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Fibre Laser Treatment of Martensitic NiTi Alloys for Load-bearing Implant Applications: Effects of Surface Chemistry on Inhibiting the *Staphylococcus aureus* Biofilm Formation

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Abstract

Biofilm infection is one of the main reasons for implant failure. It is extremely difficult to cure due to its high resistance to antibiotic treatments, and can result in substantial healthcare costs. In this study, the important shape memory NiTi alloy, in its martensitic state, was laser-treated using our newly-developed surface modification technique, aiming to tackle the biofilm infection problem. Martensitic NiTi was chosen for investigation because of its potential advantages in terms of (i) lower elastic modulus and (ii) higher damping capacity over its austenitic counterpart, giving rise to a lower risk of stress shielding and maximum stress between bones and load-bearing implants. The surfaces after laser treatment were systemically analysed using a series of surface measurement (i.e. surface roughness and water contact angle) and material characterisation (i.e. SEM-EDX, XRD and XPS) techniques. The antibacterial performance of the laser-treated surfaces was evaluated using the *Staphylococcus aureus* (or *S. aureus*) cells *in-vitro* cultured at 37 °C for 24h. Fluorescence microscopy accompanied by Live/Dead staining was employed to analyse the cell culture results. The surfaces in their as-received states and after polishing were also tested and compared with the laser-treated surfaces in order to gain a deeper insight in how different surface conditions would influence biofilm formation. Our results indicate that the surfaces after laser treatment can mitigate bacterial attachment and biofilm formation effectively. The antibacterial performance was mainly attributable to the laser-formed oxides which brought desirable changes to the surface chemistry of NiTi. The laser-induced changes in surface roughness and topography, on a micrometre scale, only played a minor role in influencing bacterial attachment. The findings of this study demonstrated for the first time that martensitic NiTi with laser treatment could be a promising choice for the next-generation implants given its superior antimicrobial resistance and favourable mechanical properties for loading bearing applications.

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1. Introduction

The unusual shape memory effect (SME) and super-elasticity (SE) coupled with good corrosion resistance, magnetic resonance compatibility and biocompatibility have made the near-equiatomic nickel titanium (NiTi) alloy a very popular choice in the medical device industry in the last decade. A medical devices market research report has shown that the global NiTi medical devices market for final medical components (namely stents, guidewires and others) was valued at US\$ 8.2 billion in 2012 whilst the global market for the NiTi semi-finished goods (namely tubes, wiring, sheets and ribbons) was amounted to US\$ 1.5 billion in 2012 [1]. The biomedical applications of NiTi span from the fields of orthodontic to orthopaedic, and vascular to neurosurgical applications [2,3].

Both the SME and SE are direct consequences of the reversible martensitic transformation between Austenite (a high-temperature phase) and Martensite (a low-temperature phase) of NiTi [4]. There are four important temperatures associated with the martensitic transformation process, namely A_s (Austenite start), A_f (Austenite finish), M_s (Martensite start) and M_f (Martensite finish) temperatures. SME is a thermal-induced transformation process which refers to the ability of a deformed martensitic NiTi (i.e. undergoes deformation at $T < M_f$) to restore to its original shape in the austenitic state upon heating (at $T > A_f$), whilst SE is a stress-induced martensitic transformation process (i.e. keeps the material at $T > A_f$ throughout the process), allowing the austenitic NiTi to sustain up to 8 % strain without plastic deformation [4]. The majority of the NiTi biomedical applications stem from utilising the SME (i.e. used in the martensitic phase at room temperature but transforms to austenite around the body temperature of 37 °C) or SE (i.e. used in the austenitic phase at 37 °C) [2,3]. In other words, NiTi medical devices are used in the austenitic state at the human body temperature to actuate either the SME or SE.

Interestingly, NiTi in its martensitic state possess a desirable characteristic of very low elastic modulus (between 26-48 GPa)[5] which is much lower than that in its austenitic state (about 83 GPa)[5] and much closer to that of the cortical bone (between 19 to 21 GPa)[6]. The Martensite of NiTi can be thermally-induced or stress-induced. The martensitic NiTi mentioned herein refers to the thermally-induced Martensite. Martensitic and austenitic NiTi possess completely different mechanical transformation behaviours. Along with biocompatibility and osseointegration as well as wear and corrosion resistance, elastic modulus is one of the most important selection criteria of load-bearing implants in bone plates for internal fixation. Large mismatching of elastic modulus between the implant material and the adjacent cortical bone (i.e. particularly if the material is stiffer than the bone, then the implant will bear more of the load) can contribute to “stress shielding effect”, which causes the subsequent bone resorption and aseptic loosening (which accounts for nearly 30 % of implant failure rate for revision)[7]. In addition, martensitic NiTi is well known for its high damping capacity [8]. The lower elastic modulus and higher damping capacity would result in (i) lower risk of stress shielding, (ii) more absorption of impact energy and (iii) lower maximum stress between the bones and implant [9]. Higher resistance to low-cycle fatigue [10] and comparable corrosion resistance to austenitic NiTi [11] are the additional benefits of martensitic NiTi. From the manufacturing point of view, martensitic NiTi is advantageous over austenitic NiTi because of ease in shape forming and machining. However, existing research in martensitic NiTi is significantly less than its austenitic counterpart. This paper serves as one of the first initiatives to explore and report the possibility of using martensitic NiTi in biomedical applications.

The high nickel (Ni) content in NiTi poses a risk of the release of harmful Ni ions into body fluids and tissues in contact. It is known that the outermost Ti oxide layer on the NiTi surface acts as a barrier to confine the diffusion/release of Ni ions [12,13]. The quality and protectiveness of the oxide layer are controlled by several attributes: composition, oxide species, oxide uniformity and homogeneity [14]. These attributes of the oxide layer depend on the previous manufacturing history (e.g. mechanical polishing, chemical passivation or etching, and heat-treatment)[13,15] and can be varied by post-process surface treatments (e.g. ion implantation [16,17], thermal oxidation [18,19] and laser treatment (LT)[15,20-25].

Among the aforementioned post-process techniques, LT including laser surface melting, nitriding and oxidation techniques offer the promising capabilities of precisely tailoring the surface characteristics (namely microstructure, chemistry, roughness and topography) in local areas as well as being accurate, repeatable and clean (a non-contact process). Positive effects of LT to reduce Ni release have been documented in

literature. The improvements mainly stem from the following two characteristics related to the laser-formed oxide layer at the outermost surface: (i) increased percentage (at.%) of TiO₂ in the oxide layer [23] and (ii) enhanced oxide homogeneity/uniformity as a result of surface homogenisation after LT (i.e. removal of the pre-existing surface inclusions and/or Ni-rich regions in the base material [15]). Moreover, the other beneficial characteristics of the laser-formed oxide layer such as reduction of surface metallic state [22-25] and oxide thickening [20] contribute to the improvement in corrosion resistance. In addition, the present author found that low Ni metal/TiO₂ ratio in the oxide layer together with the laser-induced physical changes in roughness and topography can promote better cell-surface interactions (i.e. higher attachment and viability of mesenchymal stem cells (MSCs) compared to the untreated NiTi [26]).

Taken together, the focus of existing literature on the laser-formed oxide layer on NiTi after LT has been on the evaluation of corrosion, Ni ion release and biocompatibility (i.e. formation of bone-like apatite *or in-vitro* culture of bone cells: osteoblasts or MSCs) performances. It is important to point out that biofilm infection is one of the main reasons for implant failure accounting for about 15 % of failure (i.e. ranked second to the failure caused by aseptic loosening)[7]. Biofilm infection is extremely difficult to cure due to its high resistance to antibiotic treatments and can result in substantial healthcare costs due to the need to remove the infected foreign body to cure infection [27,28]. A biofilm-mediated infection starts with initial bacterial adherence to a surface, followed by growth, replication, and production of protective extracellular substances which shield the resulting sessile bacterial community. Initial reversible bacterial adherence starts within the first few seconds and extends to approximately 2h, after which adherence becomes irreversible [29]. Accordingly, bio-fouling by bacterial is much faster than the corrosion and Ni ion release processes (usually take days to months to occur). Bacterial adherence can interfere with the tissue regeneration processes on the implant surfaces [30], and also exacerbate the corrosion behaviours of NiTi [31] thus worsening the Ni ion release problem. Surface chemistry is known to be a key factor, along with hydrophobicity, surface roughness and topography, to impact the initial adhesion and aggregation of bacteria (e.g. *Staphylococcus epidermidis* [32], *Staphylococcus aureus* [33], and *Escherichia coli* [34]). Up to the present, no effort has been devoted to investigating the effectiveness of the laser-formed oxide layer on inhibiting the bacterial adherence and biofilm formation on NiTi surface after LT.

In this study, martensitic NiTi alloy were laser-treated using the newly-developed LT technique [33,35,36]. This LT technique is designed to operate in an open air environment with the treatment area protected by coaxially delivered Ar, and hence no additional gas chamber and tubes are required. It can be applied to treat the free-form, complexly-shaped surfaces that are commonly seen in biomedical implants. The major objective of this study was to investigate the surface chemistry effects of NiTi after LT on inhibiting the *in-vitro* bacterial adherence and biofilm formation of *S. aureus*. The effects of other laser-induced surface characteristics: surface roughness, topography and wettability were also analysed and systemically compared with the two untreated NiTi samples, namely as-received and mechanically-polished surfaces.

2. Methodology

2.1. Material preparation

The near-equiatomic NiTi (55 % Ni, balance Ti by weight) alloy used in this study was sourced from American Elements (Los Angeles, California, USA). The NiTi material was purchased in form of a plate with the size of 250 mm x 250 mm and thickness of 2 mm. The samples, in form of circular discs of 15 mm in diameter, were cut from the NiTi plate using the electrical discharge machining (EDM) wire-cut method. Before laser treatment, the sample surfaces were sequentially ground with a range of sandpapers from 120 to 400 grit to ensure the consistency in surface roughness and to remove the pre-existing surface oxide layer. The disc samples after grinding were cleaned in an ultrasonic bath filled with ethanol for 10 mins, then rinsed in distilled water for 10 mins and eventually dried completely in air.

2.2. Thermal transformation analysis

The transformation temperatures of the NiTi sample, namely Martensite start (M_s), Martensite finish (M_f), Austenite start (A_s) and Austenite finish (A_f) were measured using a differential scanning calorimeter (DSC – Pyris™ Diamond, PerkinElmer Instruments, Waltham, MA, USA). The temperature range scanned in the DSC analysis was between -150 and 150 °C, and the scanning rate for heating and cooling was 10 °C /min.

2.3. Laser surface treatment

The laser treatment was performed using a fully automated continuous-wave (CW) 200 W fiber laser system. The laser system was integrated by Micro Lasersystems BV (Driel, The Netherlands). The fibre laser was manufactured by SPI Lasers UK Ltd (Southampton, UK). The laser wavelength was 1064 nm (or 1.06 μ m). The disc sample was irradiated with a laser beam with the spot size of 100 μ m under the shielding of high purity Ar gas. The Ar gas was delivered coaxially with the laser beam via a standard laser nozzle, and thus, an Ar pressure zone was established between the tip of the laser nozzle and substrate surface. The laser-irradiated surface was fully covered by a number of overlapping laser-tracks with overlapping ratio of 50 % in track width. The graphical representation of the laser treatment setup is given in Fig.1.

A preliminary study was conducted to identify the process window in terms of laser power, scanning speed and shielding gas pressure. The objective of this preliminary study was to create the laser-treated (LT) surface without any observable surface defects, e.g. micro-cracks and discolouration. The laser power of 40 W, scanning speed of 25 mm/s and Ar flow rate of 30 L/min was found to be the optimal set of processing parameters that could yield the highest quality LT surface.

In order to gain a more comprehensive understanding on the effects of different surface conditions on the antibacterial behaviour of NiTi, the following four surface conditions: as-received surface, surface after polishing (i.e. mechanically-polished with 4000 grit sandpaper), LT surfaces with 40W and 45W were tested and compared with each other. In addition to the LT surface obtained by the optimal laser power of 40W, the surface treated by a higher laser power, namely 45W was also investigated. The reason was to understand whether the laser-induced defects (e.g. surface discolouration) would present any unfavourable effects to encourage more bacterial attachment on the NiTi surface. The four surface conditions were grouped into two categories: (i) the untreated group, namely as-received and polished surfaces and (ii) the LT groups, namely LT40W and LT45W surfaces hereafter.

Fig.1. Graphical representation of the laser treatment setup. The inset at the bottom-right showing the magnified view for selected area in the LT surface. The idea of overlapping laser tracks, with 50 % overlapping ratio in track width, is illustrated in the inset at the top-right corner.

2.4. Surface morphology and composition analysis (SEM-EDX)

The morphology and composition of the LT surfaces were analysed by Scanning Electron Microscopy (SEM – Leo 1455VP) and Energy Dispersive X-Ray Spectroscopy (EDX – Oxford Instruments X-Max detector with INCA software). All SEM images and EDX data were acquired with a 20 kV beam, at magnifications between 100x and 2000x and using backscattered electron (BSE) detection.

2.5. Crystal structure analysis (XRD)

XRD data were acquired using a Bruker D8 Advance Eco diffractometer in Bragg-Brentano geometry with a Cu X-ray tube operated at 40 kV 25 mA, fixed source slit (0.6 mm), 2.5° Soller slits and 0.02 mm Ni K_β filter on the source arm. Scans were made over 10° – 80° in 2θ. The detector arm was fitted with a fixed secondary slit (9.5 mm), 2.5° anti-scatter slits and a multi-channel linear detector (Bruker SSD160). All samples were ultrasonically agitated for 10 mins in acetone to remove any loose materials and weakly-adherent contaminants prior to XRD analysis.

2.5.1. Peak positions for the phases of NiTi

It is well known that NiTi alloys transform from a martensitic phase (monoclinic) through an intermediate phase (rhombohedral) to an austenite phase (body centred cubic). Iijima et al [37] used micro-area XRD to investigate clinical NiTi wires after exposure to a range of test liquids, and identified the presence of both the intermediate (rhombohedral) and the austenite phases. In a follow-up work Iijima et al [38] provided a useful table of references and peak positions to help in distinguishing peak positions of the three major NiTi phases.

This is adapted as Table 1 below. The potential for peak overlap and mis-assignment is clear from this table.

Table 1. Reference peak positions for three phases of NiTi, adapted from Iijima et al [38]

Diffraction angle 2 θ	Austenite (BCC) ICDD standard 18-0899 hkl plane	Martensite (monoclinic) ICDD standard 35-1281 hkl plane	R-phase (rhombohedral, Riva et al [39]) hkl plane
39.22		020	
41.36		1-11	
42.2			112
42.7			300
42.8	110		
43.92		002	
44.92		111	
61.58			222
61.98	200		
77.82			412
78.16	211		
78.18			330

In this work, either the reference data in Table 1 or equivalents from the International Centre for Diffraction Data (ICDD) PDF-4 database were used. PDF 01-072-3504 represented the hexagonal (intermediate or rhombohedral) form of NiTi, PDF 01-072-8487 represented the monoclinic (Martensite) form and PDF 01-072-0442 represented the Ti₂Ni (cubic phase).

2.6. Surface composition and oxidation state analysis (XPS)

XPS data were acquired using a bespoke UHV chamber fitted with Specs GmbH (Berlin) 150 mm hemispherical analyser with 9-channeltron detection, monochromated Al K α X-ray source at 1486.6 eV energy, and low energy electron flood gun for charge neutralisation. Survey scans over the whole binding energy range of interest were acquired using 50 eV pass energy and individual scans over the Ni 2p_{3/2}, Ti 2p and O 1s spectral regions were made using 20 eV pass energy. All spectra were charge corrected to C 1s = 285.00 eV (hydrocarbon contamination). Spectra were quantified using Scofield cross sections [40] corrected for the known energy dependencies of the analyser transmission and the electron effective mean free paths, normalised to 100 % of elements detected (i.e. excluding H or He). Samples were ultrasonically agitated for 10 mins in acetone to remove any loose materials and weakly-adherent contaminants prior to analysis.

2.6.1. Correction method to deduct the residual hydrocarbon overlayer

It was noted that even after ultrasonic cleaning in acetone, the samples showed relatively high proportions of carbonaceous contamination (i.e. carbon accounted for approximately 50 % of the composition in the surface region). The presence of this residual hydrocarbon overlayer made the surface composition results difficult to interpret quantitatively. To obtain the composition in the absence of the hydrocarbon overlayer, the data cannot simply be re-normalised to 100 % as the attenuation of the signals from each element depend on the kinetic energy of the photoelectron peaks. However, the authors have previously published a simple method for correction of quantitative results for the hydrocarbon contamination overlayer which overcomes this problem [41]. As the carbon found on the samples was likely to be confined to a contamination overlayer, the method has been applied to these data. In a small modification to the method, the organic portion of the O 1s contribution to the data was also included as part of the contamination overlayer. The surface composition (at. %) results before and after correction are shown in Table 4 (in the later section 3.4.1).

2.7. Surface roughness measurements

The 2D surface profiles of the samples and their arithmetic average of roughness (Ra) were measured using a portable roughness gauge (Rugosurf 10G, TESA Technology, Telford, Shropshire, UK). The surface roughness was measured in accordance with the Standard ISO4287 / JIS B0601. A total of 10 measurements were taken in different locations at the untreated and LT surfaces (i.e. perpendicular to the laser tracks). The mean with the standard error (SE) of $n = 10$ was used to indicate the Ra of the surfaces.

2.8. Contact angle measurements

The water contact angles on the sample surfaces were measured using the sessile drop method, coupled with a video-based contact angle analyzer (FTA 200, First Ten Angstroms Inc., Portsmouth, VA, USA). Image analysis was performed using the FTA 32 video software. The testing liquid was deionized water. The volume of each sessile drop was 5 μ l (i.e. controlled using a micro-litre syringe). Droplet images were captured in the direction perpendicular to the laser tracks at fixed time intervals (i.e. at least 60s). At least four measurements ($n = 4$) were made for each sample at room temperature.

2.9. Bacterial cell culture

Both the laser-treated and untreated control samples were used for bacterial adherence and biofilm formation assays. Samples were cleaned with pure ethanol (Sigma Aldrich, UK) in an ultrasonic bath for 15 min prior to bacterial cell culture. The dry, clean samples were then sterilized with 70 % ethanol for 10 min and washed three times with sterile phosphate buffered saline (PBS) before placing into individual wells of a 24-well tissue culture plate. *S. aureus* (ATCC 6538) was cultured in Müller Hinton Broth (MHB; Oxoid) overnight (18h) at 37 °C on a gyrotatory incubator with shaking at 100 rpm. The optical density of the overnight culture was adjusted to 0.3 at 550 nm, then diluted 1 in 50 with fresh sterile MHB. This provided a bacterial inoculum of approximately 10^6 colony forming units per millilitre (CFU)/ml, as confirmed by viable count. 1 ml of culture, at an inoculum not exceeding 2.4×10^6 CFU/ml, was applied to each well, such that the sample is completely submerged. Samples were incubated for 24h at 37 °C on a gyrotatory incubator with shaking at 100 rpm. At least three replicates of each condition, namely as-received, polished, laser-treated (LT) 40W and 45W were tested to ensure the consistency and validity of the results.

2.9.1 Bacterial viability analysis

After 24h of incubation, the samples were washed three times with sterile PBS to remove any planktonic bacteria. Sessile bacteria were stained by submerging samples in Live/Dead® BacLight™ stain (Molecular Probes) for 30 min at 37 °C in the dark. The fluorescent viability stain contains two components: SYTO 9 dye and propidium iodide. The SYTO 9 labels all bacteria, whereas propidium iodide enters only bacteria with damaged membranes, thus identifying non-viable cells. Green fluorescence indicates viable bacteria with intact cell membranes whilst red-yellow fluorescence indicates non-viable bacteria with damaged membranes. The stained bacteria were observed using a fluorescence microscopy (GXM-L3201 LED, GX Optical). At least eight random fields of view (FOV) were imaged per sample. The surface areas covered by the sessile, biofilm-associated bacteria were calculated using image analysis software (ImageJ, National Institutes of Health, <https://imagej.nih.gov/ij/>). The areas corresponding to adhered viable bacteria (green) and the non-viable bacteria (red) were determined. The total biofilm coverage was the sum of the green and red areas on the surface and the dead (or non-viable) cell coverage was the areas covered with red. Data presented are the means of measurements from the eight images. The significances of the observed differences between the sample means were analysed and compared using a Student's t-Test. A probability below $p < 0.05$ was considered as statistically significant.

3. Results and Discussion

3.1. Thermal transformation analysis

The transformation temperatures of the NiTi sample are given in Table 2. From the results in the table, it is known that the NiTi was in fully Martensitic state in the laser treatment experiment because the working temperature (room temperature about 20 °C) was lower than M_f (24.8 °C) [4, 42].

Table 2. Transformation temperatures of the NiTi sample measured by DSC

Condition	Martensite Start Temperature, M_s (°C)	Martensite Finish Temperature, M_f (°C)	Austenite Start Temperature, A_s (°C)	Austenite Finish Temperature, A_f (°C)
As-received NiTi	66.2	24.8	60.5	95.6

3.2. SEM micrograph analysis

Fig.2a-j show the overview photo of the laser-treated (LT) samples and their SEM micrographs captured at different magnifications between 100x and 2000x. In the overview photo (Fig.2f) and SEM images at lower magnification (100x and 500x in Fig.2g and 2h), a number of surface particles together with a discoloured region can be found at the surface of LT45W sample. Fig.3 shows the SEM image (100x) at the boundary between the non-discoloured and discoloured (blue) regions at the LT45W surface. The surface is rough because of presence of the particles. In comparison, the LT40W surface exhibited a silvery and smoother surface morphology without any discolouration. The recognizable surface particles (i.e. identified at the LT45W surface) is absent from the LT40W surface. However, some elongated and thin features can be observed in the higher magnification SEM (1000x in Fig.2d).

On the other hand, distinctive ripples in micrometre size ($> 1\mu\text{m}$) can be found in the LT40W and LT45W surfaces in the SEMs of 2000x. The peak-to-peak distance between the micro-ripples (i.e. indicated by the white arrowhead lines in Fig.2e and 2j) are measured using the ImageJ software and reported in Table 6 (in the later section 3.5). It is known that such micro-rippled structure resulted from the oscillation of the liquid metal due to Marangoni convection and hydrodynamic processes driven by capillary waves acting on the melt pools [43-45].

Fig.2. Overview photo and SEM images of the LT surfaces: (a-e) LT40W, (f-j) LT45W captured at different magnifications. Images are arranged with increasing magnifications down from the top (5x) to bottom rows (2000x). The photo in the top row showing the overview of surface morphology of all LT samples while the SEMs in the remaining rows showing the magnified views of surface features in the selected LT areas.

Fig.3. SEM image (100x) at the boundary between the non-discoloured and discoloured (blue) areas at LT45W surface. Inset at the bottom-right corner is a higher magnification SEM (500x) showing a more-detailed view in the discoloured area. The re-solidified particles and micro-ripples can still be observed in the inset, but the number and size of particles are greatly reduced when compared to the non-discoloured area.

Fig.4a-b provide the SEM-EDX images of the LT surfaces. The element maps in the figures show the spatial distributions of Ti, Ni, and O in the LT surfaces. The SEM-EDX results (Fig.4a) indicate that the Ti and Ni elements in the LT40W surface are uniformly distributed with no measureable spatial variation over the surface area imaged, suggesting the elongated and thin particles (extended from the boundary to the interior areas of laser tracks) are laser-induced surface features as a consequence of the complex melt pool dynamics and solidification processes during laser treatment. In the LT45W surface (Fig.4b), no significant difference between Ti and Ni, in terms of spatial variation, can be observed. The surface particles have identical composition to the surrounding material as seen by the absence of any differences in contrast in the element distribution maps. This indicates that the particles are likely to be re-solidified droplets of molten material (i.e. liquid droplets ejected from the melt pool due to the laser-pressure-melt pool interactions). Black areas (indicated by the arrows in Fig.4b) in the Ti element map are due to shadowing where raised features on the surface obstruct the X-rays from reaching the detector. The additional O element map is provided to show the enhanced intensity of oxygen (i.e. oxidation) on some of the re-solidified droplets.

Fig.4. SEM-EDX images of the LT surfaces: (a) LT40W, (b) LT45W. The element maps show the spatial distribution: Ti, Ni, and O in the LT surfaces. Several black and white spots appear in the element maps of Ti and O respectively.

3.3. XRD microstructure analysis

XRD is a common analytical tool used to study phase changes of NiTi alloys. The XRD patterns for the untreated and laser-treated samples are shown in Fig.5. The pattern for the as-received sample was relatively weak, with broad features on a relatively high background indicating a relatively poor degree of crystallinity and small crystallites. Similar to the as-received sample, the pattern recorded for the polished sample was weak and broad on a relatively high intensity background, indicating poor crystallinity. However, in this case the monoclinic form (Martensite) appeared more dominant, and the broad shoulder-like component to the pattern visible in Fig.5 at approximately 40° has become a relatively well-developed peak. This may indicate the partial removal of the intermediate hexagonal (rhombohedral) phase during polishing, suggesting this higher temperature phase was originally present only as a surface layer possibly originating from heating during manufacture or cutting to shape.

Fig.5. Room-temperature XRD profiles of the untreated and LT samples. The scanning diffraction angles are between 10° and 80°. Inset at the top-right corner showing the enlarged profiles of as-received and polished samples between 35° and 50°.

Regarding the XRD pattern for the laser-treated sample namely LT40W, dramatic changes compared to the as-received and the polished samples were seen. Diffraction peaks were sharper and more intense on a low intensity background. This indicated a much higher degree of crystallinity with larger crystallites. Elements of the original pattern were still present in the range 35 – 50°. The peak around 42.5° was much more intense, as were several other features, and a new peak at approximately 13.6° was present. This peak and several others in the pattern including the most intense line were matched by the ICDD reference pattern 01-072-0442 Ti_2Ni (i.e. a cubic phase). It is particular to note that the reference peak for this phase expected at approximately 71° was only weakly present, and this can be attributed to preferential orientation of the Ti_2N crystallites at the sample surface. Likewise, the LT45 W sample also showed a mix of the monoclinic (Martensite) and rhombohedral (intermediate) NiTi phases, with a strong contribution from Ti_2Ni . There appeared to be stronger preferential orientation effects in this sample (LT45W) with all expected peaks being present. This can be attributed to further development of an oriented surface crystallinity due to the increased laser power and therefore higher local temperatures and longer re-crystallisation times. In particular, the peak due to the monoclinic phase at 60° was prominent, and minor peaks in the range 28 – 30° attributed to both the monoclinic and the rhombohedral phases were visible.

3.3.1. Calculation of X-ray penetration depth in the XRD measurements

The geometry for the XRD measurement is shown in Fig.6.

Fig.6. Geometry of the XRD measurement.

X-rays enter from the left at an angle θ to the surface and are scattered through an angle 2θ at a depth d , representing the penetration depth. The path length l travelled by the X-rays in the sample is given from the geometry by

$$l = 2a = 2d/\sin \theta$$

X-rays are attenuated in the sample according to

$$\frac{I}{I_0} = e^{-(\mu/\rho)x}$$

Where I/I_0 is the reduced intensity after passing through a length x of material with mass absorption coefficient μ and density ρ [46].

Values of mass absorption coefficient, density and hence the linear absorption coefficient are shown in Table 3 [47].

Table 3. Mass attenuation coefficients, densities and linear absorption coefficients for Ni and Ti.			
Element	μ , $\text{cm}^2 \text{g}^{-1}$	ρ , g cm^{-3}	Linear absorption coefficient $\mu\rho$, cm^{-1}
Ni	4.952×10^1	8.908	441.1
Ti	2.023×10^2	4.507	911.8
Mean:			676.4

The alloy is 50:50 Ni:Ti and X-ray absorption is additive [46] therefore the linear absorption coefficient is the mean of the values for Ni and Ti, and is 676.4 cm^{-1} . This is the distance the Cu K α X-rays can travel in the material before their intensity is attenuated to $1/e$ of its original value, i.e. for a distance of $1/676.4 \text{ cm}$ (or $14.78 \mu\text{m}$). Using this as the value for l in the expression above gives values for the information depth of the XRD measurement ranging from $1.28 \mu\text{m}$ at $\theta = 10^\circ$ to $7.28 \mu\text{m}$ at $\theta = 80^\circ$. In the mid-range of the value for $2\theta = 42^\circ$, where the peaks of interest for NiTi occur (as shown in Fig.5), the information depth is approximately $2.65 \mu\text{m}$.

To summarise the XRD results, laser treatment resulted in the formation of the Ti_2Ni phase and no evidence was seen for any Ni-/Ti- oxides in the sub-surface region (between the depth of 1.3 and $7.3 \mu\text{m}$).

3.4. XPS surface chemistry analysis

3.4.1. XPS survey scans

The surface compositions (at. % after carbon correction as described in the previous section 2.6.1) for the untreated and laser-treated (LT) samples are provided in Table 4.

Table 4. Surface composition (at. % after correction) at the outermost surfaces of the untreated and LT samples measured by XPS. The composition (at. %) before correction are given in the parenthesis.

Sample		Ti 2p	Ni 2p _{3/2}	O 1s	C 1s (Hydrocarbon Overlayer)	Other Elements (N 1s, Na 1s, Ca 2p, Zn 2p _{3/2})
Group	Condition					
Untreated	As-received	23.2 (9.6)	4.8 (1.2)	49.1 (36.5)	(47.3)	22.9 (5.5)
	Polished	23.9 (9.5)	15.3 (3.5)	45.2 (36.0)	(47.9)	15.6 (3.0)
Laser-treated (LT)	LT40W	20.7 (7.0)	16.7 (3.1)	52.4 (32.8)	(53.9)	10.3 (3.2)
	LT45W	20.7 (9.9)	22.1 (7.0)	55.7 (36.6)	(45.8)	1.5 (0.7)

All surfaces showed the expected Ni, Ti and O with the presence of low levels of other elements namely N, Na, Zn and Ca. The presence of N was likely due to incorporation of a small amount of nitrogen in the surface layers due to (i) exposure of the samples in air [48] or (ii) sample preparation process such as grinding and polishing [49]. Na, Zn and Ca were frequently found as a low-level surface contaminant in XPS analysis of surfaces. The Ti levels remained approximately constant at 21-23 at. % for all samples and the Ni concentrations showed an increasing trend across the sample series (i.e. increased from 4.8 to 22.1 at. %).

3.4.2. XPS narrow scans

The XPS narrow scan spectra: Ni 2p, Ti 2p and O 1s for the untreated and LT samples are shown in Fig.7. As summary of the species detected in the samples analysed is shown in Table 5.

Table 5. Summary of the chemical species detected by XPS

Element	line	Assignment	BE range (eV)	Present in sample			
				As received	Polished	LT40W	LT45W
Ni	2p _{3/2}	Ni metal	852.2 - 852.5	Yes	Yes	v. weak	No
		Ni oxide/ hydroxide	855.3 - 855.8	Yes	Yes	Yes	Yes
Ti	2p _{3/2}	Ti metal	454.5	Yes	Yes	No	No
		Ti intermediate oxide	456.2	Yes	Yes	v. weak	No
		Ti in TiO ₂	458.4	Yes	Yes	Yes	Yes
O	1s	Ti-O, Ni-O	529.8	Yes	Yes	Yes	Yes
		O=C	530.9 - 531.5	Yes	Yes	Yes	Yes
		O-C	532.3 - 532.9	Yes	Yes	Yes	Yes
C	1s	C-C	285	Yes	Yes	Yes	Yes
		C-O	286 - 286.4	Yes	Yes	Yes	Yes
		C=O	287.2 - 288.1	Yes	Yes	Yes	Yes
		COO-	288.6 - 289.3	Yes	Yes	Yes	Yes

Fig. 7. XPS narrow scan spectra: Ti 2p, Ni 2p3 and O 1s for the untreated and LT surfaces: (a-c) as-received, (d-f) polished, (g-i) LT40W, (j-l) LT45W.

3.4.3. Ti 2p spectra

The Ti 2p spectra of the untreated and LT samples in Fig.7 showed strong $2p_{3/2}$ - $2p_{1/2}$ doublets corresponding to TiO_2 , with the main $2p_{3/2}$ component at typically 458.4 eV. The majority of Ti at the outermost surfaces was found in the $Ti(4+)$ state (i.e. oxidised as TiO_2). However, small amounts of metallic component and intermediate oxides: TiO and Ti_2O_3 were found on the as-received (Fig.7a) and polished (Fig.7d) samples. A very small amount of intermediate oxide was found on the LT40W sample (Fig.7g). The LT45W sample (Fig.7j) showed only TiO_2 at the surface. The ratio of the metallic peak to the oxidised peak intensity of Ti is shown in Fig.8. It can be seen that Ti metallic component was absent from the surfaces after laser treatment (namely LT40W and LT45W). The polished sample showed a slightly higher metal / oxide ratio than that of the as-received sample. As seen in the Ti and Ni spectra, the Ti and Ni at the surface became fully oxidised with the laser treatment.

Fig.8. Metallic / oxidised Ti ratio at the outermost surfaces of the as-received and LT samples.

3.4.4. Ni 2p spectra

The Ni $2p_{3/2}$ - $2p_{1/2}$ doublet is known to be of complex form in the presence of oxides of Ni, with numerous and varied shake-up structures present [50]. In this study, the metallic component was fitted with a single symmetric peak centred on 852 eV binding energy whilst the oxide component was fitted with a broader peak typically around 855 eV, and the shake-up features were fitted with up to three symmetrical components. In Fig.7, the Ni metallic component was visible on the as-received (Fig.7b) and polished (Fig.7e) samples, indicating only a very thin Ni oxide layer on these surfaces. There was little evidence for its presence on the LT40W sample (Fig.7h), and it was not present on the LT45W sample (Fig.7k). The ratio of the metallic peak to the primary oxidised peak intensity of Ni is shown in Fig.9. The results indicate that the absence of any detectable non-oxidised Ni for the LT40W and LT45W samples was clear. Comparing with the as-received sample, the increase in the metal / oxide ratio on the polished sample was visible (i.e. due to the removal of surface oxide during polishing).

Fig.9. Metallic / oxidised Ni ratio at the outermost surfaces of the as-received and LT samples.

3.4.5. O 1s spectra

The oxygen spectra for the untreated (Fig.7c and 7f) and LT samples (Fig.7i and 7l) typically showed a relatively sharp lower binding energy metal-bonded component and a broader but relatively featureless higher binding energy component typical of organic oxygen, e.g. bonded within the hydrocarbon contamination layer. The metal-bonded component was typically found in the range 529.4 – 530.0 eV. The carbon-bonded oxygen was typically fitted with two components at approximately 531.2 eV and 532.6 eV representative of O=C and O-C bonding respectively, typically associated with residual organic contamination.

To summarise, these Ni-/Ti-oxides were detected by XPS but not seen in the XRD analysis. Therefore, they were confined to the outermost surface layers only.

3.5. Surface roughness analysis

Fig.10(a-d) provide the 2D roughness profiles for the untreated and LT surfaces. The roughness profile of as-received surface (Fig.10a) shows a typical randomly rough surface. Some deep narrow valleys (i.e. resulted from the manufacturing defects such as scratches, grooves, pits, etc.) are observed. In contrast, a very smooth surface can be seen from the polished surface. Inset in Fig.10b is an enlarged view of the selected segment showing the submicron (< 1 μ m) surface finish in the polished surface. Related to the LT samples, rough surfaces can be observed with periodic repeating patterns appearing throughout the surface profiles. Such repeating patterns are highlighted in dashed boxes in Fig.10c for LT40W and Fig.10d for LT45W. In comparison with LT40W surface, the amplitude and wavelength of the repeating pattern in LT45W surface are larger and longer. Further, the roughness within the repeating pattern of LT45W surface are more fluctuating and irregular, indicating the roughness irregularity increasing with higher laser power.

Fig.10. 2D roughness profiles of the untreated and LT surfaces: (a) as-received, (b) polished, (c) LT40W, (d) LT45W. All profiles are plotted with the same scale. The measurements were made in the direction perpendicular to the laser tracks.

The roughness parameter, namely the arithmetic average of roughness (R_a), is extracted from the surface profiles in Fig.10 and plotted in a bar chart in Fig.11. The mean R_a values of the untreated and LT surfaces

are given in Table 6. As seen from the results, the as-received surface has the highest Ra among all samples but the value drops to a very low level after polishing (i.e. Ra of the polished surface is the lowest). Regarding the LT group, the LT45W surface has a higher Ra than the LT40W surface, and particularly, its value is only slightly smaller than that of the as-received surface. The order of Ra (from highest to lowest) is: as-received > LT45W > LT40W > polished.

Table 6. Mean Ra values of the untreated and LT surfaces extracted from the roughness profiles in Fig.10 and the peak-to-peak distance between the micro-ripples in LT surfaces measured in the SEM micrographs in Fig.2.

Sample		Arithmetic Average of Roughness, Ra (μm) (Measured by Surface Roughness Tester)			Peak-to-Peak Distance between Micro-ripples (μm) (Indicated in Fig.2.) (Measured by the ImageJ Software)		
Group	Condition	Mean	SE	n	Mean	SE	n
Untreated	As-received	1.2	0.06	10	---	---	---
	Polished	0.1	0.004	10	---	---	---
Laser-treated (LT)	LT40W	0.7	0.02	10	4.8	0.7	10
	LT45W	1.1	0.02	10	3.6	0.5	10

Fig.11. Mean Ra values extracted from the surface roughness profile in Fig.10. The error bars indicate the SE of $n = 10$.

3.6. Contact angle analysis

The water contact angles measured on the untreated and LT surfaces are given in the bar chart in Fig.12. The mean contact angles are tabulated in Table 7. The results show that the polished surface has the highest contact angle followed by the as-received surface which has a slightly lower value. All LT surfaces have a noticeably lower contact angle than surfaces in the untreated group. Within the LT group, the contact angle on LT40W surface is higher than that on LT45W surface. The order of water contact angle is polished > as-received > LT40W > LT45W.

Table 7. Water contact angle of the untreated and LT samples measured by sessile drop method

Sample		Water Contact Angle (degree)		
Group	Condition	Mean	SE	n
Untreated	As-received	68.4	2.4	4
	Polished	73.2	0.6	4
Laser-treated (LT)	LT40W	61.4	1.4	4
	LT45W	57.5	0.4	4

Fig.12. Mean water contact angles measured from the untreated and LT surfaces. The error bars indicate the SE of n = 4.

3.7. Bacterial cell culture of *S. aureus*

The images obtained by fluorescence microscopy, accompanied by Live/Dead staining (Fig.13a-d), show a marked contrast between adherence and biofilm formation of *S. aureus* on the untreated surfaces, namely in its as-received state or polished (Fig.13a-b) compared to LT surfaces (Fig.13c-d). Live/Dead staining will stain live (viable) bacteria green (SYTO 9 nucleic acid stain), while dead (non-viable) bacteria are stained red (propidium iodide). As seen in Fig.13a-b, a large percentage of the as-received and polished surfaces are stained in green, indicating extensive bacterial adherence and the establishment of biofilm, with a minimal amount of red staining (Fig.13c-d). In comparison, a mix of green and red staining on a dark background can be observed from the LT surfaces, and more importantly, the area covered in adhered viable biofilm is significantly less than on the untreated surfaces. The small number of adherent bacterial on the LT surfaces exist as planktonic cells or micro-colonies. The orange/red staining of a proportion of these bacteria on the LT surface indicates that the bacterial cells that do adhere encounter an inhospitable environment and become non-viable.

Fig.13. Fluorescence microscopy images of *S. aureus* cells captured on the untreated and LT surfaces: (a) as-received, (b) polished, (c) 40W, (d) 45W (after 24h cell culture accompanied by Live/Dead staining). Green colour refers to the alive bacterial cells whilst red colour refers to the dead cells.

Fig.14a-b show the total biofilm coverage (Fig.14a) and deal cell coverage (Fig.14b) measured from the untreated and LT surfaces. In Fig.14a, the as-received surface had the highest biofilm coverage among all samples. The differences are statistically significant at $p < 0.05$ when compared with the polished surface and at $p < 0.01$ when compared with the LT surfaces, namely LT40W and LT45W. The polished surface had a higher biofilm coverage than the LT surfaces at the statistical significance of $p < 0.05$. No significant differences are found between the LT40W and LT45W surfaces. The order of total biofilm coverage is: as-received > polished > LT40W and LT45W. In Fig.14b, the as-received surface had the highest viable bacterial coverage among all samples. The differences are statistically significant at $p < 0.05$ when compared with the polished surface and at $p < 0.01$ when compared with all LT surfaces. Within the LT group, the LT40W surface had a statistically significantly higher dead cell coverage than the LT45W surface at $p < 0.05$. Unlike the case in the total biofilm coverage (Fig.14a), the order of dead cell (non-viable) coverage between the polished and LT surfaces (in Fig.14b) cannot be straightforwardly established because the variance in data set for the polished surface overlapped with those for the LT surfaces, even though the mean coverage of polished surface is lower than LT40W surface and higher than LT45W surface.

Fig.14.(a) Total biofilm coverage and (b) dead cell coverage measured from the fluorescence microscopy images in Fig.13. Error bars are the SE of $n = 8$. The total biofilm coverage in (a) refers to the total sum of green and red areas in surfaces after cell staining. The deal cell coverage in (b) refers only to the red areas after cell staining. The cell coverage measurements were done using the ImageJ software.

It is evident from these fluorescence microscopy images (Fig.13a-d) and the accompanying graphs of statistical analysis for total biofilm coverage (Fig.14a) and dead cell coverage (Fig.14b) that the untreated surfaces showed significantly greater attachment, adherence, and biofilm formation than surfaces that have undergone laser treatment, which display a markedly reduced biofilm coverage.

3.7.1. Biofilm coverage comparison between the as-received and polished surfaces

Surface roughness has been reported extensively in literature regarding its effects on influencing bacterial attachment. It is generally accepted that, within a certain range, the surface roughness (in terms of Ra value) correlates positively with the amount of bacterial attachment [51]. The major reason supporting this argument is that the topographic features in a rougher surface, e.g. pits and troughs provided higher surface area for the bacteria to attach and meanwhile protects them from environmental disturbances (i.e. shear forces)[52]. Particularly, the surfaces with topographic features which are comparable in size to the bacterial cells (approximately 1 to 2 μm) would encourage the strongest bacterial attachment given that the contact area between the cells and substrate surface was maximised [52-54]. This relationship is found to hold for the samples before laser treatment in our study. The as-received sample (i.e. a rougher surface with Ra of 1.2 μm) had a statistically significant higher biofilm coverage on the surface as compared to the polished sample (i.e. a smoother surface with Ra of 0.1 μm).

It is known that bacterial attachment, adhesion and the subsequent formation of biofilm on a surface depend on a number of interplaying physical and chemical factors of the surface, such as surface roughness and topography (physical), surface chemistry (chemical) and wettability (physiochemical)[33]. When analysing bacterial attachment, the surface chemistry and wettability need to be taken into account in addition to the surface roughness and topography. In our study the as-received and polished surfaces were covered by a naturally formed oxide in air. The XPS analysis indicated that the air-formed oxide layer on the two surfaces were mostly heterogeneous in nature, which included a mix of metallic (i.e. Ti^0 and Ni^0) and oxide (i.e. Ti-based and Ni-based oxides) components. They differed mainly in the surface composition with higher concentration of Ni metal appearing in the polished surface. The poor crystallinity in the sub-surface regions of the as-received and polished samples pointed to the fact that the surface oxide layer was more likely to be amorphous than crystalline. These observations are consistent with the findings by Pequegnat et al. [15] that mechanical polishing facilitated the formation of amorphous oxide on the surface. The water contact angle on the as-received surface was only slightly lower than the polished surface, indicating their similar wetting behaviour (i.e. hydrophobic surfaces).

Taking all this together, the higher biofilm coverage on the as-received surface is likely due mainly to the physical factors of the following: (i) presence of manufacturing defects in surface, such as pits and grooves, which sheltered the cells from the environmental shear forces and (ii) surface roughness comparable with the size of *S. aureus* cells (i.e. around 1 μm). Further, the higher concentration of metallic Ni at the polished surface might contribute to the lower biofilm coverage given the potential toxicity effect of Ni to microorganisms [55].

3.7.2. Biofilm coverage comparison between the untreated (as-received and polished) and LT surfaces

The biofilm coverage on surfaces after laser treatment is statistically significantly lower than that on the untreated surfaces (i.e. as-received and polished surfaces) even though the laser-treated surfaces had a high surface roughness with the presence of recognizable topographical features, namely, the micro-ripples appearing within the laser-remelted tracks. Particularly, the Ra of LT45W surface (i.e. 1.1 μm) is only slightly lower than the as-received surface. It also means that the relationship of higher surface roughness encouraging more bacterial attachment is no longer valid for the case of laser-treated surfaces. It is important to note that the LT40W surface had a lower Ra value (i.e. 0.7 μm) than LT45W. Moreover, the LT45W surface possessed numerous re-solidified droplets which could potentially act as anchoring points for the bacterial cells to attach and could also become the shelter to reduce the influence of environmental shear forces on the initially-attached cells. Such droplets could not be found on the LT40W surface. Having said this, both the laser-treated surfaces (i.e. LT40W and LT45W) exhibited similarity in the biofilm coverage. It indicates that the lower biofilm coverage measured on the laser-treated surfaces cannot be explained by surface roughness and topography.

The enhanced protection of the laser-treated surfaces against bacterial attachment might be attributable to two possible mechanisms: firstly by producing a surface environment which is inhospitable to primary attachment of the bacteria, and secondly through bactericidal mechanisms that serve to kill surface-attached bacterial cells. These might be based on the combined effects of (i) increased hydrophilicity, (ii) enhanced surface chemical stability and (iii) micro galvanic action between the Ti_2Ni and $NiTi$ matrix. All of which will be explained in detail in the following:

First, it is known that surface hydrophobicity of bacterial cells plays an important role to control the initial attachment (i.e. hydrophobic surfaces favoured the attachment of hydrophobic cells)[56,57]. *S. aureus* cells are known to be hydrophobic (water contact angle of 72° , which is attributed to the hydrophobic teichoic and lipoteichoic acid constituents of the *S. aureus* cell wall [58]) and thus should prefer attaching to hydrophobic surfaces. The water contact angles measured on the laser-treated surfaces were smaller than the untreated surfaces, indicating an improved wetting behaviour after laser treatment. The increased hydrophilicity (that is, decreased hydrophobicity) of laser-treated surfaces could discourage the initial attachment of *S. aureus* cells due to mismatch of hydrophobicity.

Second, laser treatment led to the formation of a more homogenous and crystalline oxide layer. It has been reported that surface defects in the oxide layer can act as traps for charge carriers contributing to bacterial attachment (i.e. surface carriers are responsible for the electrostatic interactions between the bacterial cells and substrate [59]). The XPS analysis indicates that the surface defect states in the oxide layers after laser treatment was significantly reduced through the complete oxidation mechanism. The oxide layers were predominately composed of Ti- and Ni-oxides after laser treatment, and the metallic states of Ni and Ti were reduced to a very tiny amount at the LT40W surface and dropped to zero at the LT45W surface, indicating that they had a lower capacity to donate electrons to the bacterial cells and thus resulting in less bacterial attachment [33].

In addition, the laser-treated surfaces were subjected to several thermal cycles as adjacent laser tracks were made during the laser treatment process [15]. Such subsequent thermal cycles can lead to higher crystallinity of oxide. Crystallinity of the oxide layer has a direct effect to influence the bacterial attachment with the crystalline TiO_2 exhibiting a stronger antibacterial effect than the amorphous TiO_2 [60]. The XRD results of the laser-treated samples showed a much higher crystallinity in the sub-surface regions. It is believed that the oxide layers after laser treatment were more crystalline than the untreated surfaces. Moreover, the secondary phase (i.e. Ti_2Ni) in surfaces after laser treatment can form a micro galvanic pair [61,62] with the $NiTi$ matrix with $NiTi$ acting as the cathode and Ti_2Ni , the anode, due to their different potentials in the presence of electrolyte [63]. A mechanism proposed by Gallo et al. [63] suggests that the cathodic reaction between the galvanic pair can create a proton-depleted region between the bacterial membrane and substrate, leading to the adenosine triphosphate (ATP) synthesis and death of bacterial cells.

To summarise, we purpose that the higher biofilm coverage on the as-received surface, compared with the polished surface, is primarily due to its higher surface roughness and presence of manufacturing defects in surface that are hospitable to the bacterial cells. The effect of surface chemistry and wettability coming from the air-formed oxide layer plays a minor role to control bacterial attachment. In contrast, the substantial reduction of biofilm coverage on laser-treated surfaces, compared with the as-received and polished surfaces, resulted chiefly by the advantageous changes in surface chemistry and hydrophilicity accompanied with presence of the laser-formed oxide layer. The laser-induced surface roughness and topography (in micrometre scale) have a little effect on bacterial attachment.

4. Conclusions

In this study, the antibacterial performance of martensitic NiTi after laser treatment was systematically investigated and compared with the untreated NiTi in its as-received state and after polishing. The findings indicated that laser treatment can significantly improve the resistance of NiTi against bacterial attachment and biofilm formation.

The important results arisen from this study can be summarised as follows:

- i) The optimal processing parameters for laser-treating the NiTi was identified as 40W (laser power), 25 mm (scanning speed), 100 μm (laser spot diameter) and 30 L/min (Ar gas flow rate). Discolouration and re-solidified droplets appeared when laser-treating the surfaces with higher laser power, namely 45W.
- ii) The samples after laser treatment showed a mix of martensitic and rhombohedral phases with the precipitation of Ti_2Ni phase in the sub-surface region. The surfaces were completely oxidized after laser treatment, and the laser-formed oxide layers at the outermost surface were found be more homogenous and crystalline than the naturally-formed air oxides on the untreated surfaces.
- iii) The surface roughness after laser-treatment was somewhat lower than the as-received surface but much higher than the polished surface, indicating the laser-treated surfaces were still rough. However, the water contact angles on the laser-treated surfaces were smaller than the untreated surfaces, indicating the increased hydrophilicity (that is, decreased hydrophobicity) after laser treatment.
- iv) The biofilm coverage on the surfaces after laser-treatment was significantly reduced on account of the (a) improved homogeneity and higher crystallinity of laser-formed oxides at the outermost surface, (b) increased surface hydrophilicity, as well as (c) possible micro-galvanic action between the Ti_2Ni precipitates and NiTi matrix in the sub-surface region.

Though the laser treatment technique was applied to martensitic NiTi in this study, it is reasonably postulated that it would apply equally well to austenitic NiTi and in fact to any Ti-based alloys, for which the resistance against initial bacterial adherence and the subsequent biofilm formation relied on the quality of the outermost TiO_2 surface layer.

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FIBRE LASER TREATMENT (LT) OF MARTENSITIC NITI ALLOY IN OPEN AIR ENVIRONMENT

Surface Properties After LT

(i) Complete oxidation of TiO_2 layer at the outmost surface

Absence of metallic Ti

(ii) Increased surface hydrophilicity

(iii) Precipitation of Ti_2Ni in the subsurface region

Enhanced Antibacterial Resistance After LT

Green colour refers to alive bacterial cells whilst red colour refers to dead cells

Graphical abstract

Research Highlight

- Martensitic NiTi alloy is proposed for load-bearing applications;
- Laser treatment (LT) in open air was applied to modify the surface of martensitic NiTi;
- The outermost surface layer of laser-treated NiTi did not contain metallic Ti or Ni;
- The laser-treated NiTi contained Ti_2Ni precipitates in the subsurface region;
- Martensitic NiTi after LT exhibited an enhanced antibacterial resistance.

ACCEPTED MANUSCRIPT