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Foddai, A. C. G., & Grant, I. R. (2015). An optimised milk testing protocol to ensure accurate enumeration of viable Mycobacterium avium subsp. paratuberculosis by the PMS-phage assay. *International Dairy Journal*, *51*, 16-23. https://doi.org/10.1016/j.idairyj.2015.07.004

Published in:
International Dairy Journal

Document Version:
Peer reviewed version

Accepted Manuscript

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PII: S0958-6946(15)00140-5

DOI: 10.1016/j.idairyj.2015.07.004

Reference: INDA 3857

To appear in: *International Dairy Journal*

Received Date: 4 April 2015

Revised Date: 11 July 2015

Accepted Date: 12 July 2015

Please cite this article as: Foddai, A.C.G., Grant, I.R., An optimised milk testing protocol to ensure accurate enumeration of viable *Mycobacterium avium* subsp. *paratuberculosis* by the PMS-phage assay, *International Dairy Journal* (2015), doi: 10.1016/j.idairyj.2015.07.004.

An optimised milk testing protocol to ensure accurate enumeration of viable *Mycobacterium avium* subsp. *paratuberculosis* by the PMS-phage assay

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Abstract

There is interest in determining levels of *Mycobacterium avium* subsp. *paratuberculosis* (MAP) contamination in milk. The optimal sample preparation for raw cows' milk to ensure accurate enumeration of viable MAP by the peptide-mediated magnetic separation (PMS)-phage assay was determined. Results indicated that milk samples should be refrigerated at 4 ℃ after collection and MAP testing should commence within 24 h, or samples can be frozen at -70 ℃ for up to one month without loss of MAP viability. Use of Bronopol is not advised as MAP viability is affected. The vast majority (> 95%) of MAP in raw milk sedimented to the pellet upon centrifugation at 2500 × *g* for 15 min, so this milk fraction should be tested. De-clumping of MAP cells was most effectively achieved by ultrasonication of the resuspended milk pellet on ice in a sonicator bath at 37 kHz for 4 min in 'Pulse' mode.

1. Introduction

Due to its possible association with various human conditions, such as Crohn's disease and ulcerative colitis (Pierce, 2010) and Type 1 diabetes (Rosu et al., 2009), the presence of viable *Mycobacterium avium* subsp. *paratuberculosis* (MAP) cells in milk and dairy products is viewed as a potential food safety issue. MAP is the causative agent of Johne's disease, a contagious, chronic and sometimes fatal infection that primarily affects the small intestine of domestic ruminants such as cattle, sheep and goats (Stabel, 1998). Dairy cattle infected by MAP can shed live organisms in both milk and faeces, so potentially bulk milk could be contaminated either directly or indirectly by this bacterium. Whilst uncertainty remains about the public health/food safety significance of MAP, it would be prudent to monitor milk for presence of MAP to assess levels of contamination and hence potential for human exposure. The general desire is that our food should come from disease-free (including Johne's disease-free) animals (Regulation (EC) No 853/2004; European Parliament, 2004).

There is interest amongst dairy producers and processors to identify a rapid method that could be adopted to detect the presence of viable MAP in raw milk or dairy products. Whilst a number of conventional and quantitative PCR methods, able to detect and estimate numbers of MAP in milk have been developed in recent years (Timms, Gehringer, Mitchel, Daskalopoulos, & Neilan, 2011), the problem remains of differentiating whether a PCR positive result is due to DNA from viable or dead cells (Hanna, Connor, & Wang, 2005). More recently, use of a phage-based test (*FASTPlaque*TB^{TM} assay, Biotec Laboratories Limited, Ipswich), as a quicker alternative to culture, for rapid detection of viable MAP was described (Stanley et al.,

2007); its application to test naturally infected milk and milk products was subsequently reported (Botsaris, Liapi, Kakogiannis, Dodd, & Rees, 2013; Botsaris et al., 2010). This phage amplification method exploits the ability of a mycobacteriophage to replicate and amplify only within viable mycobacterial cells. The number of viable cells can be quickly estimated within 24-48 h based on the count of plaques (lysed areas) produced when infected cells burst in a lawn of *Mycobacterium smegmatis* (indicator) cells in an agar plate. We have previously optimised the original *FASTPlaque*TB^{TM} test conditions to maximise detection sensitivity and achieve accurate enumeration (Foddai, Elliott, & Grant, 2009). A magnetic separation step using paramagnetic beads coated with two MAP-specific peptide binders (aMp3 and aMptD; Stratmann, Dohmann, Heinzmann, & Gerlach, 2006; Stratmann, Strommenger, Stevenson, & Gerlach, 2002) was introduced before the phage assay to add specificity for MAP (Foddai, Elliott, & Grant, 2010b), since the phage involved has a broader host range (Rybniker, Kramme, & Small, 2006). The peptide-mediated magnetic separation (PMS) both captures and concentrates MAP cells, separating them from non-target microorganisms and other possible inhibitors, thereby increasing the specificity and sensitivity of the subsequent phage assay.

In light of the possibility of the PMS-phage assay being adopted by the dairy industry for milk testing, certain aspects of the milk testing protocol being applied at Queen's University Belfast were investigated to ensure that milk sample storage and preparation before testing were optimal. The following were examined: a) effect of milk storage conditions prior to commencement of testing on the viability of MAP cells, and identification of the best milk storage conditions to be applied when immediate analysis of milk samples is not possible; b) the partitioning of MAP to milk

pellet, cream and whey fractions after centrifugation, to identify which milk fraction should be tested to most accurately determine MAP contamination levels; and c) potential to introduce a de-clumping step before PMS to maximise accuracy of MAP counts obtained with the phage assay. The optimised phage assay was previously demonstrated to be a useful research tool for studies involving milk spiked with pure cultures of MAP (Foddai, Elliott, & Grant, 2010a), as it provides quantitative results for viable cells within 24 h. Therefore, the optimised phage assay was principally used throughout this study, although conventional culture on solid agar and qPCR were employed in some experiments to supplement or confirm phage assay results.

2. Materials and methods

2.1. Bacterial strains

Four MAP strains, including two type strains (ATCC 19698 and NCTC 8578) and two milk isolates (796PSS and 806R; Grant, Ball, & Rowe, 2002), were grown in a shaker incubator for 4–6 weeks at 37 ℃ to statio nary phase in Middlebrook 7H9 broth containing 10% OADC supplement (both from Difco) and 2 µg mL^{-1} mycobactin J (Synbiotics Europe SAS, Lyon, France). Cultures on solid agar for comparison of colony counts (cfu mL^{-1}) with plaque counts (pfu mL^{-1}) achieved by phage assay were obtained by spreading appropriate dilutions of *Mycobacterium avium* subsp. *paratuberculosis* onto Herrold's egg yolk medium (comprised of 9 g bacteriological peptone, 4.5 g sodium chloride, 15.3 g bacteriological agar, 2.7 g beef extract, 4.1 g sodium pyruvate, 27 mL glycerol, 5 mL 2% malachite green, 100 mL sterile egg yolk, 2 mg L^{-1} mycobactin J) and no antibiotics (HEYM). Agar plates were wrapped in

Duraseal laboratory sealing film (Diversified Biotech, MA, USA) and incubated at 37 ℃ for 4–6 weeks before counting colonies. *Mycobacterium smegmatis* mc^2 155, to be used as fast-growing sensor cells for the phage assay (see below) was cultured for 2–3 days at 37 ℃ to stationary phase in the same 7H9 medium without the addition of mycobactin J.

2.2. Optimised phage amplification assay

The optimised phage assay was carried out as previously described by Foddai et al. (2009). Briefly, after a previous overnight incubation of samples at 37 ℃ in 1 mL of 7H9 medium containing 2 mM $CaCl_2$, 100 µL D29 mycobacteriophage (10^8 pfu mL^{-1}) was added to samples before incubation at 37 ℃ for 2 h during which, particularly post-PMS samples, were shaken periodically every 20–30 min to keep uniform resuspension of paramagnetic beads. Virucide (100 mM ferrous ammonium sulphate (FAS, Sigma), 100 µL) was then added, samples were mixed thoroughly by accurate inverting and rolling (10 min at room temperature) in order to inactivate all exogenous seed phage before addition of 5 mL 7H9 medium containing 2 mM $CaCl_2$. The samples were returned to the incubator at 37 ℃ for a further 90 min before being plated onto agar plates prepared with 1 mL *Mycobacterium smegmatis* mc^2 155 (10^8 cfu mL^{-1}) and tempered 7H9 agar. Plaques were counted the following day after incubation of plates at 37 ℃ overnight.

2.3. Peptide magnetic separation

PMS was performed on 1 mL of sample using 5 µL biotinylated-aMp3 peptide- and 5 µL biotinylated-aMptD peptide-coated MyOne^{TM} Tosylactivated Dynabeads® (Life Technologies), prepared in-house as described previously (Foddai et al., 2010b). Magnetic separation was carried out using the Dynal BeadRetriever (Life Technologies) as described previously (Foddai et al., 2010b). Magnetic capture was carried out for 30 min at room temperature under continuous mixing, followed by two washes in 1 mL Phosphate buffered saline (PBS) containing 0.05% Tween 20 (PBS-T20, Sigma), and final resuspension of the beads in 1 mL 7H9 broth containing 10% OADC and 2 mM $CaCl_2$. For experiments involving PMS of spiked raw milk samples, 7H9 medium was also supplemented with NOA Antimicrobial Supplement (Product code 3A201N-300, Abtek Biological Ltd, Liverpool UK; final concentrations per mL^{-1} of broth: Nystatin 50 IU, Oxacillin 2 µg, Aztreonam 30 µg). NOA inhibited growth of any milk bacteria adhering to the paramagnetic beads after PMS without impacting viability of the MAP cells, or affecting subsequent phage assay.

2.4. Investigation of use of ultrasound sonication to achieve MAP de-clumping

The ultrasonic cleaning unit employed in this set of experiments was the Ultrasonic PH 30 (Fisher Scientific Ltd), which has two ultrasonic frequencies (37 kHz and 80 kHz) and three operation modes (Sweep, Pulse, Degas). Initially, 2 mL aliquots of broth suspensions containing approximately 10^5–10^6 MAP pfu mL^{-1} were processed through different ultrasonication treatments applied at ambient temperature (20–21 ℃), using the three operation m odes at 37 and 80 kHz, for increasing time periods (0 to 5 min). Conventional de-clumping method consisting of vortexing samples with five sterile 3 mm glass beads for 2 min, routinely used at

Queen's University Belfast (Rowe, Grant, Dundee, & Ball, 2000) to disperse clumps of MAP cells, was included as the control treatment for each experiment. Once the most promising ultrasonication setting had been established, the effect of ultrasonication treatment of samples on ice (suggested by Schleig et al., 2005), rather than directly in water bath at ambient temperature, was also evaluated. The effect of each de-clumping treatment was evaluated by comparing pfu, or cfu, counts obtained before and after treatment.

To investigate the potential for MAP cells to reclump quickly in samples not immediately processed after ultrasonication, 2 mL broth and whole UHT milk suspensions of four MAP strains containing approximately 10^5–10^6 cells mL^{-1} were de-clumped by the optimised ultrasonication treatment (identified during course of experiments outlined above), and then incubated for 0 min, 15 min, 30 min, 1 h and overnight (approximately 18 h) in a shaker incubator (100 rpm) at 37 ℃ before being processed in duplicate through the phage assay. If pfu counts recorded after each incubation time decreased then this was taken as evidence of re-clumping.

2.5. Assessment of the fractionation of MAP cells in spiked milk samples upon centrifugation

Raw bulk tank milk from herds without a previous history of Johne's disease or ultra-high temperature (UHT) processed milk purchased from a local supermarket was used for these experiments. Milk samples (10 mL raw and 50 mL UHT) were inoculated with broth suspensions of MAP to give an initial concentration of approximately 10^6 pfu mL^{-1}. Samples were centrifuged for 15 min at 2,500 × *g* in an Eppendorf centrifuge (model 5702) at room temperature to separate three milk

fractions (cream, whey, pellet). After determining the weight of each milk fraction, samples were de-clumped by optimised ultrasonication and processed immediately through the PMS-phage assay. For the cream and pellet fractions, which had been resuspended in 1 mL PBS-T20, the number of MAP cells was calculated as pfu counts per entire fraction. For the whey fraction the number of cells was determined by processing 1 mL of whey through the PMS-phage assay and then correcting the pfu count to take into account the total volume of the original whey fraction. The experiment was carried out in duplicate for each bacterial strain considered.

Quantitative PCR targeting both IS900 and f57 specific target-genes of MAP, used either alone (direct qPCR) or combined with prior PMS (PMS-qPCR), was carried out in parallel with the PMS-phage assay to verify the results obtained. DNA for samples processed through direct qPCR was extracted from each milk fraction as described by Hanifian, Khani, Barzegari, and Shayegh (2013). For samples to be processed through qPCR after PMS, bead samples were resuspended in a final volume of 50 µL RNAse/DNAse free water and DNA was released from captured cells by heating samples at 95 ℃ for 25 min. A 2.5 µL aliquot of the supernatant, after centrifugation to sediment beads, was used for each qPCR reaction. Quantitative PCR targeting both IS900 and f57 was performed as described previously by Donaghy, Johnson, and Rowe (2010). An internal amplification control, labelled with VIC at the 5'-end (EXO IPC - Exogenous Internal Positive Control Reagent) purchased from Applied Biosystems was included for each qPCR reaction. Comparison of EXO-IPC C_T values obtained in negative test control (NTC) wells and unknown samples was undertaken to assess the possible presence of PCR inhibitors in samples tested. Higher EXO IPC C_T values in unknown samples relative to NTC wells were taken as indicative of PCR inhibition. Each qPCR reaction was

performed in a final volume of 25 µL including: TaqMan Universal 2X PCR master mix (Applied Biosystems), EXO IPC 10× mix and EXO IPC 10× DNA (Applied Biosystems), 10 µM of each forward and reverse primer and 5 µM of specific probe (Donaghy et al., 2010), and 2.5 µL template DNA. A calibration curve with DNA released from known amount of MAP (10-10^5 pfu mL^{-1}) suspended in 7H9 broth was included with each qPCR run to allow quantification of numbers detected. qPCR reactions were performed using an Eco™ Real-Time PCR system (Illumina, Inc) with the following thermal cycling conditions: stage 1: 50 ℃ for 2 min; stage 2: 95 ℃ for 10 min and stage 3 (40 cycles): 95 ℃ for 15 s and 60 ℃ for 1 min.

2.6. Assessment of the effect of refrigeration, freezing, and use of preservative on numbers of viable MAP in milk

Whole UHT milk samples spiked with moderate (~10^4 cfu 50 mL^{-1}) and low levels (~10^2 cfu 50 mL^{-1}) of the four strains were processed through the PMS-phage assay applied after centrifugation at 2,500 × *g*, resuspension of milk pellet fraction in 1 mL PBS-T20 and de-clumping by optimised ultrasonication. Samples were divided between three storage categories: 1) fresh - samples processed immediately or after overnight at 4 ℃; 2) frozen - samples tested after 1 week, 2 weeks and 1 month at -70 ℃; and 3) chemically preserved - milk samples t reated with a milk preservative [1 mL of a solution containing 1% Bronopol (Sigma-Aldrich, UK) added to 50 mL spiked milk, (i.e., final concentration 10 mg 50 mL^{-1} milk)] and tested after 1 and 2 weeks storage at 4 ℃. Duplicate milk samples were tested for each strain under all experimental conditions. Counts (pfu 50 mL^{-1}) were compared for each storage condition relative to the starting bacterial spiking level; a decrease in pfu count was

considered to be an indication of loss of viability by a proportion of MAP cells present.

2.7. Statistical analysis of results

The statistical significance of the increase or reduction in plaque (pfu) and colony (cfu) counts observed for each treatment applied to broth and milk samples inoculated with viable MAP was assessed by a paired *t* test (Instat 3; GraphPad, La Jolla, CA); differences with $P < 0.05$ were considered significant. Correlation between numbers of MAP detected through PMS-phage assay and qPCR was assessed using linear regression analysis (Microsoft Excel).

3. Results

3.1. Use of ultrasonication to disperse MAP clumps

To determine the optimal ultrasonication treatment to disperse clumps of MAP cells various treatments were studied. Of the different sonication settings applied in a sonicator bath at room temperature, the greatest degree of de-clumping was consistently achieved in samples processed through ultrasonication using Pulse mode; mean $\log_{10}$ increase in pfu counts for MAP strains ATCC 19698 and NCTC 8578 were 0.17±0.02 and 0.23±0.08 ($P < 0.01$), respectively (Fig. 1A). Corresponding increases observed for samples de-clumped by vortexing samples with glass beads were 0.35±0.06 and 0.44±0.04, respectively ($P < 0.001$). Generally similar trends in terms of effects of each de-clumping treatment were observed with

265 cfu counts on HEYM (Fig. 1B), although log_{10} increases tended not to be as large as
266 observed with pfu mL^{-1} results.

267 Significantly greater de-clumping was achieved when ultrasonication was
268 applied to samples on ice in the sonicator bath ($P < 0.001$), rather than directly in the
269 water bath at ambient temperature (Fig. 2A). This was the case for all strains with
270 the exception of MAP strain 806R which showed considerable variability in counts
271 (large error bar) when vortexed with glass beads. Mean log_{10} increase in pfu counts
272 observed for two type strains (ATCC 19698 and NCTC 8578) and two milk isolates
273 (796PSS and 806R) were 0.90±0.08, 0.87±0.14, 0.36±0.03 and 0.51±0.08,
274 respectively, with overall mean of 0.66±0.08. The corresponding mean log_{10} increase
275 in pfu counts observed for samples de-clumped by vortexing with glass beads was
276 lower for each MAP strain (0.60±0.10, 0.44±0.04, 0.20±0.02 and 0.37±0.30,
277 respectively, with overall mean log_{10} increase in pfu of 0.40±0.12, $P<0.01$). No
278 significant increase in pfu counts was observed for samples subjected to extended
279 incubation on ice for 5 or 10 min before the ultrasonication treatment ($P > 0.05$ in
280 both cases). Generally, similar trends in terms of effects of each de-clumping
281 treatment were observed with cfu counts on HEYM (Fig. 2B), although log_{10}
282 increases tended not to be as large as observed with pfu mL^{-1} results.

283 The possibility of MAP cells re-clumping quickly after sonication treatment
284 was estimated by comparing counts recorded from samples enumerated at various
285 times after de-clumping. Mean pfu counts recorded for spiked milk samples
286 immediately after de-clumping and after each incubation time did not vary
287 significantly ($P = 0.96$), suggesting that no re-clumping occurred (Fig. 3A). Similar
288 results were obtained for MAP broth cultures processed through the phage assay
289 started after 0 min, 15 min, 30 min and 1 h ($P = 0.06$, Fig. 3B). However, a significant

290 increase in pfu mL^{-1} counts (~ 2 log_{10}) was consistently recorded for MAP broth
291 cultures incubated overnight in 7H9-OADC-2 mM $CaCl_2$ at 37 ℃ before the phage
292 assay commenced (Fig. 3B). This substantial increase in pfu mL^{-1} was not
293 unexpected, as it is consistent with our previous finding (Foddai et al., 2009) that
294 there is enhanced phage infection of MAP cells and increased pfu counts when they
295 have been incubated overnight in the presence of 2 mM $CaCl_2$.
296
297 *3.2. Fractionation of MAP cells within milk upon centrifugation*
298
299 Upon centrifugation of 10 or 50 mL volumes of MAP spiked milk at 2500 × *g*
300 for 15 min, the vast majority of the MAP cells in both raw and UHT homogenised milk
301 were consistently recovered from the milk pellet. Mean log_{10} pfu observed from both
302 homogenised and non-homogenised milk inoculated with various MAP strains was
303 6.28±0.25 and 6.38±0.06, respectively, corresponding to approximately
304 81.53±5.76% and 96.7±1.7%, respectively, of the initial inocula. The percentage of
305 bacterial cells recovered from the milk whey fraction ranged from 11.4 to 25.6%
306 (overall mean 18.4±5.8%) for UHT homogenised milk and 0.4 to 2.9% (mean
307 1.6±0.21%) for raw milk. The percentage of MAP recovered from the cream was
308 lower than expected, on the basis of previous studies, in both cases; overall mean
309 observed was 1.6±0.21% for whole raw and 0.06±0.02% for UHT homogenised milk.
310 No significant difference in fractionation between MAP strains employed to spike raw
311 and UHT milk was observed ($P > 0.05$).
312 Numbers of MAP in each milk fraction estimated via direct-qPCR were more
313 variable than PMS-qPCR, but similar trends in numbers of MAP cells per fraction
314 were generally observed. Lack of sensitivity was observed when qPCR was applied

alone on DNA extracted from each raw milk fraction after centrifugation; C_T values obtained from EXO IPC assay from samples were generally higher than those obtained from negative test control (NTC) samples indicating the presence of PCR inhibitors. Numbers of MAP estimated through IS900 and f57 direct-qPCR were consistently lower (by 1–2 $\log_{10}$) than corresponding values achieved through PMS-phage assay and PMS-qPCR ($P < 0.05$ and $P < 0.01$, respectively). Best correlation was observed between qPCR and phage assay results ($r^2 = 0.8568$ for IS900 qPCR and $r^2 = 0.7631$ for f57 qPCR) when PMS was applied before qPCR for both types of spiked milk samples (Fig. 4A,B). No significant variance of EXO-IPC C_T values for all post-PMS DNA samples relative to NTC wells was observed, indicating apparent absence of PCR inhibitors in these samples.

3.3. Effect of various milk storage conditions pre-testing on MAP viability

The effect of refrigeration, freezing, and use of preservative on numbers of viable MAP in milk was evaluated. Counts (pfu) detected from samples processed immediately or after one overnight at 4 ℃ were similar ($P > 0.05$, Fig. 5A,B), suggesting that milk analysis can be performed on fresh or overnight refrigerated milk samples without adverse effect on MAP counts. Minimal reduction in numbers of MAP detected ($P > 0.05$) was observed from samples refrigerated for 3 days before milk analysis. Mean $\log_{10}$ decrease in pfu counts for both moderate-spiked (Fig. 5A) and low-spiked (Fig. 5B) milk samples was 0.41±0.08 and 0.15±0.10, respectively. No adverse effect on pfu counts ($P > 0.05$) was also observed from milk samples previously frozen at -70 ℃ for 1 or 2 weeks and 1 month before testing. Log_{10} decrease in pfu counts ranged from 0.29±0.23 to 0.52±0.20 $\log_{10}$ for moderate (Fig.

5A), and from 0.01±0.20 and 0.20±0.24 for low-spiked level (Fig. 5B), suggesting minimal impact of frozen storage at -70 ℃ on viability of MAP cells. In contrast, addition of Bronopol, as a milk preservative, in combination with refrigeration at 4 ℃ for 1 and 2 weeks was found to severely impact viability of MAP cells. In the case of one MAP strain (806R) viability was so adversely affected that no viable MAP were detected in low spiked milk when testing was preceded by refrigeration for 2 weeks in the presence of Bronopol (Fig. 5B). Counts (pfu) observed for milk samples processed after 1 and 2 weeks were consistently 1-2 $\log_{10}$ lower than those observed from fresh samples (Fig. 5A,B). Mean $\log_{10}$ decrease in pfu counts observed in moderate and low-spiked milk samples was 1.96±0.38 and 1.04±0.20 after 1 week ($P < 0.01$) and 2.25±0.56 and 1.70±0.40 after 2 weeks ($P < 0.001$), respectively.

4. Discussion

The objective of this study was to develop an optimised protocol for sample processing and PMS-phage assay testing of raw milk samples to ensure rapid detection and accurate enumeration of viable MAP, so a number of aspects of the current milk testing protocol were investigated. The first part of the study focused on the development of a non-invasive de-clumping method to maximise accuracy of MAP cells obtained through PMS-phage assay. Due to the hydrophobic nature of its cell wall, MAP cells tend to aggregate in clumps, the presence of which in test milk samples can lead to underestimates of MAP numbers by a factor of 100-1000 cells (Klijn, Herrewegh, & de Jong, 2001; Pearce et al., 2001). Various de-clumping

options, including repeated syringing (Keswani & Frank, 1998; Sung & Collins, 1998), vortexing with glass beads (Rowe et al., 2000), and sonication (Chiodini & Hermon-Taylor, 1993; Stabel, Steadham, & Bolin, 1997) have previously been applied in the preparation of MAP inocula by other researchers. The method routinely applied at Queen's University Belfast involves vortexing the MAP culture with five sterile 3 mm glass beads for 2 min. However, the sterile glass beads would generally be added to 7H9 broth at the time of preparation, not at the point of de-clumping, which would be the case in the context of milk testing. Introduction of glass beads to each test milk sample would be laborious and may increase the risk of introducing exogenous MAP DNA to the milk sample, if this approach were to be taken. A similar situation would exist in the case of syringing, the use of which would also require extra costs for additional sterile consumables. For these reasons, only treatment of samples in sealed tubes in a sonicator bath would seem to be a realistic proposition. The impact of different sonicator bath power settings and treatment temperatures and times on the viability of MAP cells was evaluated. Amongst the different sonication settings involving low-frequency ultrasound treatment (37 and 80kHz) at ambient temperature, no significant de-clumping was observed. A small increase in MAP numbers compared to vortexing with glass beads (control treatment) suggested a minimal effect on disruption of MAP clumps. With the possible exception of one of the MAP field strains (MAP 806R), ultrasonication at 37kHz on ice achieved significantly greater de-clumping of MAP suspensions than the control method (vortexing with glass beads). When grown in broth, 806R forms particularly large aggregates of cells and generally proves difficult to de-clump and achieve a visually uniform suspension; the large error bars for vortexing of MAP 806R (Fig. 2A) is reflective of this situation. Overall, no adverse effect of de-

clumping by ultrasonication at 37 kHz in Pulse mode for 4 min on the viability of MAP cells was observed.

Milk samples to be tested for MAP generally require initial centrifugation to reduce large volumes (20–50 mL) to a 1 mL test volume appropriate for PMS. The PMS-phage assay and qPCR results obtained during the present study indicate that the milk pellet is the location of maximal numbers of MAP after centrifugation of a milk sample at 2,500 × *g* for 15 min at ambient temperature. Mean percentage recovery of MAP in the pellet estimated based on pfu counts for spiked homogenised UHT milk and spiked raw milk were >80% and 95%, respectively. Considerably lower proportions of MAP cells were generally recovered from the whey (18.4% and 1.6% for spiked homogenised UHT and raw milk, respectively) and cream (0.06% and 1.6% for spiked homogenised UHT and raw milk, respectively) fractions. Thus, in the case of raw milk samples negligible numbers of MAP would be lost if the whey and cream fractions were discarded after centrifugation; the picture for homogenised milk seems to be rather different. Comparison of results achieved by the two detection methods showed a better correlation between phage assay and PMS-qPCR results than direct qPCR results; some evidence of PCR inhibition was obtained for direct qPCR, but not for PMS-qPCR. PMS was, therefore, confirmed to be an effective method to achieve selective concentration of target cells plus removal of PCR inhibitors from test samples. Two previous fractionation studies carried out on raw milk inoculated with MAP showed that the vast majority of the original inoculum sediment to the milk pellet after centrifugation (Grant, Ball, & Rowe, 1998; Van Brandt et al., 2010). Another study (Gao, Odumeru, Raymond, & Mutharia, 2005) reported that MAP cells preferentially partitioned to the cream fraction upon centrifugation, leading to a recommendation that the pellet and cream fractions

should be combined for MAP testing of milk. The centrifugation conditions adopted by Van Brandt et al. (2010; 974 × *g* for 36 min at 4 ℃) and Gao et al. (2005; 3100 × *g* for 30 min, temperature not stated) differ from those used by Grant et al. (1998) and in the current study (2,500 × *g* for 15 min at room temperature). In particular, temperature of centrifugation, or temperature of milk at time of centrifugation, may have impacted the differing fractionation profiles observed. When processing raw milks in our laboratory, we purposely allow milk samples to attain ambient temperature before starting to test and always centrifuge at ambient temperature (not at 4 ℃) to ensure that sedimentation of MAP cells to the pellet is maximised.

Another important consideration for the establishment of an optimised milk testing protocol was to discover the best milk storage condition to be adopted for milk samples if milk analysis cannot start immediately upon delivery of samples. Raw milk samples for MAP testing are usually collected miles away from the testing laboratory, and refrigeration, use of milk preservatives, or freezing samples are some of the methods used to preserve milk samples after collection or upon delivery. As the phage assay exploits the ability of D29 mycobacteriophage to replicate only within viable mycobacterial cells, maintaining the viability of MAP cells in test samples is critical. To the best of our knowledge, no information is currently available in the literature on how storage conditions including refrigeration, freezing, and use of preservative affect viability of MAP cells in milk. Results of the present study indicate that processing milk on the same day as collection or after overnight storage at 4 ℃ are the ideal conditions to maximise detection and achieve accurate enumeration of viable MAP cells. If this is not possible, then freezing of milk samples at -70 ℃ for up to one month could be alternatively applied with minimal impact on viable MAP counts. In contrast, the addition of milk preservative (Bronopol) to milk

samples which are then refrigerated for 1 or 2 weeks at 4 ℃ weeks should be avoided as these conditions significantly affected viability of MAP; this could lead to underestimation of the number of MAP cells originally present in milk samples, or false negative results if $<10^2$ MAP cells are present per 50 mL milk. Our findings are in general agreement with the literature available on the subject. Use of milk preservatives, particularly Bronopol, have already been reported to cause bacterial membrane damage affecting both Gram-negative (*Escherichia coli* and *Pseudomonas aeruginosa*) and Gram-positive (*Staphylococcus aureus*) bacteria (Shepherd, Waigh, & Gilbert, 1988; Stretton & Manson, 1973). This might explain the reduction in bacterial count as observed in our study and in other milk surveys previously carried out (Amores et al., 2010; Pinnow et al., 2001). Freezing of milk samples at -80 ℃ without adverse effect on the via bility of bacterial cells has previously been also reported for other pathogenic bacteria, including coagulase-negative staphylococci and Gram-negative bacilli (Sanchez et al., 2003).

5. Conclusions

Milk samples should be refrigerated at 4 ℃ after c ollection and MAP testing should commence within 24 h, or, when this is not possible, samples can be frozen at -70 ℃ for up to one month without significant l oss of MAP viability. The vast majority (>95%) of MAP in raw milk were found to sediment to the pellet fraction upon centrifugation at 2500 × *g* for 15 min, so this milk fraction should be tested. De-clumping of MAP aggregates before PMS is most effectively achieved by ultrasonication of the resuspended milk pellet on ice in a sonicator bath at 37 kHz for 4 min in ‘Pulse’ mode. Our findings regarding optimal milk storage and preparation

prior to MAP testing by the PMS-phage assay should theoretically be equally applicable to milk samples to be cultured; however the effect of chemical decontaminants on viability of MAP cells that had been previously frozen and thawed would be an additional consideration.

Acknowledgments

The authors acknowledge financial support from the Nestlé Research Center, Lausanne, Switzerland, to pursue this research.

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Figure legends

Fig. 1. Comparison of the de-clumping effects of vortexing with glass beads for 2 min (Control) and three ultrasonication treatments (Pulse Mode at 37 kHz for 2, 3 or 4 min). Middlebrook 7H9 broth cultures of various MAP strains containing 10^5–10^6 pfu mL^{-1} were placed in a steel basket and processed in a sonicator bath at ambient temperature (20 ℃). MAP was enumerated by (A) phage assay (pfu mL^{-1}) and (B) culture on HEYM (cfu mL^{-1}). The larger the increase in $\log_{10}$ pfu mL^{-1} observed, the greater the de-clumping effect of the treatment. For each MAP strain results are mean ± standard deviation of two experiments with duplicate samples per experiment.

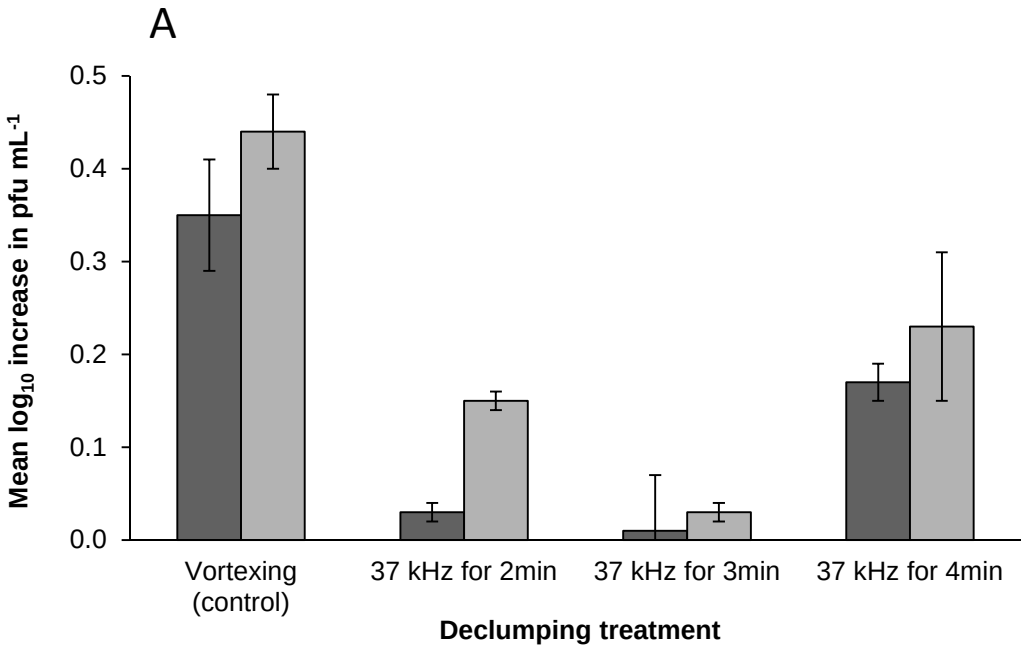
Fig. 2. De-clumping effects of ultrasonication (Pulse Mode at 37kHz for 4 min) on ice compared with vortexing with glass beads for 2 min (Control). Middlebrook 7H9 broth cultures of four MAP strains containing 10^5–10^6 pfu mL^{-1} were processed in a sonicator bath on ice. MAP was enumerated by (A) phage assay (pfu mL^{-1}) and (B) culture on HEYM (cfu mL^{-1}). The larger the increase in $\log_{10}$ pfu or cfu mL^{-1} observed, the greater the de-clumping effect of the treatment. For each MAP strain results are mean ± standard deviation of two experiments with duplicate samples per experiment. Bars represent (left to right): vortexing with glass beads for 2 min (control, ■), ultrasonication on ice at 37 kHz for 4 min (■), incubation for 5 min on ice before ultrasonication on ice at 37 kHz for 4 min (■), and incubation for 10 min on ice before ultrasonication on ice at 37 kHz for 4 min (■).

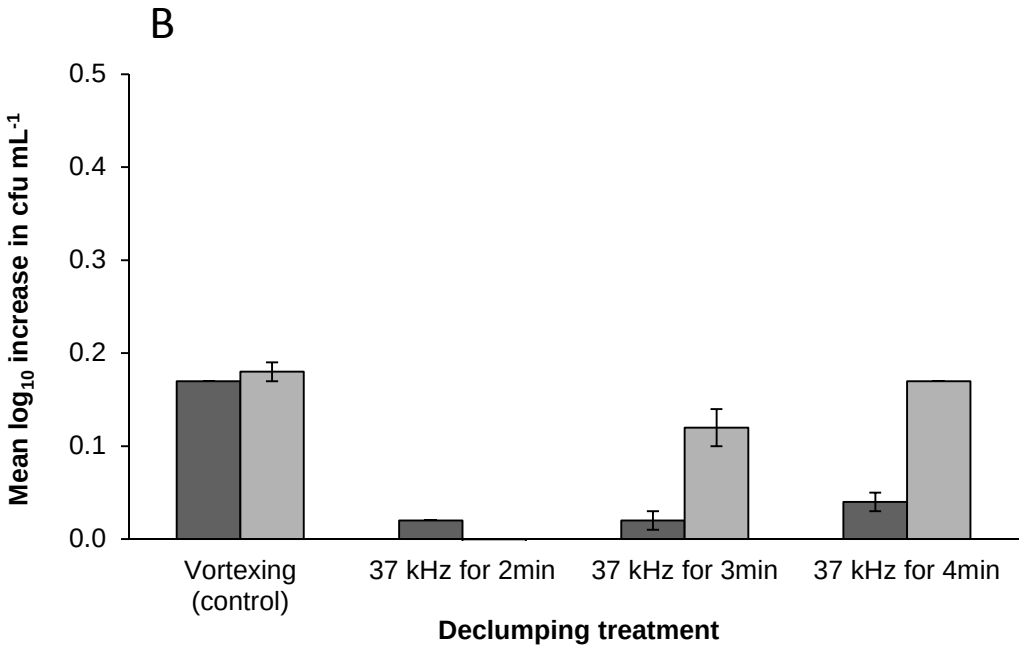
Fig. 3. Investigation of possible re-clumping of MAP cells in (A) spiked milk samples and (B) Middlebrook 7H9 broth cultures after ultrasonication treatment. Samples were processed through the phage amplification assay immediately after ultrasonication (0 min, ■) and at increasing times (15 min, ■; 30 min, ■; 1 h, ■; overnight at 37 ℃, ■) post-ultrasonication. No significant decrease in MAP count, which would have been indicative of re-clumping, was observed. For each MAP strain, $\log_{10}$ pfu mL^{-1} results are mean ± standard deviation of two experiments with duplicate samples per experiment.

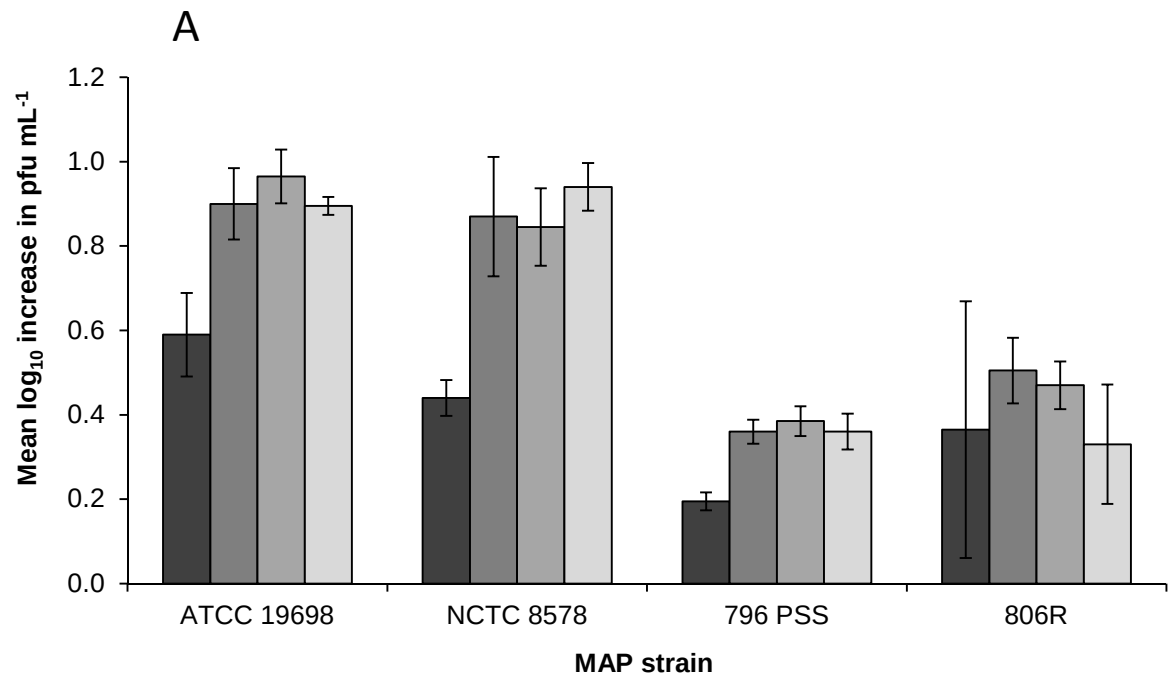
Fig. 4. Relationship between MAP counts obtained by PMS-phage assay and corresponding numbers of MAP estimated by (A) PMS-IS900 qPCR and (B) PMS-f57 qPCR for cream (white), whey (grey) and pellet (black) fractions after centrifugation of 10 mL raw milk (circles) and 50 mL homogenised UHT milk (diamonds) originally spiked with approximately 10^6 pfu MAP mL^{-1}. Each data point represents the mean of two samples.

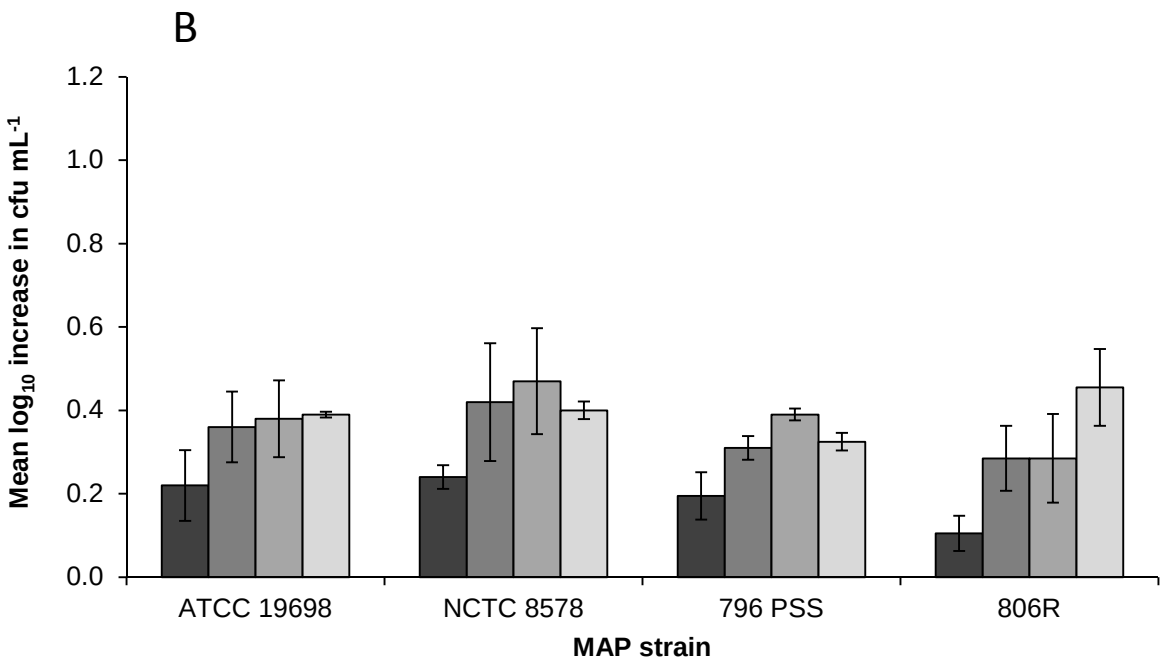
Fig.5. Impact of pre-testing storage conditions on numbers of viable MAP in milk samples spiked with 10^4 pfu 50 mL^{-1}. Four different MAP strains were used for spiking experiments. Milk samples were processed through the PMS-phage assay immediately after spiking and after different pre-testing storage conditions: (A) refrigeration at 4 ℃, with and without addition of Bronopol preservative, and (B) freezing at -70 ℃. For each MAP strain results are mean ± standard deviation of two experiments with duplicate samples per experiment. Bars represent (left to right): immediate testing (■), 4 ℃ overnight (■), 4 ℃ for 3 days (■), 1 week at 4 ℃ plus

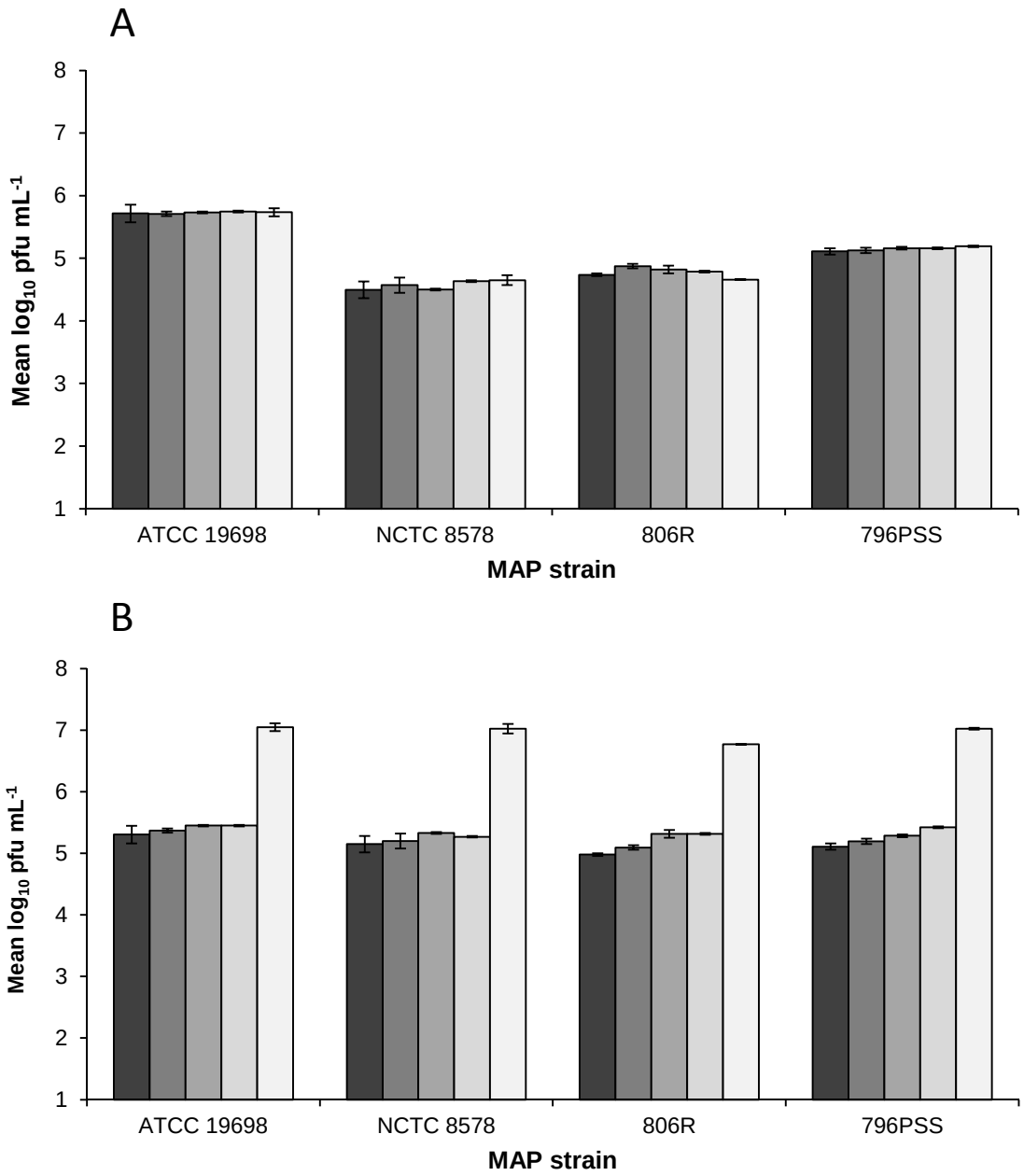
Bronopol (■), 2 weeks at 4 ℃ plus Bronopol (■), 1 week at -70 ℃ (■), 2 weeks at -70 ℃ (□), and 1 month at -70 ℃ (⊡).

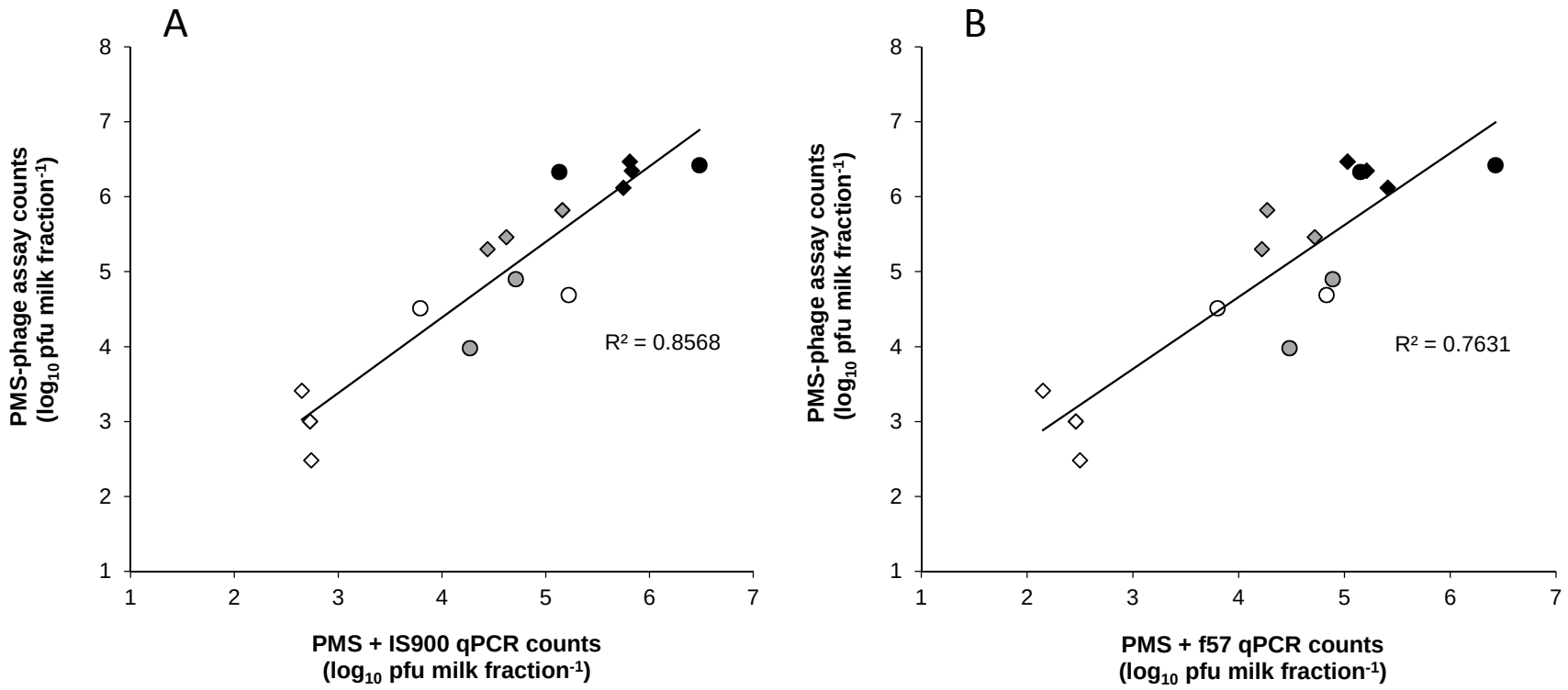

A
0.5
0.4
0.3
0.2
0.1
0.0
Mean log10 increase in pfu mL-1
Vortexing (control)
37 kHz for 2min
37 kHz for 3min
37 kHz for 4min
Declumping treatment

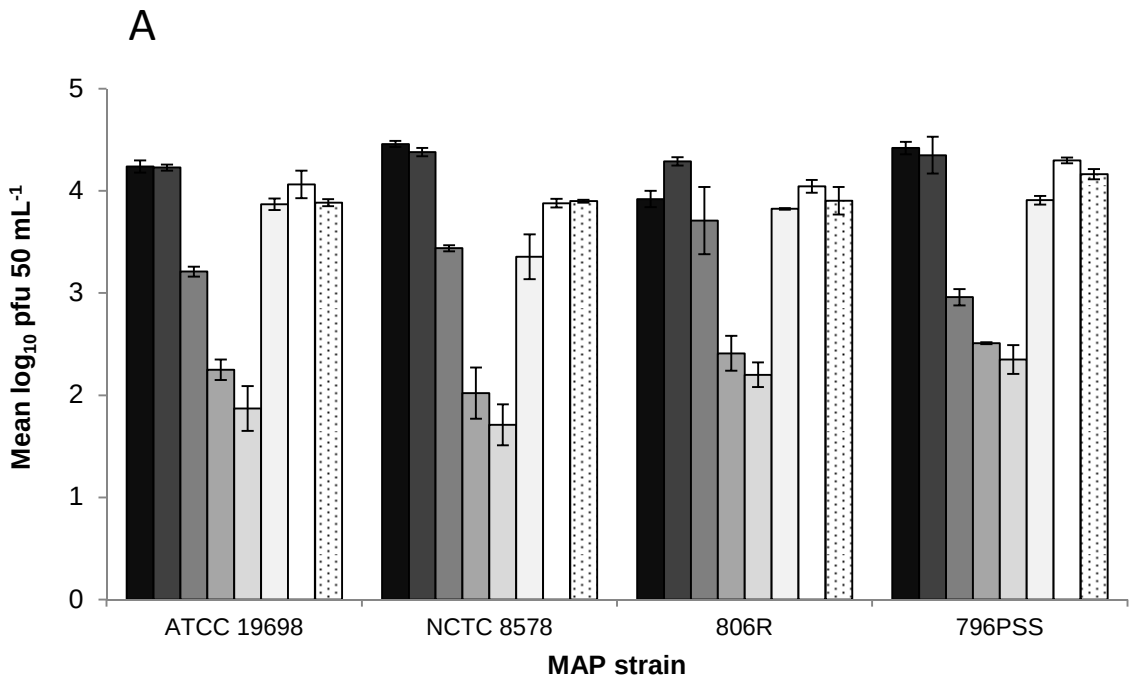

B
0.5
0.4
0.3
0.2
0.1
0.0
Mean log10 increase in cfu mL-1
Vortexing (control)
37 kHz for 2min
37 kHz for 3min
37 kHz for 4min
Declumping treatment

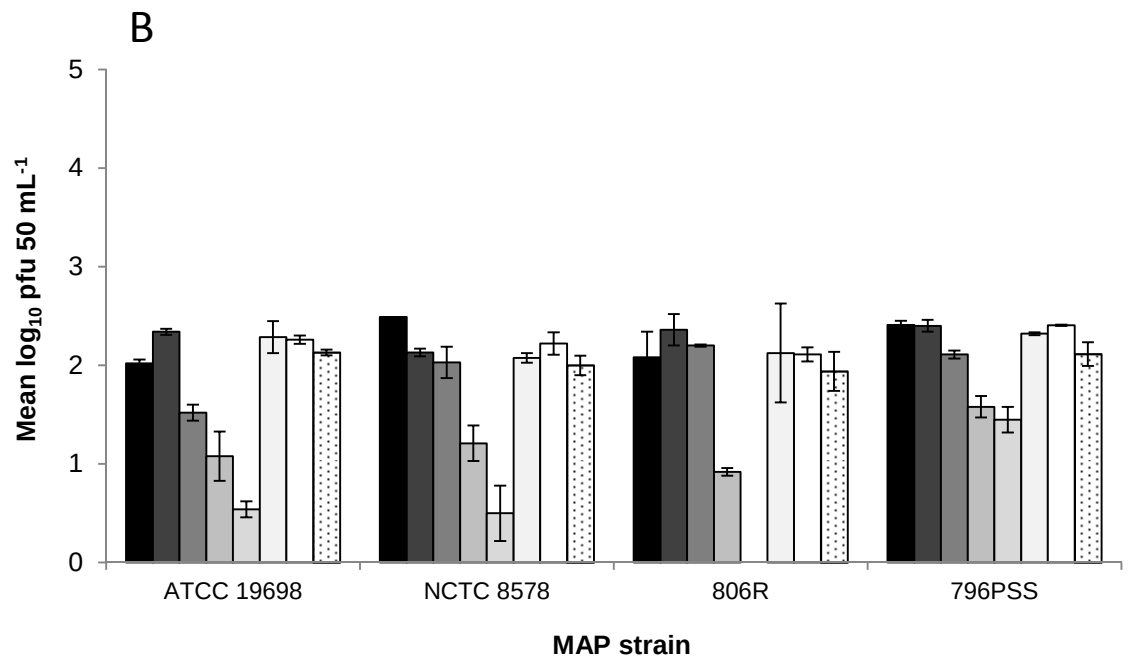

A
1.2
1.0
0.8
0.6
0.4
0.2
0.0
Mean log10 increase in pfu mL-1
ATCC 19698
NCTC 8578
796 PSS
806R
MAP strain


B
1.2
1.0
0.8
0.6
0.4
0.2
0.0
Mean log10 increase in cfu mL-1
ATCC 19698
NCTC 8578
796 PSS
806R
MAP strain


A
8
7
6
5
4
3
2
1
Mean log10 pfu mL-1
ATCC 19698
NCTC 8578
806R
796PSS
MAP strain
B
8
7
6
5
4
3
2
1
Mean log10 pfu mL-1
ATCC 19698
NCTC 8578
806R
796PSS
MAP strain


A
PMS-phage assay counts
(log10 pfu milk fraction-1)
PMS + IS900 qPCR counts
(log10 pfu milk fraction-1)
R² = 0.8568
B
PMS-phage assay counts
(log10 pfu milk fraction-1)
PMS + f57 qPCR counts
(log10 pfu milk fraction-1)
R² = 0.7631


A
Mean log10 pfu 50 mL-1
0
1
2
3
4
5
ATCC 19698
NCTC 8578
806R
796PSS
MAP strain


B
Mean log10 pfu 50 mL-1
0
1
2
3
4
5
ATCC 19698
NCTC 8578
806R
796PSS
MAP strain