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## **Development of release methods for critically endangered freshwater pearl mussels (*Margaritifera margaritifera*)**

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1 Original Research

2 **Development of release methods for captive-bred freshwater pearl**  
3 **mussels (*Margaritifera margaritifera*)**

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18

19 **Running title**

20 *Ex situ* conservation of endangered *M. margaritifera*

21

22 **Keywords**

23 Conservation, endangered species, freshwater pearl mussel, juvenile culture, mussel silo,  
24 release, survival rate

## 1 **Abstract**

2 Biodiversity loss is a global problem with freshwater bivalves considered amongst the most  
3 endangered biota. The freshwater pearl mussel, *Margaritifera margaritifera*, is declining  
4 throughout its range owing to habitat degradation and overexploitation. In most of its range,  
5 populations are regarded as reproductively non-functional which has led to the development  
6 of captive breeding programmes. A novel method of releasing *M. margaritifera* was trialled,  
7 with captive-bred juveniles being released into the rivers caged in ‘mussels silos’ (protective  
8 concrete domes with ventilation creating upwelling to ensure water through flow). We  
9 released 240 juvenile mussels and survival and growth rates were monitored for 18 months  
10 post-release for three size classes: A (13.01-20.00mm); B (10.01-13.00mm); and C (4.01-  
11 10.00mm). We explicitly tested two experimental treatments; one where sediment was added  
12 to each silo (allowing mussels to orientate and burrow) and one without sediment. Survival  
13 by the end of the experiment at month 18 was significantly higher for the largest size class at  
14 97% (though growth was lowest in this cohort), and lowest for the smallest size class at 61%  
15 (though growth was highest in this cohort). Survival and growth were unaffected by the  
16 experimental treatment suggesting that adding sediment offered no advantage. Growth was  
17 positively correlated with both water temperature and the particle size of suspended solids  
18 (both of which were collinear, peaking in summer). There are a large number of *ex situ*  
19 breeding programmes for freshwater pearl mussels throughout Europe and our finding  
20 suggest that the use of ‘mussel silos’ could be a useful tool to protecting juvenile mussels  
21 allowing them to be released at a relatively early stage of development, minimising the risk of  
22 domestication.

## 1 **1.1 Introduction**

2 Biodiversity loss is a global problem occurring across all ecosystems and taxa (Brooks et al.,  
3 2002, 2006; Hooper et al., 2012). Freshwater ecosystems have a higher extinction rate than  
4 terrestrial systems (Saunders et al., 2002) and freshwater mussels are considered among the  
5 most globally endangered biota (Geist and Auerswald, 2007; Geist and Kuehn, 2005; Strayer  
6 et al., 2004). Ricciardi and Rasmussen (1999) estimated that at least 127 imperilled mussel  
7 species would disappear within the next 100 years, suggesting the extinction rate could be as  
8 high as 6.4% per decade. Numerous *ex situ* conservation programmes for both aquatic and  
9 terrestrial species have been developed in an attempt to help reduce predicted biodiversity  
10 losses (Bolland et al., 2010).

11 Across North America and Europe facilities have been developed to cultivate endangered  
12 or threatened freshwater mussels using various methods with the aim of releasing them if  
13 suitable habitat remains or is restored (Jones et al., 2006). In North America an intensive  
14 cultivation system is often used. Juveniles are propagated in laboratory conditions and kept in  
15 baskets or raceways in either flow-through or re-circulating systems with detritus added as a  
16 food resource (Gum et al., 2011; Jones et al., 2005; Thomas et al., 2010). A number of  
17 hatcheries in Europe use a similar approach (Buddensiek, 1995; Schmidt and Vandr , 2010),  
18 although a small number use less intensive approaches that involve bringing mussels and fish  
19 host into closer proximity under semi-natural environmental conditions (Gum et al., 2011;  
20 Preston et al., 2007). A number of intermediate rearing cages have been developed and  
21 trialled to release mussels at an early stage directly into the release site to avoid the risk of  
22 domestication during prolonged periods in captivity, such as Buddensiek sheet cages  
23 (Buddensiek, 1995), gauze bags (Schmidt and Vandr , 2010) and mussel silos (Barnhart et  
24 al., 2007), Table 1.

1 The freshwater pearl mussel, *Margaritifera margaritifera*, is a long-lived, globally  
2 endangered bivalve with populations having declined by up to 90% throughout its Holarctic  
3 range over the course of the 20<sup>th</sup> century (Bauer, 1988; Cosgrove et al., 2000; Bolland et al.,  
4 2010; Reid et al., 2013). These declines have primarily been linked to pollution driven by  
5 nutrient enrichment and habitat degradation (Beasley and Roberts, 1999; Hastie et al., 2003;  
6 Reis, 2003; Österling et al., 2010; Österling and Högberg, 2014), the lack of suitable host fish  
7 (Geist et al., 2006; Arvidsson et al., 2012; Österling and Larsen, 2013), and historically pearl  
8 fishing on a local scale (Cosgrove et al., 2000). The freshwater pearl mussel has a complex  
9 lifecycle, during which the parasitic glochidia (larvae released from the gills of the female  
10 mussel) spend up to nine months on the gills of a host fish, normally salmon, *Salmo salar*, or  
11 trout, *Salmo trutta* (Geist et al., 2006). *M. margaritifera* is regarded as an indicator species  
12 because it is sensitive to environmental change and is regarded as a keystone species due to  
13 its filtering because of its filtering capacity (Geist, 2010).

14 *M. margaritifera* is generally found in clean, moderate-fast flowing waters and is  
15 associated with a stable cobble-boulder substratum including sand for burrowing (Hastie *et*  
16 *al.*, 2000, 2003; Wilson *et al.*, 2011). Juvenile mussels spend the first four to five years  
17 completely burrowed in the riverbed and require high rates of oxygen exchange between free  
18 moving and interstitial water (Buddensiek, 1995). If conditions are muddy or silty the  
19 juveniles will suffocate as a result of clogging of the interstitial spaces inhibiting the  
20 exchange of oxygen (Hastie et al., 2000; Geist and Auerswald 2007).

21 In Northern Ireland, the freshwater pearl mussel, which anecdotal evidence suggests was  
22 once found abundantly throughout the country (Beasley et al., 1998), is recently extinct from  
23 ten rivers and its range is now restricted to short stretches of just six rivers (Fig. 1).  
24 Remaining remnant populations can be separated into three genetically distinct  
25 metapopulations which should be managed separately in any proposed captive breeding

1 programmes (Wilson *et al.* 2012). Recent surveys suggest there approximately 22,000  
2 mussels remain throughout their range in Northern Ireland but that all populations are  
3 regarded as reproductively “non-functional” owing to a lack of recruitment (Reid *et al.*,  
4 2013). Total species extirpation in Northern Ireland has been predicted by the year 2098  
5 (Wilson and Roberts, 2011). Surveys have revealed there are fewer than 1000 wild mussels  
6 in the Ballinderry River (Reid *et al.*, 2013); these declines are linked primarily to habitat  
7 degradation and declines in host fish (Horton *et al.*, 2015).

8       Consequently, an *ex situ* captive breeding programme was established in 1998 with 100  
9 adults taken as broodstock for a custom-built facility on the Ballinderry River (Fig. 1; Preston  
10 *et al.*, 2007). This system uses a semi-natural approach to cultivation, allowing natural  
11 fertilisation of mussels, with water from mussel tanks draining into tanks containing suitable  
12 host fish for infection. Infected fish are then transferred a vivarium (a tank set up to mimic  
13 the natural river with substratum and controlled flow) where glochidia to fall off (excyst) the  
14 gills of the fish and burrow into river substratum about nine months later (Preston *et al.*,  
15 2007). Previously, 350 hatchery-reared juvenile mussels (*ca.* 8-10 years old) were released  
16 directly into the Ballinderry River at three sites in February and August 2009 with subsequent  
17 survival monitored by passive integrated transponders (PIT) tags (Wilson 2010). Recovered  
18 shells show mortality rates during the first 18 months post-release ranging from 4.3 - 14.3%  
19 with the estimated maximum mortality calculated from mussels which were not recovered  
20 again ranging from 33.6-36.6% suggesting hatchery-reared mussels could survive at least  
21 initial release into the river.

22       The semi-natural cultivation system used in Northern Ireland means that when individuals  
23 are large enough to be collected from the vivarium, they are too large for Buddensiek cages  
24 (Buddensiek, 1995). This study aimed to test the use of mussel silos (Fig. 2 and 3), designed  
25 by Barnhart *et al.* (2007), as an alternative release method facilitating the release of larger

1 juvenile mussels than are used by other programmes. We hypothesised that releasing larger  
2 individuals will result in greater survival and thus success for the captive breeding and release  
3 programme. Mussel silos use the Bernoulli effect (Barnhart et al., 2007), to draw a  
4 continuous supply of water through a hole in the centre of a concrete dome (Barnhart *pers.*  
5 *comm.*). The main objectives of this study were to test: 1) The efficacy of mussel silos at  
6 improving juvenile *M. margaritifera* survival; 2) the minimum size at which juvenile mussels  
7 can be released with comparable (or better) survival to other methods; 3) the variation in  
8 survival and growth rate among different size classes, treatments and release sites.

9

## 1 **1.2 Materials and Methods**

### 2 **1.2.1 Site selection**

3 Extensive surveys were carried out along the Ballinderry River catchment to identify two  
4 suitable sites to trial the release of juvenile *M. margaritifera* into mussel silos. Site 1 was a  
5 tributary of the main Ballinderry River channel selected because of high water quality and the  
6 presence of suitable habitat, such as cobbles with gravel for burrowing and bankside  
7 vegetation (following Wilson *et al.*, 2011; Horton *et al.*, 2015). Site 2 was on the main  
8 Ballinderry River channel with suitable habitat (following Wilson *et al.* 2011) and a nearby  
9 extant remnant, non-functional, adult mussel population.

10

### 11 **1.2.2 Selection and tagging of mussels**

12 Juvenile mussels were collected from the *ex situ* culturing vivarium and divided into three  
13 size classes, with 80 individuals in each; Class A = 13.01 - 20.00 mm (mean 15 mm), Class B  
14 = 10.01 - 13.00 mm (mean 11 mm) and Class C = 4.00 - 10.00 mm (mean 8 mm). Prior to  
15 release, each mussel was quantified and tagged with a bee tag (EH Thorne (Beehives) Ltd.,  
16 Lincolnshire, UK) for identification purposes. Each mussel was swabbed with alcohol, lightly  
17 sanded and had alcohol applied again until dry. The bee tag was then attached using Loctite  
18 Precision Super Glue.

19

### 20 **1.2.3 Experimental design**

21 Four individuals from each size class were included in each silo (12 individuals per silo). At  
22 each of the two sites 10 silos were deployed, five of which had the sediment treatment and  
23 five of which had the no sediment treatment. Mussels in the sediment treatment had a small  
24 amount of river substrate gravel included within the chamber (which was filled up to 2.5 cm  
25 from the top of the chamber) where mussels were held. The no sediment treatment had



1 nothing included with the mussels which is what had been trialled previously (Barnhart et al.,  
2 2007). Sediment was included as a treatment to test the hypothesis that mussels exposed to  
3 sediment should have a higher survival and growth than mussels not exposed to sediment as  
4 they would be able to orientate and anchor themselves within the sediment rather than being  
5 vulnerable to the water flow.

6

#### 7 **1.2.4 Maintenance and monitoring**

8 Site visits were carried out once a week when conditions permitted to ensure there were no  
9 blockages interrupting water flow through the mussel silo chamber. Mussel silo chambers  
10 were opened once a month and survival (0/1) was recorded for each individual over an 18  
11 month period (from September 2013 to March 2015). During the course of the experiment,  
12 one mussel chamber, which was part of the sediment treatment, was lost at Site 2 either by  
13 being washed out or stolen, and was excluded from analysis.

14 Temperature and siltation data were collected each month to provide descriptions of  
15 background conditions and help interpret results. An eleventh silo containing gravel without  
16 mussels was included at each site as a sediment trap to quantify the settlement of suspended  
17 solids; this was emptied and refilled each month. Sediment analysis was carried out with the  
18 Department of the Environment Marine Division. HOBO Pendant® Temp/Light, 64K data  
19 loggers (Tempcon Instrumentation Ltd., West Sussex, UK) were deployed at each site,  
20 logging temperature every two hours.

21 Mussel growth was quantified over three consecutive six monthly periods (0-6, 6-12 and  
22 12-18 months) after release. Instantaneous growth rate ( $r$ ) was calculated as:

$$23 \quad r = \ln(Sl_t) - \ln(Sl_{t-1})$$

24

1 where  $Sl$  was shell length in millimetres (mm) at time period  $t$  (current measurements) or  $t-1$   
2 (previous measurement) expressed as a natural logarithm.

3

#### 4 **1.2.5 Data analysis**

5 Survival and growth of juvenile *M. margaritifera* post-release were examined using General  
6 Linear Mixed Models (GLMMs). Survival (0/1) was fitted using a binomial logistic  
7 distribution whilst growth was fitted using a gamma distribution (i.e. data were highly left  
8 skewed). Individual mussel identity was included as a nested Random Factor within Site i.e.  
9 Site (Individual\_ID) fitted with an autoregressive error structure (AR1) to account for  
10 multiple measurements per individual per site. Site (1/2), Size Class (A, B and C), and  
11 Treatment (Sediment/No Sediment) were fitted as fixed factors. For the survival model, the  
12 effect of time was fitted as eighteen Months (0-18 inclusive) whilst for the growth model;  
13 time was fitted as three Periods (0-6, 6-12 and 12-18 months). Two-way interactions were  
14 initially fitted but subsequently dropped as they were not significant. *Post-hoc* pairwise  
15 differences in survival between size classes by the end of the experiment at month 18 were  
16 tested by *t*-tests. The relationship between growth rate and water temperature and sediment  
17 particle size was examined using Pearson's correlation whilst differences in monthly water  
18 temperature and sediment particle size was tested between site using paired *t*-tests. Graphs  
19 show combined data from sites 1 and 2. Results examine 228 mussels rather than 240 as one  
20 cell was lost before mortality and growth could be monitored. All statistical analysis was  
21 performed using IBM SPSS v21 and all plots were drawn in SigmaPlot v12.

### 1 1.3 Results

2 Post-release survival varied significantly ( $F_{df=21,4082} = 8.950, p < 0.001$ ) among size classes, site  
3 and month but not between experimental sediment treatments (Table 2a). Survival by the end  
4 of the experiment at month 18 differed significantly between all size classes (*post-hoc*  
5 pairwise contrasts  $p < 0.020$ ) where the largest size class A (~15 mm) had the greatest survival  
6 (97%), followed by the intermediate size class B (~11 mm; 86% survival) and the lowest  
7 survival was for the smallest size class C (~8 mm; 61% survival). Survival rates in the  
8 smallest size class C stabilised after 9-10 months (Fig. 4). Survival (mean percentage  $\pm$  1SD)  
9 was lower at Site 1 ( $76 \pm 43\%$  survival) compared to Site 2 ( $87 \pm 34\%$  survival). Survival in  
10 silos without the sediment treatment was  $82 \pm 39\%$  compared to  $81 \pm 40\%$  for those with the  
11 sediment treatment 18 months post release.

12 Growth rates varied significantly ( $F_{df=6,455} = 82.989, p < 0.001$ ) among size classes, site and  
13 time periods (three six monthly periods), but not between experimental sediment treatments  
14 (Table 2b). The instantaneous growth rate (mean  $\pm$  1SD) was similar between the largest size  
15 class A ( $0.014 \pm 0.009$ ) and the intermediate size class B ( $0.015 \pm 0.010$ ) but substantially  
16 higher (+38%) in the smallest size class C ( $0.020 \pm 0.032$ ) 18 months post release. Growth  
17 rate was 53% lower at Site 1 ( $0.023 \pm 0.027$ ) than at Site 2 ( $0.050 \pm 0.055$ ). Growth was 4.3  
18 fold greater during the summer period (6-12 months post release) than the winter period (0-6  
19 and 12-18 months post release), but did not vary between sediment treatments (Fig 5) despite  
20 mussels exposed to sediment having a 9% higher growth rate ( $0.038 \pm 0.050$ ) than those in  
21 the no sediment treatment ( $0.035 \pm 0.041$ ). Growth was positively correlated with both water  
22 temperature ( $r_p = 0.733, p < 0.001$ ) and sediment particle size ( $r_p = 0.217, p < 0.001$ ). Particle  
23 size was weakly correlated with water temperature ( $r_p = 0.100, p = 0.009$ ) with sediment  
24 deposition being greatest during summer low water flow (Kyle *pers. obs.*). Particle size was  
25 significantly smaller (paired  $t_{df=12} = 3.269, p = 0.007$ ) at Site 1 ( $592.1 \pm 114.4\mu\text{m}$ ) than Site 2

- 1 (713.6 ± 154.1µm) whilst water temperature was lower but not significantly so (paired  $t_{df=17} =$
- 2 0.318,  $p=0.754$ ) at Site 1 (8.9 ± 3.0°C) than Site 2 (9.1 ± 3.4°C).

## 1 **1.4 Discussion**

2 Larger mussels were shown to have the greatest survival post-release (size class A; 13.01-  
3 20.00 mm) and were virtually all alive after 18 months but had the lowest growth rate. Small  
4 mussels had the highest mortality of almost 40% (size class C; 4.01 – 10.00 mm) after 18  
5 months post-release but had the highest growth rate. Post-parasitic *M. margaritifera* spend  
6 the first four to five years of their life completely submerged in the sediment pedal feeding  
7 before making the transition to filter feeding (Bauer and Vogel, 1987; Geist and Auerswald,  
8 2007). Mussel size varies across its range (Miguel et al., 2004; Ziuganov, 2004) and age  
9 determination of the shell is most accurate when the mussel is sacrificed (Helama and  
10 Valovirta, 2008), therefore, very little is understood about when exactly the pedal-filter  
11 feeding transition takes place and whether it is a gradual change or sudden. It is possible that  
12 a least a proportion of the mussels in size class C were still in the pedal feeding stage and  
13 food availability was a limiting resource contributing to their higher mortality rates. They  
14 would also be more vulnerable to being physically covered by silt.

15 This study had a mean mussel survival of 81%, which was relatively high compared to  
16 similar release studies (Table 1). A previous study examining *Villosa iris* using mussel silos  
17 designed by Barnhart *et al.*, (2007) showed a similarly high survival rate ranging from 81-  
18 88% (Johnson et al., 2014). Studies using various other cage release methods of *M.*  
19 *margaritifera* suggest a large degree of variation in survival from 0.21 - 82 % (Hastie and  
20 Young, 2003; Johnson et al., 2014; Schmidt and Vandr , 2010; Wilson, 2010). Buddensiek  
21 cages are widely used as an intermediate release method for juvenile *M. margaritifera*  
22 (Schmidt and Vandr , 2010; Eybe et al., 2013). However, mussels released in Buddensiek  
23 cages have been shown to have highly variable survival rates. Buddensiek (1995) released  
24 juveniles in Buddensiek cages ranging from <500 - >900  $\mu\text{m}$  and found 100% of individuals  
25 less than <700  $\mu\text{m}$  died. Wilson (2010) carried out direct releases into the sediment of

1 mussels, which were *ca.* 10 years old and had a high survival rate, suggesting size (age) at  
2 release is a strong indicator of survival. Based on these findings it would be recommended  
3 that mussels in size class C (4.00 - 10.00 mm) should not be released using mussel silos but  
4 should be maintained within the hatchery facility and released when they attain a minimum of  
5 size class B (10.01 - 13.00 mm).

6 Whilst there was no significant difference in water temperature between sites, highest  
7 survival and growth were coincident with highest water temperature (i.e. at Site 2). Growth  
8 was positively correlated with water temperature and it is well known that many important *M.*  
9 *margaritifera* life stages are achieved only when a certain threshold of degree days have been  
10 experienced (Scheder et al., 2011), highlighting the importance of site selection before  
11 release (Bolland et al., 2010). Sediment deposition was greatest during low summer flow  
12 when temperatures were highest. Thus, particle size was correlated with water temperature  
13 but this probably reflects total levels of sediment deposition rather than a skew in sediment  
14 particle size distribution with temperature. Site 1 had a higher level of sediment deposition  
15 than Site 2 which could have interrupted water flow through mussel silos, therefore, limiting  
16 food causing lower growth and higher mortality at Site 1.

17 Experimental sediment treatment had no effect on survival or growth rates of juvenile *M.*  
18 *margaritifera* in mussel silos. Previous investigations by Barnhart *et al.* (2007) didn't include  
19 sediment, however in this experiment sediment was added to allow mussels to orientate  
20 themselves within the cage system.

21 The greater survival of larger size classes suggest that there are benefits to rearing juvenile  
22 mussels to larger sizes before being released with silos, with little effect of domestication.  
23 Mussel silos provide a high survival rate in release studies (Barnhart et al., 2007; Johnson et  
24 al., 2014), and can be securely fitted within a river to ensure they are not washed away during  
25 flood events. Although sediment was included to allow mussels to orientate themselves, it

1 was not found to be of benefit, with no sediment included the silos require relatively little  
2 husbandry whereas Buddensiek cages need regular cleaning (Scheder et al., 2014). Recent  
3 surveys have shown that the *M. margaritifera* population in the Upper Ballinderry River  
4 catchment is < 1,000 individuals and is regarded as reproductively non-functional (Reid et  
5 al., 2013), with extinction estimated to be in the year 2098 (Wilson and Roberts, 2011). Thus,  
6 the successful release of juvenile mussels via mussel silos coupled with catchment restoration  
7 work nearing completion (Horton et al., 2015) should improve the chances of recreating a  
8 reproductively functional population with a varied age structure. This work has included  
9 bank revetment works, erecting stock proof fencing along the length of the river and closing  
10 open cattle drinkers (Horton et al., 2015). Findings reported in the present paper are widely  
11 applicable to other small populations of *M. margaritifera* which are reproductively non-  
12 functional (i.e. not recruiting). A number of *ex situ* captive breeding programmes are  
13 producing large numbers of juveniles (Preston *et al.*, 2007; McIvor and Aldridge, 2008, and  
14 references therein; Scheder and Gumpinger, 2008). Such mussels could be grown in silos, as  
15 described above, to a size when they can be released into restored natal rivers. Future work  
16 should therefore establish the size at which mussels can be transferred from silos to natural  
17 sediments show survival levels comparable to those reported by Wilson (2010) for mussels  
18 which have recently reached sexual maturity. An advantage of this approach is the mussels in  
19 the silos also serve as biological indicators that test whether water quality is suitable for  
20 mussel release.

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1 **References**

- 2 Arvidsson BL, Karlsson J, Österling ME. 2012. Recruitment of the threatened mussel  
3 *Margaritifera margaritifera* in relation to mussel population size, mussel density and  
4 host density. *Aquatic Conservation: Marine and Freshwater Ecosystems* **22**: 526–532.
- 5 Barnhart MC, Fobian TB, Whites DW, Ingersoll CG. 2007. Mussel silos: Bernoulli flow  
6 devices for caging juvenile mussels in rivers in: Fifth Biennial Symposium of the  
7 Freshwater Mollusk Conservation Society, Little Roc, Ark.
- 8 Bauer G. 1988. Threats to the freshwater pearl mussel *Margaritifera margaritifera* L. in  
9 Central Europe. *Biological Conservation*, **45**: 239–253.
- 10 Bauer G, Vogel C. 1987. The parasitic stage of the freshwater pearl mussel (*Margaritifera*  
11 *margaritifera* L.) I. Host response to glochidiosis. *Archiv Für Hydrobiologie* **76**: 393-  
12 402.
- 13 Beasley C, Roberts D. 1999. Towards a strategy for the conservation of the freshwater pearl  
14 mussel *Margaritifera margaritifera* in County Donegal, Ireland. *Biological*  
15 *Conservation* **89**: 275–284.
- 16 Beasley CR, Roberts D, Mackie TG. 1998. Does the freshwater pearl mussel, *Margaritifera*  
17 *margaritifera* L., face extinction in Northern Ireland? *Aquatic Conservation: Marine and*  
18 *Freshwater Ecosystems* **8**: 265–272.
- 19 Bolland JD, Bracken LJ, Martin R, Lucas MC. 2010. A protocol for stocking hatchery reared  
20 freshwater pearl mussel *Margaritifera margaritifera*. *Aquatic Conservation: Marine and*  
21 *Freshwater Ecosystems* **20** 695–704.

- 1 Brooks TM, Mittermeier RA, Mittermeier CG, da Fonseca GAB, Rylands AB, Konstant WR,  
2 Flick P, Pilgrim J, Oldfield S, Magin G, Hilton-Taylor C. 2002. Habitat Loss and  
3 Extinction in the Hotspots of Biodiversity. *Conservation Biology* **16**: 909–923.
- 4 Brooks TM, Mittermeier RA, da Fonseca GAB, Gerlach J, Hoffmann M, Lamoreux JF,  
5 Mittermeier CG, Pilgrim JD, Rodrigues ASL. 2006. Global biodiversity conservation  
6 priorities. *Science* **313**: 58–61.
- 7 Buddensiek V. 1995. The culture of juvenile freshwater pearl mussels *Margaritifera*  
8 *margaritifera* L. in cages: A contribution to conservation programmes and the  
9 knowledge of habitat requirements. *Biological Conservation* **74**: 33–40.
- 10 Cosgrove PJ, Young MR, Hastie LC, Gaywood M, Boon PJ. (2000) The status of the  
11 freshwater pearl mussel *Margaritifera margaritifera* Linn. in Scotland. *Aquatic*  
12 *Conservation: Marine and Freshwater Ecosystems* **10**: 197–208.
- 13 Eybe T, Thielen F, Bohn T, Sures B. 2013. The first millimetre - rearing juvenile freshwater  
14 pearl mussels (*Margaritifera margaritifera* L.) in plastic boxes. *Aquatic Conservation:*  
15 *Marine and Freshwater Ecosystems* **23**: 964–975.
- 16 Geist J. 2010. Strategies for the conservation of endangered freshwater pearl mussels  
17 (*Margaritifera margaritifera* L.): a synthesis of Conservation Genetics and Ecology.  
18 *Hydrobiologia* **644**: 69–88.
- 19 Geist J, Auerswald K. 2007. Physicochemical stream bed characteristics and recruitment of  
20 the freshwater pearl mussel (*Margaritifera margaritifera*). *Freshwater Biology* **52**:  
21 2299–2316.

- 1 Geist J, Kuehn R. 2005. Genetic diversity and differentiation of central European freshwater  
2 pearl mussel (*Margaritifera margaritifera* L.) populations: implications for conservation  
3 and management. *Molecular Ecology* **14**: 425–439.
- 4 Geist J, Porkka M, Kuehn R. 2006. The status of host fish populations and fish species  
5 richness in European freshwater pearl mussel (*Margaritifera margaritifera*) streams.  
6 *Aquatic Conservation: Marine and Freshwater Ecosystem* **16**: 251–266.
- 7 Gum B, Lange M, Geist J. 2011. A critical reflection on the success of rearing and culturing  
8 juvenile freshwater mussels with a focus on the endangered freshwater pearl mussel  
9 (*Margaritifera margaritifera* L.). *Aquatic Conservation: Marine and Freshwater*  
10 *Ecosystems* **21**: 743–751.
- 11 Hastie LC, Boon PJ, Young MR 2000. Physical microhabitat requirements of freshwater  
12 pearl mussels, *Margaritifera margaritifera* (L.). *Hydrobiologia* **429**: 59–71.
- 13 Hastie LC, Cooksley SL, Scougall F, Young MR, Boon PJ, Gaywood MJ. 2003.  
14 Characterization of freshwater pearl mussel (*Margaritifera margaritifera*) riverine  
15 habitat using River Habitat Survey data. *Aquatic Conservation: Marine and Freshwater*  
16 *Ecosystems* **13**: 213–224.
- 17 Hastie LC, Young MR. 2003. Conservation of the Freshwater Pearl Mussel I: Captive  
18 Breeding Techniques. Conserving Natura 2000 Rivers Conservation Techniques, Series  
19 No. 2. *English Nature, Peterborough*.
- 20 Helama S, Valovirta I. 2008. The oldest recorded animal in Finland: Ontogenetic age and  
21 growth in *Margaritifera margaritifera* (L. 1758) based on internal shell increments.  
22 *Memoranda Societas pro Fauna et Flora Fennica* **84**: 20–30.

- 1 Hooper DU, Adair EC, Cardinale BJ, Byrnes JEK, Hungate BA, Matulich KL, Gonzalez A,  
2 Duffy JE, Gamfeldt L, O'Connor MI. 2012. A global synthesis reveals biodiversity loss  
3 as a major driver of ecosystem change. *Nature* **486**: 105–108.
- 4 Horton M, Keys A, Kirkwood L, Mitchell F, Kyle R, Roberts D. 2015. Sustainable catchment  
5 restoration for reintroduction of captive bred freshwater pearl mussels *Margaritifera*  
6 *margaritifera*. *Limnologica – Ecology and Management of Inland Waters* **50**: 21–28.
- 7 Johnson GC, Krstolic JL, Ostby BJK. 2014. Influences of Water and Sediment Quality and  
8 Hydrologic Processes on Mussels in the Clinch River. *JAWRA Journal of the American*  
9 *Water Resources Association* **50**: 878–897.
- 10 Jones JW, Hallerman EM, Neves RJ. 2006. Genetic management guidelines for captive  
11 propagation of freshwater mussels (Unionoidea). *Journal of Shellfish Research* **25**: 527–  
12 535.
- 13 Jones JW, Mair RA, Neves RJ. 2005. Factors affecting survival and growth of juvenile  
14 freshwater mussels cultured in recirculating aquaculture systems. *North American*  
15 *Journal of Aquaculture* **67**: 210-220.
- 16 McIvor A, Aldridge DC. 2008. The cultivation of the freshwater pearl mussel, *Margaritifera*  
17 *margaritifera*. Countryside Council for Wales.
- 18 Miguel ES, Monserrat S, Fernández C, Amaro R, Hermida M, Ondina P, Altaba CR. 2004.  
19 Growth models and longevity of freshwater pearl mussels (*Margaritifera margaritifera*)  
20 in Spain. *Canadian Journal of Zoology* **82**: 1370–1379.

- 1 Österling ME, Arvidsson BL, Greenberg LA. 2010. Habitat degradation and the decline of  
2 the threatened mussel *Margaritifera margaritifera*: influence of turbidity and  
3 sedimentation on the mussel and its host. *Journal of Applied Ecology* **47**: 759–768.
- 4 Österling M, Högberg JO. 2014. The impact of land use on the mussel *Margaritifera*  
5 *margaritifera* and its host fish *Salmo trutta*. *Hydrobiologia* **735**: 213–220.
- 6 Österling ME, Larsen BM. 2013. Impact of origin and condition of host fish (*Salmo trutta*) on  
7 parasitic larvae of *Margaritifera margaritifera*. *Aquatic Conservation: Marine and*  
8 *Freshwater Ecosystems* **23**: 564–570.
- 9 Preston SJ, Keys A, Roberts D. 2007. Culturing freshwater pearl mussel *Margaritifera*  
10 *margaritifera*: a breakthrough in the conservation of an endangered species. *Aquatic*  
11 *Conservation: Marine and Freshwater Ecosystems* **17**: 539–549.
- 12 Reid N, Keys A, Preston JS, Moorkens E, Roberts D, Wilson CD. 2013. Conservation status  
13 and reproduction of the critically endangered freshwater pearl mussel (*Margaritifera*  
14 *margaritifera*) in Northern Ireland. *Aquatic Conservation: Marine and Freshwater*  
15 *Ecosystems* **23**: 571–581.
- 16 Reis J. 2003. The freshwater pearl mussel [*Margaritifera margaritifera* (L.)] (Bivalvia,  
17 Unionoida) rediscovered in Portugal and threats to its survival. *Biological Conservation*  
18 **114**: 447–452.
- 19 Ricciardi A, Rasmussen JB. 1999. Extinction Rates of North American Freshwater Fauna.  
20 *Conservation Biology* **13**: 1220–1222.
- 21 Saunders DL, Meeuwig JJ, Vincent ACJ. 2002. Freshwater Protected Areas: Strategies for  
22 Conservation. *Conservation Biology* **16**: 30–41.

- 1 Scheder C, Gumpinger C. 2008. The freshwater pearl mussel (*Margaritifera margaritifera*  
2 Linné, 1758) in Upper Austria- A species treated with extinction and current measures  
3 for its sustained protection. *Romanian Journal of Biology* **52**: 53–59.
- 4 Scheder C, Gumpinger C, Csar D. 2011. Application of a five stage field key for the larval  
5 development of the freshwater pearl mussel (*Margaritifera margaritifera* Linné, 1758)  
6 under different temperature conditions- A tool for the approximation of the optimum  
7 time for host fish infection in captive breeding. *Ferrantia* **64**: 13–22.
- 8 Scheder C, Lerchegger B, Jung M, Csar D, Gumpinger C. 2014. Practical experience in the  
9 rearing of freshwater pearl mussels (*Margaritifera margaritifera*): advantages of a work-  
10 saving infection approach, survival, and growth of early life stages. *Hydrobiologia* **735**:  
11 203–212.
- 12 Schmidt C, Vandr  R. 2010. Ten years of experience in the rearing of young freshwater pearl  
13 mussels (*Margaritifera margaritifera*). *Aquatic Conservation: Marine and Freshwater*  
14 *Ecosystems* **20**: 735–747.
- 15 Strayer DL, Downing JA, Haag WR, King TL, Layzer JB, Newton TJ, Nichols SJ. 2004.  
16 Changing Perspectives on Pearly Mussels, North America’s Most Imperiled Animals.  
17 *Bioscience* **54**: 429-439.
- 18 Thomas GR, Taylor J, Garcia de Leaniz C. 2010. Captive breeding of the endangered  
19 freshwater pearl mussel *Margaritifera margaritifera*. *Endangered Species Research*  
20 **12**:1-9.
- 21 Wilson CD. 2010. Empirical approaches to the conservation of *Margaritifera margaritifera*.  
22 PhD thesis, Queen’s University Belfast.

- 1 Wilson CD, Roberts D. 2011. Modelling distributional trends to inform conservation  
2 strategies for an endangered species. *Diversity and Distributions* **17**: 182–189.
- 3 Wilson CD, Beatty GE, Bradley CR, Clarke HC, Preston SJ, Roberts D, Provan J. 2012a. The  
4 importance of population genetic information in formulating ex situ conservation  
5 strategies for the freshwater pearl mussel (*Margaritifera margaritifera* L.) in Northern  
6 Ireland. *Animal Conservation* **15**: 593–602.
- 7 Wilson CD, Roberts D, Reid N. 2011. Applying species distribution modelling to identify  
8 areas of high conservation value for endangered species: A case study using  
9 *Margaritifera margaritifera* (L.). *Biological Conservation* **144**: 821–829.
- 10 Ziuganov VV. 2004. Arctic and southern freshwater pearl mussel *Margaritifera*  
11 *margaritifera* with long and short life span as a model system for testing longevity  
12 mechanisms. *Advances in Gerontology = Uspekhi Gerontologii/ Rossiiskaia akademiia*  
13 *nauk, Gerontologicheskoe obshchestvo* **14**: 21–30.

14

1 **Table 1** Average survival rates using different methods of caged release for freshwater  
 2 mussels.

Species	Method	Cage Mechanism	Survival	Reference
<i>Villosa iris</i>	Mussel silos	See method	81-88%	Johnson <i>et al.</i> , 2014
<i>Margaritifera margaritifera</i>	Buddensiek sheet cages	Acrylic plate with drilled holes which are surrounded by a fine mesh. Each hole houses one mussel (~1mm). The box is placed upright in the river to allow constant flow of fresh water through the mesh	0.21-80%	Schmidt and Vandré, 2010; Eybe <i>et al.</i> , 2013
	Mussel cages	Modified Buddensiek sheet cages	1-7%	Hastie and Young, 2003
	Gauze bags	Gauze bags are filled with sieved gravel collected from mussel rivers. Mussels are placed in the bag which is then embedded in the river bed.	0.7-6.0%	Schmidt and Vandré, 2010
	Sediment mussel baskets	Plastic bowls are placed under a water outlet. Plastic colanders lined with gauze and filled with sieved gravel are set into the water filled bowl. Juveniles are placed in the gravel in the colander.	2.9-82%	Hastie and Young, 2003; Schmidt and Vandré, 2010

3



1 **Table 2** Generalised Linear Mixed Model (GLMM) results for **a)** survival and **b)** growth  
 2 rates of juvenile *Margaritifera margaritifera*. The function used to fit each model is shown in  
 3 parentheses. Two-way interactions were initially fitted by subsequently dropped as they were  
 4 not significant. (n.df = nominator and d.df = denominator degrees of freedom respectively).

<b>Explanatory variable</b>	<b><i>F</i></b>	<b>n.df</b>	<b>d.df</b>	<b><i>p</i></b>
<b>a) Survival (binomial logistic)</b>				
Site	4.059	1	4082	<b>0.044</b>
Size class	13.382	2	4082	<b>&lt;0.001</b>
Month (0-18 inclusive)	10.389	17	4082	<b>&lt;0.001</b>
Treatment	0.016	1	4082	0.898
<b>b) Growth rate (gamma)</b>				
Site	48.011	1	455	<b>&lt;0.001</b>
Size class	3.235	2	455	<b>0.040</b>
Period (0-6, 6-12, 12-18)	195.425	2	455	<b>&lt;0.001</b>
Treatment	0.581	1	455	0.446

5