil, C., Penk,
- 429 M.R., Aldridge, D.C., Caffrey, J.M., 2017. Invader Relative Impact Potential: a new
- 430 metric to understand and predict the ecological impacts of existing, emerging and future
- 431 invasive alien species. J. Appl. Ecol. 54(4), 1259–1267.
- 432 Englund, G., Ohlund, G., Hein, C.L., Diehl, S., 2011. Temperature dependence of the
- 433 <u>functional response. Ecol. Lett. 14(9), 914–921.</u>
- 434 Fernández-arhex, V., Corley, J.C., 2003. The functional response of parasitoids and its
- 435 <u>implications for biological control. Biocontrol Sci. Technol. 13(4), 403–413.</u>
- 436 Fonseca, D.M., Keyghobadi, N., Malcolm, C.A., Mehmet, C., Schaffner, F., Mogi, M.,
- 437 Fleischer, R.C., Wilkerson, R.C., 2004. Emerging vectors in the *Culex pipiens* complex.
- 438 <u>Sci. 303(5663), 1535–1538.</u>
- 439 Hajek, A., 2007. Introduction of a fungus into North America for control of gypsy moth, in:
- 440 Vincent, C., Goettel, M.S., Lazarovits, G. (Eds.) Biological control: a global perspective.
- 441 <u>CAB International, Wallingford, pp. 53–62.</u>
- Hammond, D., Pryce, A.R., 2007. Climate change impacts and water temperature.
- 443 Environment Agency Science Report SC060017/SR. Bristol.
- 444 Harbach, R.E., 2012. Culex pipiens: Species versus species complex taxonomic history and
- 445 perspective. J. Am. Mosq. Control Assoc. 28(4), 10–23.
- 446 Heisswolf, A., Klemola, N., Ammunét, T., Klemola, T., 2009. Responses of generalist
- 447 <u>invertebrate predators to pupal densities of autumnal and winter moths under field</u>
- 448 <u>conditions. Ecol. Entomol. 34(6), 709–717.</u>
- 449 Holling, C.S., 1959. Some characteristics of simple types of predation and parasitism. Can.

#### 450 <u>Entomol. 91(7), 385–398.</u>

- 451 Hothorn, T., Bretz, F., Westfall, P., 2008. Simultaneous inference in general parametric
- 452 <u>models. Biom. J. 50(3)</u>, 346–363.
- 453 Hubalek, Z., Halouzka, J., 1999. West Nile fever a remerging mosquito-borne viral disease
- 454 <u>in Europe. Emerg. Infect. Dis. 5(5), 643–650.</u>
- 455 <u>Huey, R.B., 1982. Temperature, physiology, and the ecology of reptiles, in: Gans, C., Pough,</u>
  456 F.H. (Eds.), Biology of the Reptilia. Academic Press, New York, pp. 25–91.
- 457 Hulme, M., Jenkins, G.J., Lu, X., Turnpenny, J.R., Mitchell, T.D., Jones, R.G., Lowe, J.,
- 458 Murphy, J.M., Hassell, D., Boorman, P., McDonald, R., Hill, S., 2002. Climate change
- 459 scenarios for the United Kingdom. The UKCIP Scientific Report. Tyndall Centre for
- 460 Climate Change Research, School of Environmental Sciences, University of East
- 461 Anglia, Norwich.
- 462 Juliano, S.A., 2001. Non-linear curve fitting: predation and functional response curves, in:
- 463 Scheiner, S.M., Gurevitch, J. (Eds.), Design and Analysis of Ecological Experiments.
- 464 Oxford University Press, Oxford, pp. 178–196.
- 465 <u>Kalinoski, R.M., DeLong, J.P., 2016. Beyond body mass: how prey traits improve predictions</u>
  466 <u>of functional response parameters. Oecologia 180(2), 543–550.</u>
- 467 Kay, B.H., V.S. Nam., 2005. New strategy against Aedes aegypti in Vietnam. Lancet
- **468** <u>365(9459), 613-617.</u>
- 469 Kosiyachinda, P., Bhumiratana, A., Kittayapong, P., 2003. Enhancement of the efficacy of a
- 470 combination of *Mesocyclops aspericornis* and *Bacillus thuringiensis* var. *israelensis* by
- 471 community-based products in controlling *Aedes aegypti* larvae in Thailand. Amer. J.
- 472 Trop. Med. Hyg. 69(2), 206–212.

- 473 Laverty C., Dick, J.T.A., Alexander, M.E., Lucy, F.E., 2015. Differential ecological impacts
- 474 of invader and native predatory freshwater amphipods under environmental change are
- 475 revealed by comparative functional responses. Biol. Invasions 17(6), 1761–1770.
- 476 Laverty, C., Brenner, D., McIlwaine, C., Lennon, J.J., Dick, J.T.A., Lucy, F.E., Christian,
- 477 <u>K.A., 2017a. Temperature rise and parasitic infection interact to increase the impact of</u>
- 478 <u>an invasive species. Int. J. Parasitol. 47(5), 291–296.</u>
- 479 Laverty, C., Green, K.D., Dick, J.T.A., Barrios-O'Neill, D., Mensink, P.J., Médoc, V.,
- 480 Spataro, T., Caffrey, J.M., Lucy, F.E., Boets, P., Britton, J.R., Pegg, J., Callagher, C.,
- 481 <u>2017b. Assessing the ecological impacts of invasive species based on their functional</u>
- 482 responses and abundances. Biol. Invasions 19(5),1653–1665.
- 483 Laybourn-Parry, J., Abdullahi, B.A., Tinson, S.V., 1988. Temperature-dependent energy
- 484 partitioning in the benthic copepods *Acanthocyclops viridis* and *Macrocyclops albidus*.
- 485 <u>Can. J. Zool. 66(12), 2709–2713.</u>
- 486 Lester, P.J., Harmsen, R., 2002. Functional and numerical responses do not always indicate
- 487 the most effective predator for biological control: An analysis of two predators in a two-
- 488 prey system. J. Appl. Ecol. 39(3), 455–468.
- 489 Loetti, V., Schweigmanna, N., Burronia, N., 2011. Development rates, larval survivorship
- 490 and wing length of *Culex pipiens* (Diptera: Culicidae) at constant temperatures. J. Nat.
- 491 <u>Hist. 45(36-36), 2207–2217.</u>
- 492 Long, W.C., Whitefleet-Smith, L., 2013. Cannibalism in red king crab: habitat, ontogeny, and
- 493 the predator functional response. J. Exp. Mar. Biol. Ecol. 449, 142–148.
- 494 Marten, G.G., 1990. Elimination of *Aedes albopictus* from tire piles by introducing
- 495 *Macrocyclops albidus* (Copepoda, Cyclopidae). J. Am. Mosq. Control Assoc. 6(4), 689–

496 693.

- 497 <u>Marten, G.G., Reid, J.W., 2007. Cyclopoid copepods. J. Am. Mosq. Control Assoc. 23(2),</u>
  498 <u>65–92.</u>
- 499 <u>Medlock, J.M., Leach, S.A., 2015. Effect of climate change on vector-borne disease risk in</u>
- 500 the UK. Lancet Infect. Dis. 15(6), 721–730.
- 501 Medlock, J.M., Vaux, A.G.C., Cull, B., Schäffner, F., Gillingham, E., Pfluger, V., Leach, S.,
- 502 2017. Detection of the invasive mosquito species *Aedes albopictus* in southern England.
- 503
   Lancet Infect. Dis. 17(2), 140.
- 504 Micieli, M.V., Marti, G., García, J.J., 2002. Laboratory evaluation of *Mesocyclops annulatus*
- 505 (Wierzejski, 1892) (Copepoda: Cyclopidea) as a predator of container-breeding
- 506 mosquitoes in Argentina. Mem. Inst. Oswaldo Cruz. 97(6), 835–838.
- 507 Murugan, K., Benelli, G., Panneerselvam, C., Subramaniam, J., Jeyalalitha, T., Dinesh, D.,
- 508 Nicoletti, M., Hwang, J.S., Suresh, U., Madhiyazhagan, P., 2015. Cymbopogon citratus-
- 509 synthesized gold nanoparticles boost the predation efficiency of copepod *Mesocyclops*
- 510 *aspericornis* against malaria and dengue mosquitoes. Exp. Parasitol. 153, 129–138.
- 511 Nam, V.S., Yen, N.T., Duc, H.M., Tu, T.C., Thang, V.T., Le, N.H., Le Loan, L., Huong,
- 512 V.T.Q., Khanh, L.H.K., Trang, H.T.T., Lam, L.Z.Y., Kutcher, S.C., Aaskov, J.G.,
- 513 Jeffery, J.A.L., Ryan, P.A., Kay, B.H., 2012. Community-based control of Aedes aegypti
- 514 by using *Mesocyclops* in Southern Vietnam. Am. J. Trop. Med. Hyg. 86(5), 850–859.
- 515 Novich, R.A., Erickson, E.K., Kalinoski, R.M., DeLong, J.P., 2014. The temperature
- 516 <u>independence of interaction strength in a sit-and-wait predator. Ecosphere 5(10), 1–19.</u>
- 517 O'Neil, R.J., 1990. Functional response of arthropod predators and its role in the biological
- 518 control of insect pests in agricultural systems, in: Dunn, P.E., Baker, R.R. (Eds.) New

- 519 directions in biological control: alternatives for suppressing agricultural pests and
- 520 diseases. Alan R. Liss, Inc., New York, pp. 83–96.
- 521 R Core Team, 2016. R: A language and environment for statistical computing. R Foundation
- 522 for Statistical Computing, Vienna.
- 523 Rall, B.C., Brose, U., Hartvig, M., Kalinkat, G., Schwarzmüller, F., Vucic-Pestic, O.,
- 524 Petchey, O.L., 2012. Universal temperature and body-mass scaling of feeding rates.
- 525 Philos. Trans. R. Soc. Lond., Series B Biol. Sci. 367(1605), 2923–2934.
- 526 Ruybal, J.E., Kramer, L.D., Kilpatrick, A.M., 2016. Geographic variation in the response of
- 527 *Culex pipiens* life history traits to temperature. Parasit. Vectors 9(116), 1–9.
- 528 Siraj, A.S., Oidtman, R.J., Huber, J.H., Kraemer, M.U.G., Brady, O.J., Johansson, M.A.,
- 529 Perkins, T.A., 2017. Temperature modulates dengue virus epidemic growth rates
- 530 through its effects on reproduction numbers and generation intervals. PLOS Negl.
- 531 <u>Trop. Dis. 11(7), e0005797.</u>
- 532 Solomon, M.E., 1949. The natural control of animal populations. J. Anim. Ecol. 18, 1–35.
- 533 South, J., Dick, J.T.A., McCard, M., Barrios-O'Neill, D., Anton, A., 2017. Predicting
- 534 predatory impact of juvenile invasive lionfish (*Pterois volitans*) on a crustacean prey
- 535 <u>using functional response analysis: effects of temperature, habitat complexity and light</u>
- 536 regimes. Environ. Biol. Fish. 100(10), 1155–1165.
- 537 <u>Tinson, S., Laybourn-Parry, J., 1986. The distribution and abundance of benthic cyclopoid</u>
- 538 <u>copepods in Esthwaite Water, Cumbria. Hydrobiol. 131(3), 225–234.</u>
- 539 Toscano, B.J., Rombado, B.R., Rudolf, V.H.W., 2016. Deadly competition and life-saving
- 540 predation: the potential for alternative stable states in a stage-structured predator-prey
- 541 system. Proc. R. Soc. B. 283, 20161546.

- 542 <u>Townroe, S., Callaghan, A., 2014. British container breeding mosquitoes: The impact of</u>
- 543 urbanisation and climate change on community composition and phenology. PLoS
  544 ONE 9(4), e95325.
- 545 Trexler, J.C., McCulloch, C.E., Travis, J., 1988. How can the functional response best be
- 546 <u>determined? Oecologia 76(2), 206–214.</u>
- 547 Van Driesche, R., Bellows, T.S., 2011. Biological control. Springer, Berlin.
- 548 Veronesi, R., Carrieri, M., Maccagnani, B., Maini, S., Bellini, R. 2015. Macrocyclops albidus
- 549 (Copepoda: Cyclopidae) for the biocontrol of Aedes albopictus and Culex pipiens in
- 550 <u>Italy. J. Am. Mosq. Control Assoc. 31(1), 32–43.</u>
- 551 Wasserman, R.J., Alexander, M.E., Weyl, O.L.F., Barrios-O'Neill, D., Froneman, P.W., Dalu,
- 552 <u>T., 2016. Emergent effects of structural complexity and temperature on predator-prey</u>
- 553 <u>interactions. Ecosphere 7(2), e01239.</u>
- 554 Yee, D.A., 2016. Thirty years of Aedes albopictus (Diptera: Culicidae) in America: An
- 555 introduction to current perspectives and future challenges. J. Med. Entomol. 53(5), 989–
- 556 <u>991.</u>
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# 564 Tables

# 565 Table 1

	Species	Temperature (°C)	<i>C</i> (mJ)	$P_r$ (mJ)	Fecundity (%)
	M. albidus	12	31210	2355	7.55
	M. albidus	16	26150	2907	11.12
	M. albidus	20	29150	3691	12.66
	M. viridis	12	34433	2851	8.28
	M. viridis	16	25311	3020	11.93
	M. viridis	20	24960	1671	6.70
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575 Table 2

	Species	Temperature (°C)	1 <sup>st</sup> order term	р	Attack rate ( <i>a</i> )	р	Handling time ( <i>h</i> )	р
	M. albidus	12	-0.02	< 0.001	1.28	< 0.001	0.07	< 0.001
	M. albidus	16	-0.03	< 0.001	1.92	< 0.001	0.04	< 0.001
	M. albidus	20	-0.04	< 0.001	2.37	< 0.001	0.04	< 0.001
	M. viridis	12	-0.03	< 0.001	0.95	< 0.001	0.07	< 0.001
	M. viridis	16	-0.03	< 0.001	1.90	< 0.001	0.04	< 0.001
	M. viridis	20	-0.04	< 0.001	2.19	< 0.001	0.03	< 0.001
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Agent A, agent B comparison	Mean FR parameter (± SD)	Mean abundance (ind. $m^{-2} \pm SD$ )	RCP	CIs $p_{\text{RCP}} > 1 (\%)$
M. albidus, M. viridis	1/ <i>h</i> : 22.80 (± 8.37), 24.41 (± 10.55)	6727 (± 1018.23), 562 (± 288.50)	16.77	6.79 – 23.78 99.97***
M. albidus, M. viridis	<i>a</i> : 1.98 (± 0.72), 1.77 (± 0.67)	6727 (± 1018.23), 562 (± 288.50)	19.34	8.18 – 27.35 99.99***

Agent A, agent B comparison	Temperature (°C)	Mean FR parameter (± SD)	Fecundity (%)	RCP	CIs <i>p</i> <sub>RCP</sub> >1 (%)
M. albidus, M. viridis	12	1/ <i>h</i> : 15.88 (± 4.36), 13.99 (± 2.67)	7.55, 8.28	1.07	0.77 – 1.34 51.93
M. albidus, M. viridis	16	1/ <i>h</i> : 22.12 (± 7.64), 25.98 (± 9.68)	11.12, 11.93	0.90	0.53 – 1.21 32.58
M. albidus, M. viridis	20	1/ <i>h</i> : 30.42 (± 5.24), 33.25 (± 6.73)	12.66, 6.70	1.80	1.39 – 2.17 98.21*
M. albidus, M. viridis	12	<i>a</i> : 1.43 (± 0.52), 1.05 (± 0.34)	7.55, 8.28	1.37	0.82 – 1.83 66.71
M. albidus, M. viridis	16	a: 2.17 (± 0.51), 2.02 (± 0.39)	11.12, 11.93	1.04	0.77 – 1.28 49.04
M. albidus, M. viridis	20	<i>a</i> : 2.34 (± 0.75), 2.24 (± 0.54)	12.66, 6.70	2.09	1.39 – 2.69 95.34*

	Species	Temperature (°C)	Mean max. feeding rate $(1/h)$	$Q_{10}$ value
	M. albidus	12 – 20	15.88 - 30.42	2.25
	M. albidus	12 – 16	15.88 - 22.12	2.29
	M. albidus	16 – 20	22.12 - 30.42	2.22
	M. viridis	12 – 20	13.99 – 33.25	2.95
	M. viridis	12 – 16	13.99 – 25.98	4.70
	M. viridis	16 – 20	25.98 - 33.25	1.85
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#### 622 **Captions**

Table 1. Fecundity (%) calculations, entailing proportion of total consumed energy (C)

624 devoted to production of eggs  $(P_r)$  in adult female *M. albidus* and *M. viridis* across 12 °C, 16

625 °C and 20 °C. Data relating to energetics adapted from Laybourn-Parry et al. (1988; n = 5;

626 Eqn. 6).

627 Table 2. Results of logistic regression to denote functional response type across all predator

628 and temperature treatments, alongside the starting attack rate (a) and handling time (h)

629 parameter estimates and associated *p*-values generated using the Rogers' random predator



Table 3: Mean Relative Control Potential (RCP) using maximal abundances from Tinson and

632 Laybourn-Parry (1986; n = 8) for *M. albidus* and *M. viridis*, alongside uncertainties reflected

through 60% confidence intervals (CIs) and probability that the RCP output exceeds 1 using

bootstrapped maximum feeding and attack rates. Asterisks denote significant levels of

635 certainty that the RCP score is greater than 1 (\* > 95%, \*\* > 99%, \*\*\* > 99.9%).

Table 4: Mean Relative Control Potential (RCP) using fecundities (%) across temperature

637 change for *M. albidus* and *M. viridis* alongside uncertainties reflected through 60%

638 confidence intervals (CIs) and probability that the RCP output exceeds 1 using bootstrapped

639 maximum feeding and attack rates. Asterisks denote significant levels of certainty that the

640 RCP score is greater than 1 (\* > 95%, \*\* > 99%, \*\*\* > 99.9%).

Table 5:  $Q_{10}$  coefficient (Eqn. 7) values associated with mean bootstrapped maximum feeding rates for both predators between temperature gradients.

643 Figure 1. Functional responses of *M. albidus* (a, b, c) and *M. viridis* (d, e, f) towards first

644 instar *C. pipiens* larvae at 12 °C (a, d), 16 °C (b, e) and 20 °C (c, f) over the 24 hour

645 experimental period. Means are  $\pm$  SE at each prey density (n = 4).

646	Figure 2. RCP biplots comparing <i>M. albidus</i> and <i>M. viridis</i> using abundance estimates (a, b; n
647	= 8) and fecundity calculations (c, d; Table 1), with FR parameters of maximum feeding (a,
648	c) and attack rates (b, d). FR parameters in abundance biplots are pooled bootstrapped
649	estimates across all temperatures ( $n = 90$ ); those in fecundity plots are temperature-specific
650	estimates ( $n = 30$ ). Increasing CP is read from bottom left to top right. Abundance and FR
651	parameter means are $\pm$ SE.
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