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The role of anaerobic bacteria in the cystic fibrosis airway

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Abstract (word count, 192)

Purpose of review

Anaerobic bacteria are normal commensals but are also considered opportunistic pathogens and have been identified as persistent members of the lower airway community in people with cystic fibrosis (CF) of all ages and stages of disease. Currently, the role of anaerobic bacteria in CF lower airway disease is not well understood. Therefore, this review describes the recent studies relating to the potential pathophysiological role(s) of anaerobes within the CF lungs.

Recent findings

The most frequently identified anaerobic bacteria in the lower airways are common to both CF and healthy lungs. Studies have shown that in CF the relative abundance of anaerobes fluctuates in the lower airways with reduced lung function and increased inflammation associated with a decreased anaerobic load. However, anaerobes found within the lower airways also produce virulence factors, may cause a host inflammatory response and interact synergistically with recognised pathogens.

Summary

Anaerobic bacteria are potentially members of the airway microbiota in health but could also contribute to the pathogenesis of lower airway disease in CF via both direct and indirect mechanisms. A personalised treatment strategy that maintains a normal microbial community may be possible in the future. **Key words:** anaerobes, microbial community, respiratory infection.

Introduction

Cystic fibrosis (CF) is the most common lethal inherited condition affecting the Caucasian population. Clinical characteristics are caused by the absence or reduction of cystic fibrosis transmembrane conductance regulator (CFTR) protein function, which perturbs ion channels at the apical membrane of epithelial cells and results in a multisystem disease. Disease of the lower airways is considered the most important manifestation as it is the cause of morbidity and mortality in a large percentage of people [1].

Lower airway disease is secondary to chronic bacterial infection and a dysregulated host inflammatory response that is dominated by neutrophils [2]. Currently, a limited range of bacteria which cause lower airway infection, including *Staphylococcus aureus*, *Haemophilus influenzae*, *Pseudomonas aeruginosa* and *Burkholderia cepacia* complex, have been correlated with worse outcomes of people with CF (Figure 1) [3]; therefore, they are recognised as CF pathogens and are usually targeted by antibiotic treatment regimens [4]. These pathogens are routinely detected using aerobic culture-based methods [5]; however, such methods fail to culture obligate anaerobic bacteria whilst non-target facultative anaerobic microorganisms isolated are often considered "oropharyngeal flora".

During the past decade, unexpected bacterial community richness (number of species) within the CF lower airways has been demonstrated [6-12]. Furthermore, anoxic niches are known to exist within CF sputum, which supports the survival of anaerobic bacteria in the lower airways [13,14]. Many anaerobic species detected in CF respiratory samples are also present in the microbiota of the oral cavity and in a recent study, periodontal-associated

anaerobic bacteria were found in plaque and sputum samples of some people with CF [15*]. Therefore, detection of oral-associated bacteria in the lungs may be attributed to migration from the oral cavity to the lower airways [16-18].

Despite the accumulating knowledge of the range and abundance of bacteria in the diverse, dynamic and spatially heterogeneous lower airway communities, the contribution of obligate and non-target facultative anaerobic bacteria (henceforth referred to collectively as "anaerobic bacteria") to the pathogenesis of CF lower airway disease is currently not well understood. In fact, there remains much deliberation about whether anaerobic bacteria are pathogenic in the CF lower airways, commensals with negligible clinical impact or members of a "normal" airway microbiota. This review focuses on the most recent knowledge pertaining to the potential role of anaerobes detected within the polymicrobial communities of the CF lungs.

Potential pathophysiological role of anaerobic bacteria in chronic lower airway infection

Ecological analyses generally describe that a reduced bacterial diversity, including decreased richness of anaerobes, leads to dominance of a recognised pathogen and is associated with antibiotic use, increasing age and poorer lung function [19-22]. Furthermore, it has been hypothesied that certain anaerobic bacteria potentially form part of a normal airway microbiota; the most frequently identified anaerobic genera (*Prevotella*, 83%; *Veillonella*, 54%) in CF sputum were also commonly detected in induced sputum of healthy controls [23*].

Variation in the abundance of anaerobic bacteria: impact on clinical status

It was recently shown that worse lung function (measured by lung clearance index) and increased inflammation (measured by blood C-Reactive Protein) were associated with a lower total load of anaerobic bacteria in clinically stable people with CF (Figure 2) [23*]. This suggests that a more diverse lung microbiota is potentially associated with health whilst a lower load of anaerobic bacteria may reflect microbiota disruption and disease progression (Figure 2) [23*]. However, some anaerobes in low abundance can have an adverse impact on disease progression, as has been demonstrated in periodontal disease [24].

Conflicting reports of variability in microbial community composition at the onset of an exacerbation exist with some studies reporting within-patient fluctuation whilst others finding little change compared with other time-points [9,25,26]. Recently there has been interest pertaining to microbial physiology within the polymicrobial community of the CF lower airways and using a novel microbial culture system, the physiochemical properties of the microbiota in sputum were investigated [27**]. Increased gas bubbles and reduced pH in glass capillary tubes, indicators of fermentative metabolism, at the onset of a pulmonary exacerbation coincided with a high abundance of fermentative anaerobes in a single patient with CF [27**]. Treatment with antibiotics raised the pH and reduced gas production and the abundance of anaerobes [27**].

It was also proposed that fluctuations in the abundance of specific anaerobic bacterial genera or species within the microbial community could potentially be utilised as a surrogate marker to indicate clinical status [20,28,29*]. This

includes the obligate anaerobes, *Porphyromonas* and *Prevotella* species, which when present in high relative abundance were associated with better lung function (measured by FEV₁) [28]. In contrast, an increase in the abundance of the facultative anaerobe, *Streptococcus anginosus* group, correlated with poorer lung function [28]. Furthermore, detection of *Streptococcus pyogenes* in high numbers was associated with acute pulmonary exacerbations in a small number of adults (n=15) with CF [30] whilst *Veillonella* and *Prevotella* species were predicted to represent potential surrogate markers for the onset of a pulmonary exacerbation in another recent study [29*].

Genetic adaptation and antimicrobial resistance

It is evident that anaerobic bacteria persist in the CF lower airway community, even under continuous antibiotic pressure targeting recognised pathogens [31]. In order to become established members of a bacterial community, bacteria may undergo genetic adaptation to a specific niche. For example, various clonal lineages of *P. aeruginosa* are known to accumulate mutations in genes that enable adaptation of the bacterium to the dynamic CF airways including mutations that increase resistance to antibiotics or affect virulence traits (described as "pathoadaptation") [32]. Anaerobes from the CF lower airways are also more resistant to antibiotics when compared to similar anaerobic genera from the airways of healthy control subjects [33,34]. Therefore, this may indicate that anaerobic bacteria are established members

of the lower airway community and are undergoing adaptation to antibiotic selective pressure within the lungs.

Some strains of *P. aeruginosa* are associated with worse patient outcomes and similar patterns have been reported for periodontal microorganisms where differences in pathogenicity are observed between strains of the same species [35,36]. Therefore, further work is warranted to investigate genetic adaptations and diversity of particular anaerobic species in the CF lower airway environment.

Potential virulence mechanisms

Anaerobic bacteria are normal commensals but are also considered opportunistic pathogens, often involved in polymicrobial infections. Therefore, it is plausible that anaerobic bacteria could constitute members of the normal microbiota in health but also conceivable that they could contribute to the pathogenesis of chronic lung disease in people with CF via direct or indirect mechanisms. Recent evidence from a number of studies demonstrate that anaerobes may cause a host inflammatory response, interact synergistically with recognised CF pathogens and protect recognised pathogens from antibiotics by enhanced antimicrobial resistance (Figure 3).

Inflammation of the lower airways

Anaerobic bacteria in the CF lower airways may contribute to the dysregulated inflammatory response and subsequent, tissue destruction within the lungs [37**]. Bacteria including *Prevotella melaninogenica*, *Actinomyces odontolyticus*, *Veillonella parvula*, *Fusobacterium nucleatum* and

Streptococcus sanguinis secrete various short chain fatty acids (e.g. acetic acid, propionic acid, butyric acid) *in vitro*, which were also detected in micromolar amounts in bronchoalveolar lavage (BAL) fluid [37**]. Notably, high levels of acetic acid in BAL samples correlated with elevated levels of the pro-inflammatory cytokine interleukin-8 (IL-8), which is an important mediator of neutrophil recruitment and activation [37**]. The short-chain fatty acids were found to contribute to the excessive release of IL-8 by CF bronchial epithelial cells via a specific receptor (GPR41) that was overexpressed on the surface of the CF cells compared to non-CF bronchial epithelial cells [37**]. Another study also detected short-chain fatty acids in sputum of people with CF, which influenced host inflammatory responses and nitric oxide production and when present in low concentrations, improved the growth of *P. aeruginosa in vitro* [38**].

Interaction with other microorganisms in polymicrobial communities

Anaerobic bacteria may influence the progression of CF lower airway disease by inter-species interactions that enhance the pathogenicity or colonisation ability of recognised pathogens within the lower airways. In fact, it was previously hypothesised that certain anaerobes might create a favourable microenvironment for recognised CF pathogens to colonise, cause infection and subsequently dominate the lower airways (Figure 2) [27**].

It was also reported that co-infection of *P. aeruginosa* with *V. parvula*, in an *in vivo* murine model, resulted in the recovery of a significantly higher load of *P. aeruginosa* and greater clinical deterioration compared to infection with *P. aeruginosa* alone [39*]. Conversely, pyocyanin-producing *P. aeruginosa*

isolates may increase the virulence of obligate anaerobes. A study found that increased mortality was observed when mice were intra-tracheally inoculated with *Porphyromonas gingivalis* and a dose of pyocyanin at a concentration found in the CF airways compared to the anaerobe alone [40].

Biofilm formation is a feature of chronic airway infection in CF and it was shown that *P. aeruginosa* grows as a mixed biofilm *in vitro* with various *Streptococcus* species (e.g. viridans streptococci, *Streptococcus milleri* group) [41*,42]. Virulence factor production by *P. aeruginosa* was increased when grown as a mixed biofilm compared to without *Streptococcus* species [41*]. Furthermore, in patients with bronchiectasis it was reported that detection of non-tuberculous mycobacteria (NTM) in BAL samples was associated with a higher prevalence of *Prevotella* species compared to non-NTM infected patients [43]. Therefore, *Prevotella* species may affect colonisation of NTM in chronic lung infections.

Enhanced antibiotic resistance

A high proportion of CF-associated *Prevotella* species were previously shown to produce β -lactamases [33]. In polymicrobial infections, β -lactamaseproducing bacteria may protect themselves and other members of the community from antimicrobial agents (the concept of "passive resistance") [44]. A recent study found that a β -lactamase-producing *Prevotella* species was able to shield *P. aeruginosa* from the activity of ceftazidime *in vitro*, which is used to target the pathogen during treatment of pulmonary exacerbations [45*]. Therefore, enabling *P. aeruginosa* to persist in the presence of ceftazidime may be an indirect method by which *Prevotella* species contribute to disease pathogenesis.

Should anaerobes be targeted by treatment?

There is currently a lack of evidence to conclude if anaerobic bacteria should be targeted when treating CF lower airway infection. Targeted treatment of bacteria, residing within the lungs, is also challenging with the types and formulations of antibiotics that are currently available. Anaerobic bacteria are generally susceptible to some broad-spectrum antibiotics that are routinely used in the management of CF lower airway infection (e.g. meropenem, piperacillin/tazobactam) [33]; therefore, it is likely that anaerobes are nonspecifically targeted when recognised CF pathogens are treated. Furthermore, systemic antibiotics may adversely affect anaerobic bacteria that are present in high abundance in the normal microbiota of the gastrointestinal tract and contribute to disruption of the intestinal microbial community ("dysbiosis") [46]. CF is characterised by both local and systemic inflammation and evidence is accumulating that indicates a relationship exists between the gut microbiota and the health of the airways [47-50]. For example, disruption of gut homeostasis may result in immunoregulatory effects outside the intestinal tract, which may impact respiratory diseases including chronic obstructive pulmonary disease [51]. Therefore, any potential antibiotic treatment used to target anaerobes within the CF lower airways may need to be tailored to minimise adverse effects on the gut microbiota. Development of local therapies that specifically reduce bacterial interactions, limit the host inflammatory response triggered by anaerobes or minimise growth of target

anaerobes in the lungs, e.g. hyperbaric oxygen therapy, may offer an alternative [27**,37**].

Metagenomic sequencing studies generally indicate that in the absence of anaerobes, the health of patients is worse. Studies are required to understand the activities and interactions of microbes in early-stage disease and how the evolution of the bacterial community towards a less diverse end-stage disease (as found in explanted lungs) can be prevented [52]. We have also entered the era of CFTR modulators (e.g. correctors and potentiators), which target CFTR dysfunction, and how these treatments affect development of the airway community has been under short-term surveillance [28,53,54] but longer studies are required. Furthermore, the members and functional characteristics of the airway microbiota are individual to a person. Based on analysis of lung tissue samples from a symptomatic paediatric CF patient, who underwent a right lower lobectomy [55,56], bacterial communities are also known to be spatially heterogeneous in non-end stage disease. Therefore, virulence of anaerobic bacteria may differ both within and between people with CF.

In the future it may be possible to commence a personalised treatment approach that will maintain a normal microbial community, limit pathoadaptation of bacteria residing within the lower airways, prevent dominance by a recognised CF pathogen and subsequent lung damage in the individual. In fact, the Cystic Fibrosis Microbiome-determined Antibiotic Therapy Trial in Exacerbations: Results Stratified (CFMATTERS) trail is the first randomized, controlled trial comparing the use of microbiome-directed antibiotic treatment with standard therapy (tobramycin and ceftazidime) for

people with CF experiencing respiratory infections [57**]. In this clinical trial, which commenced recruitment in 2014, DNA is extracted from sputum and sequenced. Results are reviewed by a Consensus Expert Treatment Panel, who agree on a 3rd antibiotic treatment to add to standard therapy targeting *P. aeruginosa*. This 3rd antibiotic is selected to target additional bacteria detected in the airway microbiome.

Conclusion

Anaerobic bacteria are detected in respiratory samples provided by people with CF of all ages and stages of disease suggesting that they are common and persistent members of the lower airway community. The role of anaerobic bacteria in the CF lower airways might be as members of a normal airway microbiota with disruption of this microbiota and dominance of recognised pathogens responsible for disease progression. However, anaerobic bacteria also produce virulence factors and may contribute to a dysregulated inflammatory response or promote recognised CF pathogens to colonise and infect the lower airways. Additional studies are required to understand if specific anaerobic genera or species are important in health or disease of the CF airways and if this differs between individuals. Research investigating ways to prevent the evolution of the bacterial community towards a less diverse end-stage disease without disrupting the gut microbiota is commended.

(Word count: 2312)

Key points

- Anaerobic bacteria are commonly detected in the lower airways of people with CF but their clinical significance is unknown.
- The relative abundance of anaerobes in the lower airways fluctuates over time.
- A lower abundance of anaerobes has been associated with worse lung function suggesting that anaerobes may form part of a normal airway community.
- Anaerobic bacteria produce virulence factors and may enhance pathogenicity and colonisation by recognised CF pathogens.
- A personalised treatment approach might be required to prevent the development of a microbiota that is characteristic of end-stage lung disease.

Conflicts of interest

S.C.B. has served as a member of an Advisory Board to Vertex and Raptor, received travel support to attend and speak at symposia by Gilead and Novartis and acted as a lead investigator (site) of trials sponsored by Pharmaxis, Vertex and Raptor. M.M.T. is a co-investigator on the CFMATTERS trial and has served as a member of an Advisory Board to Bayer and received travel support to attend and speak at symposia by Novartis.

References

1. Barto TL, Flume PA. Treatment of pulmonary exacerbations in adult cystic fibrosis patients: a review. Hosp Pract (1995) 2010; 38:26-34.

2. Hartl D, Gaggar A, Bruscia E, *et al.* Innate immunity in cystic fibrosis lung disease. J Cyst Fibros 2012; 11:363-382.

Cystic Fibrosis Trust. UK Cystic Fibrosis Registry 2014 Annual Report.
 2015.

4. Doring G, Flume P, Heijerman H, Elborn JS, Consensus Study Group. Treatment of lung infection in patients with cystic fibrosis: current and future strategies. J Cyst Fibros 2012; 11:461-479.

5. UK Cystic Fibrosis Trust Microbiology Laboratory Standards Working Group. Laboratory standards for processing microbiological samples from people with cystic fibrosis. 2010.

6. Harris JK, De Groote MA, Sagel SD, *et al.* Molecular identification of bacteria in bronchoalveolar lavage fluid from children with cystic fibrosis. Proc Natl Acad Sci U S A 2007; 104:20529-20533.

7. Tunney MM, Field TR, Moriarty TF, *et al.* Detection of anaerobic bacteria in high numbers in sputum from patients with cystic fibrosis. Am J Respir Crit Care Med 2008; 177:995-1001.

8. Tunney MM, Klem ER, Fodor AA, *et al.* Use of culture and molecular analysis to determine the effect of antibiotic treatment on microbial community diversity and abundance during exacerbation in patients with cystic fibrosis. Thorax 2011; 66:579-584.

9. Fodor AA, Klem ER, Gilpin DF, *et al*. The adult cystic fibrosis airway microbiota is stable over time and infection type, and highly resilient to antibiotic treatment of exacerbations. PLoS One 2012; 7:e45001.

10. Klepac-Ceraj V, Lemon KP, Martin TR, *et al.* Relationship between cystic fibrosis respiratory tract bacterial communities and age, genotype, antibiotics and *Pseudomonas aeruginosa*. Environ Microbiol 2010; 12:1293-1303.

11. Zhao J, Schloss PD, Kalikin LM, *et al.* Decade-long bacterial community dynamics in cystic fibrosis airways. Proc Natl Acad Sci U S A 2012; 109:5809-5814.

12. Hauser PM, Bernard T, Greub G, *et al.* Microbiota present in cystic fibrosis lungs as revealed by whole genome sequencing. PLoS One 2014; 9:e90934.

13. Worlitzsch D, Tarran R, Ulrich M, *et al.* Effects of reduced mucus oxygen concentration in airway *Pseudomonas* infections of cystic fibrosis patients. J Clin Invest 2002; 109:317-325.

14. Cowley ES, Kopf SH, LaRiviere A, *et al.* Pediatric Cystic Fibrosis Sputum Can Be Chemically Dynamic, Anoxic, and Extremely Reduced Due to Hydrogen Sulfide Formation. MBio 2015; 6:e00767.

*15. Rivas Caldas R, Le Gall F, Revert K, *et al. Pseudomonas aeruginosa* and Periodontal Pathogens in the Oral Cavity and Lungs of Cystic Fibrosis Patients: a Case-Control Study. J Clin Microbiol 2015; 53:1898-1907.

This article deomonstrated that periodontal-associated anaerobic bacteria are found in matched plaque and sputum samples of people with CF.

16. Venkataraman A, Bassis CM, Beck JM, *et al.* Application of a neutral community model to assess structuring of the human lung microbiome. MBio 2015; 6:e02284.

17. Bassis CM, Erb-Downward JR, Dickson RP, *et al.* Analysis of the upper respiratory tract microbiotas as the source of the lung and gastric microbiotas in healthy individuals. MBio 2015; 6:e00037.

18. Whiteson KL, Bailey B, Bergkessel M, *et al.* The upper respiratory tract as a microbial source for pulmonary infections in cystic fibrosis. Parallels from island biogeography. Am J Respir Crit Care Med 2014; 189:1309-1315.

19. Coburn B, Wang PW, Diaz Caballero J, *et al.* Lung microbiota across age and disease stage in cystic fibrosis. Sci Rep 2015; 5:10241.

20. Paganin P, Fiscarelli EV, Tuccio V, *et al*. Changes in cystic fibrosis airway microbial community associated with a severe decline in lung function. PLoS One 2015; 10:e0124348.

21. Flight WG, Smith A, Paisey C, *et al.* Rapid Detection of Emerging Pathogens and Loss of Microbial Diversity Associated with Severe Lung Disease in Cystic Fibrosis. J Clin Microbiol 2015; 53:2022-2029.

22. Stokell JR, Gharaibeh RZ, Hamp TJ, *et al.* Analysis of changes in diversity and abundance of the microbial community in a cystic fibrosis patient over a multiyear period. J Clin Microbiol 2015; 53:237-247.

*23. O'Neill K, Bradley JM, Johnston E, *et al.* Reduced bacterial colony count of anaerobic bacteria is associated with a worsening in lung clearance index and inflammation in cystic fibrosis. PLoS One 2015; 10:e0126980.

This article found that worse lung function and increased airway inflammation were associated with a lower abundance of anaerobic bacteria in the lower airway. Microbiota disruption might result in disease progression.

24. Hajishengallis G, Liang S, Payne MA, *et al.* Low-abundance biofilm species orchestrates inflammatory periodontal disease through the commensal microbiota and complement. Cell Host Microbe 2011; 10:497-506.
25. Zemanick ET, Harris JK, Wagner BD, *et al.* Inflammation and airway microbiota during cystic fibrosis pulmonary exacerbations. PLoS One 2013; 8:e62917.

26. Smith DJ, Badrick AC, Zakrzewski M, *et al.* Pyrosequencing reveals transient cystic fibrosis lung microbiome changes with intravenous antibiotics. Eur Respir J 2014; 44:922-930.

**27. Quinn RA, Whiteson K, Lim YW, *et al.* A Winogradsky-based culture system shows an association between microbial fermentation and cystic fibrosis exacerbation. ISME J 2015; 9:1024-1038.

This study used a novel microbial culture system to measure the physiochemical properties of the microbiota in sputum. The article hypothesised that acidic and gaseous products produced by fermentative anaerobes may drive the development of pulmonary exacerbation.

28. Bernarde C, Keravec M, Mounier J, *et al.* Impact of the CFTRpotentiator ivacaftor on airway microbiota in cystic fibrosis patients carrying a G551D mutation. PLoS One 2015;10:e0124124.

*29. Cuthbertson L, Rogers GB, Walker AW, *et al.* Respiratory microbiota resistance and resilience to pulmonary exacerbation and subsequent antimicrobial intervention. ISME J 2016;10:1081-1091.

This article suggested that *Veillonella* and *Prevotella* species represent potential surrogate markers of disease state due to significant changes in abundance from baseline prior to the start of antibiotic treatment of a pulmonary exacerbation.

30. Skolnik K, Nguyen A, Somayaji R, *et al.* Clinical implications and characterization of Group A *Streptoccoccus* infections in adults with cystic fibrosis. BMC Pulm Med 2015; 15:161.

31. Kramer R, Sauer-Heilborn A, Welte T, *et al.* High individuality of respiratory bacterial communities in a large cohort of adult cystic fibrosis patients under continuous antibiotic treatment. PLoS One 2015; 10:e0117436.

32. Marvig RL, Sommer LM, Molin S, Johansen HK. Convergent evolution and adaptation of *Pseudomonas aeruginosa* within patients with cystic fibrosis. Nat Genet 2015; 47:57-64.

33. Sherrard LJ, Graham KA, McGrath SJ, *et al.* Antibiotic resistance in *Prevotella* species isolated from patients with cystic fibrosis. J Antimicrob Chemother 2013; 68:2369-2374.

34. Thornton CS, Grinwis ME, Sibley CD, *et al*. Antibiotic susceptibility and molecular mechanisms of macrolide resistance in streptococci isolated from adult cystic fibrosis patients. J Med Microbiol 2015; 64:1375-1386.

35. Tai AS, Bell SC, Kidd TJ, *et al.* Genotypic Diversity within a Single *Pseudomonas aeruginosa* Strain Commonly Shared by Australian Patients with Cystic Fibrosis PLoS One 2015; 10:e0144022.

36. Tribble GD, Kerr JE, Wang BY. Genetic diversity in the oral pathogen *Porphyromonas gingivalis*: molecular mechanisms and biological consequences. Future Microbiol 2013; 8:607-620.

**37. Mirkovic B, Murray MA, Lavelle GM, *et al*. The Role of Short-Chain Fatty Acids, Produced by Anaerobic Bacteria, in the Cystic Fibrosis Airway. Am J Respir Crit Care Med 2015; 192:1314-1324.

This article demonstrated that anaerobic bacteria may contribute to the dysregulated inflammatory response in the CF airways by the secretion of various short chain fatty acids.

**38. Ghorbani P, Santhakumar P, Hu Q, *et al.* Short-chain fatty acids affect cystic fibrosis airway inflammation and bacterial growth. Eur Respir J 2015;
46:1033-1045.

This article highlighted that anaerobic bacteria produce short-chain fatty acids through fermentation. Short-chain fatty acids were detected in sputum of people with CF, which influenced host inflammatory responses.

*39. Pustelny C, Komor U, Pawar V, *et al*. Contribution of *Veillonella parvula* to *Pseudomonas aeruginosa*-mediated pathogenicity in a murine tumor model system. Infect Immun 2015; 83:417-429.

This study used an *in vivo* model to show that co-infection of a recognised CF pathogen with an anaerobe resulted in greater clinical deterioration compared to infection with the pathogen alone.

40. Benedyk M, Byrne DP, Glowczyk I, *et al.* Pyocycanin, a contributory factor in haem acquisition and virulence enhancement of *Porphyromonas gingivalis* in the lung. PLoS One 2015; 10:e0118319.

*41. Whiley RA, Fleming EV, Makhija R, Waite RD. Environment and colonisation sequence are key parameters driving cooperation and competition between *Pseudomonas aeruginosa* cystic fibrosis strains and oral commensal streptococci. PLoS One 2015; 10:e0115513.

The article reported that the pathogenic and colonisation ability of *Pseudomonas aeruginosa* was modulated by the presence of oral commensal streptococci.

42. Price KE, Naimie AA, Griffin EF, *et al.* Tobramycin-Treated *Pseudomonas aeruginosa* PA14 Enhances *Streptococcus constellatus* 7155 Biofilm Formation in a Cystic Fibrosis Model System. J Bacteriol 2015; 198:237-247.

43. Yamasaki K, Mukae H, Kawanami T, *et al.* Possible role of anaerobes in the pathogenesis of nontuberculous mycobacterial infection. Respirology 2015; 20:758-765.

44. Sherrard LJ, Tunney MM, Elborn JS. Antimicrobial resistance in the respiratory microbiota of people with cystic fibrosis. Lancet 2014; 384:703-713.

*45. Sherrard LJ, McGrath SJ, McIlreavey L, *et al.* Production of extendedspectrum beta-lactamases and the potential indirect pathogenic role of *Prevotella* isolates from the cystic fibrosis respiratory microbiota. Int J Antimicrob Agents 2016; 47:140-145.

This article was the first to demonstrate that a *Prevotella* species could protect *Pseudomonas aeruginosa* from antibiotic treatment *in vitro*.

46. Ooi CY, Durie PR. Cystic fibrosis from the gastroenterologist's perspective. Nat Rev Gastroenterol Hepatol 2016; 13:175-185.

47. Wojewodka G, De Sanctis JB, Bernier J, *et al.* Candidate markers associated with the probability of future pulmonary exacerbations in cystic fibrosis patients. PLoS One 2014; 9:e88567.

48. Li L, Somerset S. The clinical significance of the gut microbiota in cystic fibrosis and the potential for dietary therapies. Clin Nutr 2014; 33:571-580.

49. Hoen AG, Li J, Moulton LA, O'Toole GA, *et al.* Associations between Gut Microbial Colonization in Early Life and Respiratory Outcomes in Cystic Fibrosis. J Pediatr 2015; 167:138-147.

50. Bazett M, Bergeron ME, Haston CK. Streptomycin treatment alters the intestinal microbiome, pulmonary T cell profile and airway hyperresponsiveness in a cystic fibrosis mouse model. Sci Rep 2016; 6:19189.

51. Keely S, Talley NJ, Hansbro PM. Pulmonary-intestinal cross-talk in mucosal inflammatory disease. Mucosal Immunol 2012; 5:7-18.

52. Rudkjobing VB, Thomsen TR, Alhede M, *et al.* The microorganisms in chronically infected end-stage and non-end-stage cystic fibrosis patients. FEMS Immunol Med Microbiol 2012; 65:236-244.

53. Rowe SM, Heltshe SL, Gonska T, *et al.* Clinical mechanism of the cystic fibrosis transmembrane conductance regulator potentiator ivacaftor in G551D-mediated cystic fibrosis. Am J Respir Crit Care Med 2014; 190:175-184.

54. Heltshe SL, Mayer-Hamblett N, Burns JL, *et al. Pseudomonas aeruginosa* in cystic fibrosis patients with G551D-CFTR treated with ivacaftor. Clin Infect Dis 2015; 60:703-712.

55. Boutin S, Graeber SY, Weitnauer M, *et al.* Comparison of microbiomes from different niches of upper and lower airways in children and adolescents with cystic fibrosis. PLoS One 2015; 10:e0116029.

56. Brown PS, Pope CE, Marsh RL, *et al.* Directly sampling the lung of a young child with cystic fibrosis reveals diverse microbiota. Ann Am Thorac Soc 2014; 11:1049-1055.

**57. http://www.cfmatters.eu/the-project/

This website provides details of the CFMATTERS study that is underway. This is the first randomised, controlled trial, which will use 'real-time' microbiome data to inform antibiotic treatment of a chronic infection.

Figure titles and legends

FIGURE 1. Prevalence of recognised respiratory pathogens among paediatrics and adults with cystic fibrosis (UK) in 2014 (n=9432) [3].

Abbreviation: MRSA, methicillin-resistant Staphylococcus aureus.

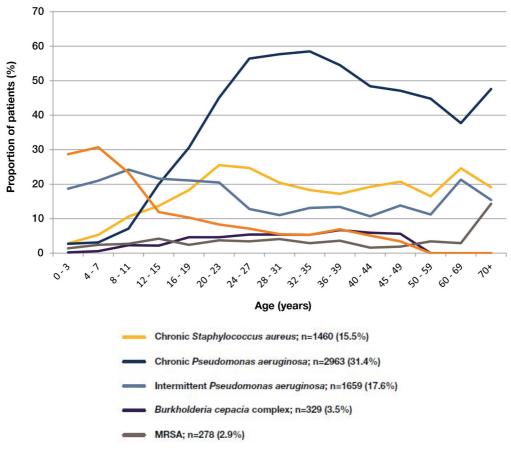
(Reproduced with permission from the UK Cystic Fibrosis Trust).

FIGURE 2. A potential dynamic relationship between the relative abundance of anaerobic bacteria, recognised pathogens, lung function and inflammation in the cystic fibrosis airways over time.

(ORIGINAL)

FIGURE 3. Potential pathogenic mechanisms of anaerobic bacteria detected in the cystic fibrosis airways. Anaerobic bacteria may [A] increase the pathogenicity of recognised cystic fibrosis pathogens [B] produce short-chain fatty acids that exaggerate the host inflammatory response [C] shield recognised cystic fibrosis pathogens from treatment with antibiotics. (ORIGINAL)





Haemophilus Influenza; n=1310 (13.9%)

FIGURE 2

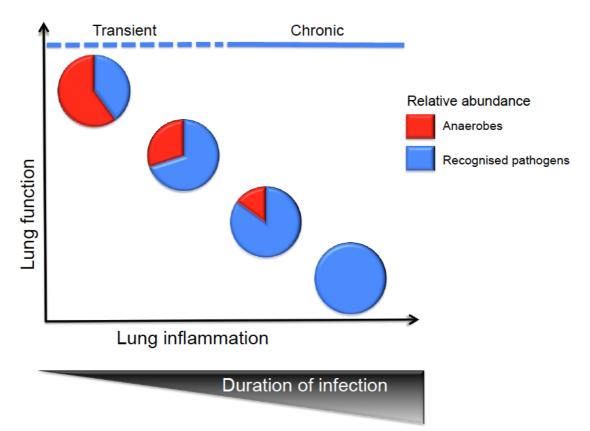


FIGURE 3

