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https://doi.org/10.1155/2014/158546

Published in: The Scientific World Journal

Document Version: Publisher's PDF, also known as Version of record

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Download date: 20. Jun. 2020
Research Article

AcT-2: A Novel Myotropic and Antimicrobial Type 2 Tryptophyllin from the Skin Secretion of the Central American Red-Eyed Leaf Frog, *Agalychnis callidryas*

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Received 13 December 2013; Accepted 8 January 2014; Published 13 February 2014

Academic Editors: A. Sacchetti and H.-S. Won

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Tryptophyllins are a diverse family of amphibian peptides originally found in extracts of phyllomedusine frog skin by chemical means. Their biological activities remain obscure. Here we describe the isolation and preliminary pharmacological characterization of a novel type 2 tryptophyllin, named AcT-2, from the skin secretion of the red-eyed leaf frog, *Agalychnis callidryas*. The peptide was initially identified during smooth muscle pharmacological screening of skin secretion HPLC fractions and the unique primary structure—GMRPPWF-NH$_2$—was established by both Edman degradation and electrospray MS/MS fragmentation sequencing. A cDNA encoding the biosynthetic precursor of AcT-2 was successfully cloned from a skin secretion-derived cDNA library by means of RACE PCR and this contained an open-reading frame consisting of 62 amino acid residues with a single AcT-2 encoding sequence located towards the C-terminus. A synthetic replicate of AcT-2 was found to relax arterial smooth muscle ($EC_{50} = 5.1$ nM) and to contract rat urinary bladder smooth muscle ($EC_{50} = 9.3$ μM). The peptide could also inhibit the growth of the microorganisms, *Staphylococcus aureus* (MIC = 256 mg/L), *Escherichia coli* (MIC = 512 mg/L), and *Candida albicans* (128 mg/L). AcT-2 is thus the first amphibian skin tryptophyllin found to possess both myotropic and antimicrobial activities.

1. Introduction

Amphibians represent the most ancient group of terrestrial vertebrates and are widely distributed globally, occurring on all continents with the exception of Antarctica [1, 2]. Their continuing persistence in the biosphere can in part be attributed to their highly developed and potent skin secretions that act as a front-line defense against potential predators and microbes [1, 2]. The biological potency of their skins and associated secretions were recognized by many ancient peoples who used them in shamanic rituals and as medicines for diverse ailments [1, 2]. Some applications of these secretions include those of bufonid toads as stimulants in traditional Chinese medicine, those of tropical poison frogs (dendrobatids) as arrow-tip poisons, and those of giant monkey frogs (*Phyllomedusa bicolor*) in “hunting magic”—a purging and sensory-enhancing ritual performed by some South-American natives prior to hunting [3].

The unlocking of the molecular secrets of amphibian defensive skin secretions was predominantly initiated by two, twentieth-century pioneer pharmacological chemists—Vittorio Erspamer (1909–1999) (endogenous peptides and biogenic amines) and John Daly (1933–2008) (exogenous alkaloids). Their combined efforts resulted in the unravelling of the molecular structures and biological actions of several thousands of such molecules representing many classes of biochemical, studies on which have served to reveal many novel regulatory systems and potential drug targets in mammalian systems [1–4].

The application of modern analytical technologies such as molecular cloning, proteomics, and mass spectrometry to the study of the molecular complexity of amphibian skin secretions continues on where the pioneers left off with ever-increasing numbers of unique molecules of diverse functions being revealed [5]. One of the major goals of such contemporary research is directed towards the identification
of natural leads for therapeutic development or to identify novel disease-related drug targets [6, 7].

The bioactive peptides represent one of the largest and most functionally diverse groups of biochemicals identified thus far from amphibian skin, and unlike the diet-acquired alkaloids, these are of endogenous origin and are genetically encoded [1–4]. While many individual peptides have been isolated, they fall into two broad functional groupings—those with pharmacological activity and those with antimicrobial activity [1–4].

Tryptophyllins (TPHs) are a large and heterogeneous group of peptides that did not fall into either category when first identified, as they were originally isolated from the skin of the South-American leaf frog, *Phyllomedusa rohdei*, through chemical means by nature of their positive Ehrlich's reaction—a color-producing chemical reaction for indoles (tryptophan residues) [2]. All original TPHs possessed a tryptophanyl residue at position 2 from the C-terminus and prolyl residues at position 2 and/or 3 from the N-terminus [2], although some TPHs more recently identified are devoid of tryptophanyl residues [2]. Due to the increasing numbers of TPHs being identified and their apparent structural heterogeneity, this peptide family has been divided into three types—(1) tryptophyllin-1 (T-1) peptides: the N-terminal doublet KP, a tryptophanyl residue at position 5 and a prolyl residue at position 7 from the N-terminus; all highly conserved in this group of peptides that typically possess 7-8 residues; (2) tryptophyllin-2 (T-2) peptides: these are variable in length, contain 4 to 7 residues, and all contain an internal PW doublet; (3) tryptophyllin-3 (T-3) peptides: the most conserved TPHs containing 13 residues with conservative substitutions at positions 2, 5, 6, and 13 from the N-terminus. The N-terminal pGlu-, –KP-, at positions 3 and 4 and substitutions at positions 2, 5, 6, and 13 from the N-terminus. conserved TPHs containing 13 residues with conservative

2. Materials and Methods

2.1. Acquisition of Skin Secretions. Adult red-eyed tree frogs, *Agalychnis callidryas* (*n* = 6), of the Costa Rican type, were housed in a purpose-designed terrarium under a 12h/12 h light/dark cycle and were fed multivitamin-loaded crickets three times per week. Skin secretions were obtained by transdermal electrical stimulation (4 ms pulse width, 50 Hz, 6 V) in accordance with the method of Tyler et al. [11] following a three-month settling-in period. The skin secretions were washed from the skin using deionized water, snap frozen in liquid nitrogen, lyophilized, and stored at −20°C prior to analyses.

2.2. Reverse Phase HPLC Fractionation of Lyophilized Skin Secretion. Five milligrams of lyophilized skin secretion were dissolved in 1 mL of 0.05/99.95 (v/v) trifluoroacetic acid (TFA)/water and clarified by centrifugation. The supernatant was directly injected onto a CECIL reverse phase HPLC system (Milton Technical Centre, Cambridge, UK) fitted with an analytical column (Phenomenex Jupiter C5; 300 Å pore size; 250 × 4 mm). The linear elution gradient employed was formed from 0.05/99.95 (v/v) TFA/water to 0.05/19.95/80.0 (v/v/v) TFA/water/acetonitrile in 80 min at a flow rate of 1 mL/min. The effluent absorbance was monitored at λ = 214 nm and fractions of approximately 1 mL were collected automatically at minute intervals. Samples (100 μL) from each chromatographic fraction were removed, lyophilized, and stored at −20°C prior to analysis for myoactivity using rat smooth muscle bioassays.

2.3. Rat Smooth Muscle Bioassays. Male Wistar rats (250–300 g) were euthanized by carbon dioxide asphyxiation followed by cervical dislocation under appropriate UK animal research licences and following local institutional guidelines. After removing the fur on the abdomen of rats, the body cavity was opened and any visible fat was removed. The exposed urinary bladder and the tail were carefully removed. All dissected tissues were placed in ice-cold Kreb’s solution (118 mM NaCl, 4.7 mM KCl, 25 mM NaHCO3, 1.15 mM NaH2PO4, 2.5 mM CaCl2, 1.1 mM MgCl2, and 5.6 mM glucose) which was aerated with 95% O2/5% CO2 gas mixture. Strips of urinary bladder and rings of dissected tail artery were prepared and tied with fine silk ligatures (0.2 mm) at each end. These preparations were attached to a fixed pin at one end and to a transducer at the other end. Kreb’s solution (at 37°C) flowing through the organ baths at 2 mL/min with a constant bubbling of 95% O2/5% CO2 maintained the tissues in a viable state for >2 h. An equilibration period of 20 min was used for each preparation after which 60 mM KCl was added to test the responsiveness of urinary bladder muscle strips and 10−4 M phenylephrine was added to test the responsiveness of the tail artery rings and to cause preconstriction. Following these tests, viable preparations were used to screen prepared samples of reverse phase HPLC fractions of *Agalychnis callidryas* skin secretion. Changes in tension of smooth muscle preparations were detected by pressure transducers connected to a PowerLab System (AD
2.4. Structural Characterization of the Novel Peptide. A single mRNA was then subjected to 5'-magnetic oligo-dT Dynabeads (Dynal Biotech, UK), as Polyadenylated mRNA was isolated from this using protection buffer that was supplied by Dynal Biotech, UK. Skin secretion were dissolved in 1 mL of cell lysis/mRNA precursor-encoding cDNA. Five milligrams of lyophilized peptide resolved was subjected to MS/MS fragmentation using a Perseptive Biosystems Voyager DE mass spectrometer (Thermo-Fisher, San Jose, CA, USA). The synthetic novel peptide on rat tail artery and urinary bladder smooth muscle. A sample of this fraction was subjected to MALDI-TOF mass spectrometry using a PS3 automated solid-phase peptide synthesizer (Protein Technologies, Inc., AZ, USA). Following cleavage of the side-chain protecting groups, the resultant material was reverse phase HPLC fraction #33 of the synthetic peptide from the resin and deprotection of primary structure of the novel peptide had been established, the optical density of supernatants following centrifugation was subjected to MALDI-TOF mass spectrometry to establish both degree of purity and authenticity of structure.

2.5. Molecular Cloning of the Novel Peptide Biosynthetic Precursor-Encoding cDNA. Five milligrams of lyophilized skin secretion were dissolved in 1 mL of cell lysis/mRNA protection buffer that was supplied by Dynal Biotech, UK. Polyadenylated mRNA was then isolated from this using magnetic oligo-dT Dynabeads (Dynal Biotech, UK), as per manufacturer's instructions. The isolated polyadenylated mRNA was then subjected to 5'- and 3'-rapid amplification of cDNA ends (RACE) procedures to obtain full-length novel peptide precursor nucleic acid sequence data using a SMART-RACE kit (Clontech UK) likewise as per manufacturer's instructions. Briefly, the 3'-RACE reactions employed a nested universal (NUP) primer (supplied with the kit) and a degenerate sense primer (SP: 3'-GGIATMGICCCICITGG-3') (I = deoxyinosine, M = A/C) that was complementary to the putative amino acid sequence, GMRPPW-, of the novel peptide. The 3'-RACE reactions were purified and cloned using a pGEM-T vector system (Promega Corporation) and sequenced using an ABI 3100 automated sequencer. The sequence data obtained from the 3'-RACE product were used to design a specific antisense primer (ASP: 5'-CGGCACTATATTGTAATTTGTGCT-3') to a defined site within the 3' nontranslated region of the novel peptide precursor-encoding transcripts. 5'-RACE was carried out using these primers in conjunction with the NUP primer and resultant products were purified, cloned, and sequenced.

2.6. Solid-Phase Peptide Synthesis. Once the unequivocal primary structure of the novel peptide had been established, it was chemically synthesized by solid-phase Fmoc chemistry using a PS3 automated solid-phase peptide synthesizer (Protein Technologies, Inc., AZ, USA). Following cleavage of the synthetic peptide from the resin and deprotection of the side-chain protecting groups, the resultant material was lyophilized and then purified by HPLC. The major product was subjected to MALDI-TOF mass spectrometry to establish both degree of purity and authenticity of structure.

2.7. Preliminary Pharmacological Characterization of the Synthetic Novel Peptide on Rat Tail Artery and Urinary Bladder Smooth Muscle. The synthetic peptide was initially prepared as a stock in Kreb's solution (118 mM NaCl, 4.7 mM KCl, 25 mM NaHCO₃, 1.15 mM Na₂HPO₄, 2.5 mM CaCl₂, 11.1 mM MgCl₂, and 5.6 mM glucose) at a concentration of 10⁻³ M. Working concentrations of peptide, ranging from 10⁻³ to 10⁻₁ M, were prepared prior to each experiment and were applied to tissues (n = 6) progressively from low to high concentrations. Data obtained from the PowerLab System (AD Instruments Pty, Ltd.) were analyzed by Student's t-test through GraphPad Prism software to obtain the mean and standard error of responses and using these datasets, dose-response curves were constructed using a best-fit algorithm.

2.8. Determination of Minimal Inhibitory Concentrations (MICs) of the Synthetic Novel Peptide for Model Microorganisms. Minimal inhibitory concentrations (MICs) of the synthetic peptide were assessed against three standard model microorganisms: the Gram-positive bacterium Staphylococcus aureus (S. aureus, NCTC 10788), the Gram-negative bacterium Escherichia coli (E. coli, NCTC 10418) and the pathogenic yeast Candida albicans (C. albicans, NCPF 1467). The synthetic peptide was initially dissolved in a small volume of dimethylsulfoxide (DMSO) and made up to 1 mL with sterile Mueller-Hinton broth (MHB). Doubling dilutions of this peptide stock solution peptide were made from 512–1 mg/L in sterile MHB and were incubated with microorganism cultures (10⁶ colony forming units (CFU)/mL) in 96-well microtiter cell culture plates for 18 h at 37 °C, in a humidified atmosphere. MHB alone was used as a negative control and each microorganism in MHB with no peptide added was used as positive controls. After incubation, the growth of microorganisms was determined by means of measuring optical density (OD) at λ = 550 nm using an ELISA plate reader (Biolise BioTek EL808). Minimal inhibitory concentrations (MICs) were defined as the lowest concentration at which no growth was detectable.

2.9. Hemolysis Assay. Peptide solutions in a range of concentrations were prepared as described in Section 2.8 but in 0.9% (w/v) aqueous NaCl solution, prior to performing the hemolysis assay. A 2% suspension of washed horse red blood cells in this solution was prepared from defibrinated horse blood (TCS Biosciences Ltd., UK) and samples of this were incubated at 37 °C for 120 min, with a range of peptide concentrations similar to those employed for the antimicrobial assays. Lysis of red cells was assessed by measurement of the optical density of supernatants following centrifugation at λ = 550 nm using an ELISA plate reader (Biolise BioTek EL808). Negative controls employed consisted of a 2% (v/v) red cell suspension alone and positive controls consisted of a 2% (v/v) red cell suspension and an equal volume of saline containing 2% (v/v) of the nonionic detergent, Triton X-100 (Sigma-Aldrich).

3. Results

3.1. Identification and Structural Characterization of the Novel Peptide. Reverse phase HPLC fraction #33 of Agalychnis callidryas skin secretion (Figure 1(a)) was found to contain...
a myoactive peptide following preliminary smooth muscle pharmacological screening and a sample of this fraction was subjected to MALDI-TOF mass spectrometry which indicated a relatively high degree of purity of a peptide with an m/z (M+H)+ of 889.25. The doubly charged ion of this peptide was subsequently identified following electrospray MS analysis and subjected to MS/MS fragmentation sequencing (Figure 1(b)). This produced the tentative amino acid sequence, GMRRPPWF, based upon identification of b- and y-ion series. The peptide was also deemed to be C-terminally amidated. Bioinformatic analysis produced no hits with any archived amphibian skin peptide but with two synthetic antifungal peptides, named PAF-26 and combi-1 [12, 13] (Figure 2(a)). Of note was the presence of residues 3–8 of the novel myotropic peptide in a C-termially-located domain of a prophenin-2-like protein from the killer whale (Orcinus orca) that contains a cathelicidin sequence (accession no. XP004284013) (Figure 2(a)). However, the origin and structural characteristics of the novel myotropic peptide, an internal -PPW-sequence and C-terminal amidation, indicated that it was a member of the amphibian skin tryptophyllin family, subtype T-2, and thus it was named Agalychnis calidryas tryptophyllin-2 (AcT-2) in accordance.

3.2. Molecular Cloning of AcT-2 Biosynthetic Precursor-Encoding cDNA. A cDNA encoding the AcT-2 precursor protein was successfully and repeatedly cloned using the RACE PCR strategy employed. The sequence was represented in at least 30 clones after employing repetitive PCR and cloning procedures. The complete nucleotide and translated open-reading frame amino acid sequences of the cloned AcT-2 biosynthetic precursor-encoding cDNA are shown in Figure 2(b). The deduced open-reading frame consisted of 62 amino acid residues and shared a similar architecture with the previously reported precursors of the tryptophyllins, PdT-1 and PdT-2 from Pachymedusa dacnicolor skin secretion (Chen et al. 2004, Wang et al. 2009) (Figure 2(c)). This consisted of an N-terminal putative signal peptide, an acidic amino acid residue-rich spacer peptide domain, a single copy of a mature AcT-2 sequence, and a C-terminal processing and amidation site (Figure 2(c)). As in the Pachymedusa dacnicolor tryptophyllin precursors, the mature peptide was flanked N-terminally by a double basic amino acid residue propeptide convertase processing site - RRR-, cleavage of which generated the N-terminus of mature AcT-2. The C-terminal region of the mature AcT-2 peptide was flanked by a tripeptide sequence, -GKK, in common with the precursor of PdT-2. The double basic amino acid motif, -KK, is removed by propeptide convertase and the resulting C-terminal glycyl (G) residue serves as an amide donor through the action of amidating enzyme complex.

3.3. Smooth Muscle Activity of Synthetic AcT-2. Synthetic AcT-2 was found to be active in smooth muscle preparations from both rat tail artery and urinary bladder but in different ways and at different potencies. In rat tail artery smooth muscle preparations, AcT-2 caused a dose-dependent relaxation with an EC50 of 5.1 nM (Figure 3(a)). In contrast, the peptide induced a dose-dependent contraction of urinary bladder smooth muscle with an EC50 of 9.3 μM (Figure 3(b)).

3.4. Antimicrobial and Hemolytic Activity of AcT-2. AcT-2 was somewhat unexpectedly found to possess a broad spectrum of antimicrobial activity. MICs obtained with the three model test organisms were as follows: S. aureus (256 mg/L), E. coli (512 mg/L), and C. albicans (128 mg/L) (Figures 4(a)–4(c)). The order of sensitivity of the test organisms employed was thus C. albicans > S. aureus > E. coli, which is a most...
unusual profile for previously reported typical amphibian skin cationic, amphipathic, \( \alpha \)-helical antimicrobial peptides. Also worthy of note was the virtual lack of hemolytic activity of AcT-2, even up to the highest concentration (512 mg/L) employed (Figure 4(d)).

### 4. Discussion

Central-American red-eyed leaf frogs (\textit{Agalychnis callidryas}) are probably the most universally recognized frogs in the world having been used extensively by advertising agencies
sequence motif (Figure 2(c)). However, despite the presence of this common internal structural feature, AcT-2 and PdT-2 display radically different potencies in stimulating the contraction of rat uterus smooth muscle (EC$_{50}$ values: AcT-2 = 9.3 µM; PdT-2 = 4 nM), indicating that other features of the primary structure influence their receptor interactions [9]. However, AcT-2 was found to possess a most potent arterial smooth muscle relaxant effect (EC$_{50}$ = 5.1 nM)—an effect not observed with PdT-2 [9]. The smooth muscle pharmacology and molecular targets for these Type-2 tryptophyllins in mammalian tissues obviously warrant further in-depth investigation.

As demonstrated in this study, AcT-2 was unexpectedly found to possess antimicrobial activity, albeit relatively low potency, using three model test microorganisms with the order of sensitivity being C. albicans > S. aureus > E. coli or in other words, yeast > Gram-positive bacterium > Gram-negative bacterium. Related to this observation was that following database interrogation with the primary structure of AcT-2, three peptides were found which displayed some degree of structural identity (Figure 2(a)). Two of these, named PAF26 and combi-1, were synthetic in nature and were found through the screening of combinatorial peptide libraries for antifungal activity [12, 13]. The third represented a cathelicidin domain of a prophenin-2-like protein deduced from a cloned cDNA of the killer whale (Orcinus orca) (accession no. XP004284013). Interestingly, all three AcT-2-like peptides were associated with antimicrobial functions, with the former two being specifically against fungi.

While AcT-2 is of relatively low potency when compared with many other amphibian skin antimicrobial peptides, most if not all of the latter act through a mechanism of microbial target cell membrane lysis and some are strongly hemolytic [17–19]. To achieve this form of nonspecific
antimicrobial action, such peptides are generally much longer in amino acid chain length than AcT-2 [17, 19]. Thus, AcT-2 may act upon a different and possibly intracellular target rather than by direct lytic action on the cytoplasmic membrane—a mode of action that has actually been proposed for some “classical” antimicrobial peptides as well [17, 19]. In support of this proposal to some degree is the lack of hemolytic activity of AcT-2 observed following its incubation with horse erythrocytes (Figure 4(d)).

In conclusion, the novel amphibian skin tryptophyllin, named AcT-2, described here, has been demonstrated to have both selective mammalian smooth muscle myotropic effects and antimicrobial activity—findings which add to our increasing knowledge of the biological effects of this enigmatic family of amphibian skin peptides.

The nucleotide sequence of AcT-2, from the skin secretion of *Agalychnis calidryas*, has been deposited in the EMBL Nucleotide Sequence Database under the accession code HG710094.

Conflict of Interests

The authors declare that they have no conflict of interests.

Authors’ Contribution

Lilin Ge and Peng Lyu contributed equally to this work.

References


