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# Susceptibility of Streptozotocin-Induced Diabetic Rat Retinal Function and Ocular Blood Flow to Acute Intraocular Pressure Challenge

Vickie H. Y. Wong,<sup>1</sup> Algis J. Vingrys,<sup>1</sup> Andrew I. Jobling,<sup>2</sup> and Bang V. Bui<sup>1</sup>

**PURPOSE.** To consider the hypothesis that streptozotocin (STZ)-induced hyperglycemia renders rat retinal function and ocular blood flow more susceptible to acute IOP challenge.

**METHODS.** Retinal function (electroretinogram [ERG]) was measured during acute IOP challenge (10–100 mm Hg, increments of 5 mm Hg, 3 minutes per step, vitreal cannulation) in adult Long-Evans rats (6 weeks old; citrate:  $n = 6$ , STZ:  $n = 10$ ) 4 weeks after citrate buffer or STZ (65 mg/kg, blood glucose >15 mM) injection. At each IOP, dim and bright flash ( $-4.56, -1.72 \log \text{cd.s.m}^{-2}$ ) ERG responses were recorded to measure inner retinal and ON-bipolar cell function, respectively. Ocular blood flow (laser Doppler flowmetry; citrate:  $n = 6$ , STZ:  $n = 10$ ) was also measured during acute IOP challenge. Retinas were isolated for quantitative PCR analysis of nitric oxide synthase mRNA expression (endothelial, *eNos*; inducible, *iNos*; neuronal, *nNos*).

**RESULTS.** STZ-induced diabetes increased the susceptibility of inner retinal (IOP at 50% response, 60.1, CI: 57.0–62.0 mm Hg versus citrate: 67.5, CI: 62.1–72.4 mm Hg) and ON-bipolar cell function (STZ: 60.3, CI: 58.0–62.8 mm Hg versus citrate: 65.1, CI: 61.9–68.6 mm Hg) and ocular blood flow (43.9, CI: 40.8–46.8 versus citrate: 53.4, CI: 50.7–56.1 mm Hg) to IOP challenge. Citrate eyes showed elevated *eNos* mRNA (+49.7%) after IOP stress, an effect not found in STZ-diabetic eyes (–5.7%,  $P < 0.03$ ). No difference was observed for *iNos* or *nNos* ( $P > 0.05$ ) following IOP elevation.

**CONCLUSIONS.** STZ-induced diabetes increased functional susceptibility during acute IOP challenge. This functional vulnerability is associated with a reduced capacity for diabetic eyes to upregulate *eNos* expression and to autoregulate blood flow in response to stress. (*Invest Ophthalmol Vis Sci.* 2013; 54:2133–2141) DOI:10.1167/iovs.13-11595

A major complication affecting many patients with diabetes is retinopathy, which is clinically characterized by vascular abnormalities, such as microaneurysms, hemorrhages, and neovascularization, which can lead to vision loss.<sup>1</sup> There is growing evidence that neuronal abnormalities might precede

microangiopathy.<sup>2</sup> For example, functional assessment of the retina using electrophysiological measures reveals changes in function<sup>3,4</sup> prior to clinically detectable microangiopathy. Indeed, Bearse et al.<sup>5</sup> and Harrison et al.<sup>6</sup> provide evidence that regions of the retina showing functional deficits are more likely to go on to show signs of microangiopathy. The presence of neuronal dysfunction may reflect subtle changes to the vasculature not readily detectable clinically. Evidence in clinical studies (none or mild nonproliferative diabetic retinopathy)<sup>7–9</sup> and in animal models suggest that changes in retinal blood flow,<sup>10</sup> impaired blood vessel reactivity,<sup>11–13</sup> and autoregulation<sup>14</sup> occur early in the course of the disease.

In addition to the development of diabetic retinopathy, the presence of vascular dysfunction or some inherent neuronal weakness may place diabetic eyes at greater risk of other neurodegenerative diseases, such as glaucoma. Glaucoma affects an estimated 60 million people worldwide<sup>15</sup> and is characterized by a progressive loss of retinal ganglion cells. The most well-defined risk factor for glaucoma is elevated intraocular pressure (IOP). Epidemiological studies have returned conflicting outcomes as to whether diabetes increases the risk of glaucoma (as reviewed in Wong et al.<sup>16</sup>). However, these studies did not set out to directly consider the association between diabetes and glaucoma. Laboratory studies provide some evidence in support of this possibility. Kanamori et al.<sup>17</sup> reported that streptozotocin (STZ)-induced hyperglycemic rats showed greater apoptosis of retinal neurons after exposure to chronic IOP elevation compared with normoglycemic rats. Kohzaki et al.<sup>18</sup> showed a reduction in the recovery of retinal ganglion cell responses 1 week after a single episode of IOP challenge, an effect not evident in citrate control rats. Whether this increased susceptibility arises from impairment to the vascular system or via a deficit in retinal neurons is not clear.

This study considered the hypothesis that STZ-induced hyperglycemia increases susceptibility of the electroretinogram and ocular blood flow to acute IOP challenge. In the same cohort of animals, we also consider a potential mechanism by comparing the effect of IOP challenge on gene expression of nitric oxide synthase (Nos)-isoenzymes (endothelial, *eNos*; inducible, *iNos*; neuronal, *nNos*) in diabetic and control eyes.

## METHODS

### Animals

All experimental procedures abide with the ARVO Statement for the Use of Animals in Ophthalmic and Vision Research and the National Health and Medical Research Council's Australian Code of Practice for the care and use of animals for scientific purposes. Animal ethics approval was obtained from the Faculty of Science Animal Ethics Committee at The University of Melbourne. Hyperglycemia was

From the Departments of <sup>1</sup>Optometry and Vision Sciences and <sup>2</sup>Anatomy and Neuroscience, The University of Melbourne, Parkville, Victoria, Australia.

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Corresponding author: Bang V. Bui, Department of Optometry and Vision Sciences, The University of Melbourne, Parkville, 3010, VIC, Australia; bvb@unimelb.edu.au.

TABLE 1. Metabolic Parameters (Blood Glucose, IOP, and Systolic Blood Pressure; Mean  $\pm$  SEM) of Control and Diabetic Animals

Week	Blood Glucose, mM $\pm$ SEM		IOP, mm Hg $\pm$ SEM		BP	SBP, mm Hg $\pm$ SEM	
	Citrate	STZ-Diabetes	Citrate	STZ-Diabetes		Citrate	STZ-Diabetes
1	5.8 $\pm$ 0.2	23.1 $\pm$ 0.9*	13.4 $\pm$ 0.4	13.4 $\pm$ 0.5	1	110.2 $\pm$ 7.6	110.3 $\pm$ 4.2
2	5.3 $\pm$ 0.2	21.6 $\pm$ 1.0*			2	92.0 $\pm$ 5.5	102.7 $\pm$ 3.9

\* Denotes significance ( $P < 0.001$ ) between citrate and STZ-diabetic animals for blood glucose (mM).

induced in 6-week-old male Long-Evans rats (Monash Animal Services, Clayton, Victoria, Australia) via a tail vein injection of 65 kg/mg STZ (MP Biomedicals, Seven Hills, New South Wales, Australia) dissolved in 0.1 M sodium citrate buffer (pH 4.5; Sigma-Aldrich Pty Ltd., Castle Hill, New South Wales, Australia). Animals were fasted overnight (~12 hours) prior to STZ-injection. Control animals were given tail vein injections of 0.1 M sodium citrate buffer (citrate group). One week following STZ-injection, blood glucose (Table 1) was measured (Ascensia Esprit Glucometer; Bayer Australia, Ltd., Pymble, New South Wales, Australia) to confirm that all STZ-treated animals had blood glucose higher than 15 mM. To prevent ketoacidosis and excessive loss of body weight, 2 units of insulin was administered each day for 2 weeks. After 2 weeks, only those animals with blood glucose higher than 20 mM continued to receive insulin injection (2 units/d). Animals were housed in a 22°C environment with a 12:12-hour light-dark cycle (maximum 40 lux), with normal rat chow (Barastoc rat chow; Ridley AgriProducts, Melbourne, Victoria, Australia) and water available ad libitum.

Electrophysiology and blood flow measurement were conducted under general anesthesia (ketamine-xylazine, 60:5 mg/kg, intramuscularly [IM]). It is worth noting that ketamine-xylazine anesthesia may have neuroprotective effects<sup>19</sup>; however, our low dose of ketamine should provide similar effects in both citrate control and STZ-injected animals. A topical anesthetic (0.5% proxymetacaine hydrochloride, eye drops, Alcaine, 5 mg/mL; Alcon Laboratories, Frenchs Forest, New South Wales, Australia) and a mydriatic (0.5% tropicamide, Mydriacyl, 5 mg/mL; Alcon Laboratories) were also given. A circulating water heat pad was used to maintain body temperature at approximately 37°C.

### Acute IOP Elevation

Baseline IOP was obtained from each animal using a noninvasive applanation tonometer (Tonopen XL; Medtronic Solan, Jacksonville, FL) using light sedation (50% dose, ketamine-xylazine, 30 mg/kg; 2.5 mg/kg, IM). This established that STZ treatment did not affect baseline IOP (Table 1). To ensure that any functional and ocular blood flow susceptibility is independent of ocular perfusion pressure (blood pressure – IOP), systolic blood pressure (SBP, Table 1) was assayed via noninvasive tail cuff sphygmomanometer (ML125; ADInstruments Pty, Ltd., Bella Vista, New South Wales, Australia). To assess ocular susceptibility to stress, one randomly selected eye underwent manometric IOP elevation from 10 to 100 mm Hg in increments of 5 mm Hg (3 minutes per step) using a height-adjustable reservoir (Hanks balanced salt solution; Sigma-Aldrich Pty Ltd.) attached to a 30-gauge cannula via polyethylene tubing. For electroretinogram (ERG) assessment, the cannula was inserted into the vitreal chamber to allow for placement of ERG electrodes. In those animals undergoing blood-flow studies, a laser Doppler flowmetry (LDF) (ML191; ADInstruments Pty, Ltd.) needle probe (MNP110XP; ADInstruments Pty, Ltd.) was inserted into the vitreal chamber and, thus, IOP was manometrically controlled via an anterior chamber cannula. Previously, we have shown that the pressure between the anterior and vitreal chamber quickly equilibrates regardless of the location of the cannula.<sup>20</sup>

### Electroretinography

Prior to functional measurements, all animals were dark-adapted overnight (approximately 12 hours) and procedures were conducted

in the dark with the aid of night-vision goggles (NVMT24021; Yukon Advanced Optics, Sellmark Corporation, Mansfield, TX) and an infrared light (IR) source ( $\lambda = 880$  nm, 18-LED IR spotlight; Jaycar, Rydalmere, New South Wales, Australia) adapted onto a dissecting microscope (Zeiss OPMI-1 Lab; Carl Zeiss Pty, Ltd., Jena, Germany).

Baseline ERG was measured 3 weeks following STZ or citrate treatment. Responses were measured across a range of calibrated (IL1700; International Light Technologies, Inc., Newbury Port, MA) luminous energies ( $-6.79$  to  $2.07$  log cd.s.m<sup>-2</sup>) delivered via a Ganzfeld integrating sphere (170 mm diameter; Photometric Solutions International, Huntingdale, Victoria, Australia). This range returns measures of photoreceptor (a-wave)<sup>21</sup> and ON-bipolar cell (b-wave).<sup>22</sup> The dimmest flashes elicit the scotopic threshold response (STR), which provides information about the integrity of retinal ganglion cells.<sup>23–25</sup>

To quantify functional susceptibility to acute IOP challenge, ERGs were recorded 4 weeks following citrate or STZ injection (1 week after baseline assessment). Two luminous energy were assayed at each step of IOP elevation. The first luminous energy was chosen to elicit an ERG dominated by a rod-driven b-wave ( $-1.42$  log cd.s.m<sup>-2</sup>), which has been widely used to assay the effects of ischemia and IOP stress.<sup>20,26,27</sup> The second, much dimmer, luminous energy ( $-5.25$  log cd.s.m<sup>-2</sup>) was chosen to elicit a retinal ganglion cell-dominated STR, a component of the ERG known to be particularly sensitive to IOP elevation.

ERGs were recorded using custom-made corneal (active) and scleral (reference, ring-shaped) chlorided silver electrodes (99.99% purity, 0.329 mm, 29-gauge; A&E Metal Merchants, Sydney, New South Wales, Australia). Both electrodes from each eye were grounded to a stainless steel electrode (F-E2-30; Grass Telefactor, West Warwick, RI) inserted subcutaneously into the tail. Eyes were lubricated after electrode placement and periodically throughout the session with 1.0% carboxymethylcellulose sodium (Celluvisc; Allergan, Irvine, CA). Signals were digitized at 4 kHz (ML785, Powerlab/8SP, 1000 $\times$  amplification; ADInstruments Pty, Ltd.), with band pass settings of  $-3$  dB at 0.3 to 1000 Hz.

### Ocular Blood Flow: Laser Doppler Flowmetry

Ocular blood flow was measured using LDF (ML191; ADInstruments Pty, Ltd.) as has been used previously.<sup>28–30</sup> This approach provides real-time continuous measurement of tissue microvascular perfusion. In the eye, LDF reflects both retinal and choroidal blood flow,<sup>27</sup> thus the term “ocular blood flow” (OBF) is used throughout the study.

The LDF needle probe (0.48 mm outer diameter  $\times$  25 mm length; MNP110XP; ADInstruments Pty, Ltd.) was inserted 2 mm behind the limbus, into the vitreal chamber. Both the LDF probe and IOP cannula (anterior chamber) were stabilized and positioned with micromanipulators (MM-3; Narishige Co., Ltd., Tokyo, Japan).

### Retinal Tissue Isolation and Real-Time PCR

On completion of in vivo experiments, eyes were enucleated and animals were euthanized (0.3 mL pentobarbitone sodium, Lethabarb; Virbac Pty, Ltd., Peakhurst, New South Wales, Australia). Retinas were excised, snap frozen in liquid nitrogen, and stored at  $-80^\circ\text{C}$  for RNA isolation. Real-time quantitative PCR was conducted, for control and IOP-treated eyes in both citrate and STZ-treated groups, to quantify mRNA expression for specific target genes of interest (*nNos*, *eNos*, and *iNos*) relative to a housekeeping gene (hypoxanthine phosphorylribo-

TABLE 2. Oligonucleotide Primers for Nitric Oxide Synthase Isoenzymes (eNos, iNos, and nNos) and the Housekeeper Gene, *HPRT*

Gene	Accession No. (PubMed.org)	Primer Sequences	Source Organism	Amplicon Size, bp
<i>eNos</i>	NM_021838.2 <sup>60</sup>	F: 5'-TGGCAGCCCTAAGACCTATG-3' R: 5'-AGTCCGAAAATGTCCTCGTG-3'	<i>Rattus norvegicus</i> (Norway rat)	243
<i>iNos</i>	NM_012611.3 <sup>60</sup>	F: 5'-CCTTGTTTCAGCTACGCCTTC-3' R: 5'-GGTATGCCCGAGTTCTTTCA-3'	<i>Rattus norvegicus</i> (Norway rat)	179
<i>nNos</i>	NM_052799.1 <sup>61</sup>	F: 5'-CCTTCCGAAGCTTCTGGCAACAGC-3' R: 5'-TGGACTCAGATCTAAGGCGGTTGG-3'	<i>Rattus norvegicus</i> (Norway rat)	474
<i>HPRT</i>	NM_000194.2 <sup>62</sup>	F: 5'-GGAGGCCATCACATTGTAGC-3' R: 5'-CCACAATCAAGACATTCTTTC-3'	<i>Homo sapiens</i> (Human)	229

F, forward; R, reverse.

syl transferase [*HPRT*]). For oligonucleotide primer sequences, see Table 2.

RNA was isolated from dissected retinal tissue using a commercially available kit (RNeasy Mini Kit; Qiagen Pty, Ltd., Doncaster, Victoria, Australia). Isolated retinas were homogenized and treated with DNase-I to eliminate possible contamination by genomic DNA. The purified RNA (10× dilution with RNase-free dH<sub>2</sub>O) was then quantified using a spectrophotometer (UV-2550; Shimadzu Corporation, Tokyo, Japan). RNA concentration was determined by the absorbance at 260 nm (RNeasy Mini Handbook for RNeasy Mini Kit, 2006; Qiagen Pty, Ltd.).

A 500-ng volume of purified RNA was transcribed to cDNA via reverse transcription (RT) reaction. An Omniscript RT Kit (Qiagen Pty, Ltd.) was used in combination with 0.5 μM oligo d(T) primers to synthesize cDNA. These samples were incubated at 37°C for 1 hour, followed by dilution in dH<sub>2</sub>O into 5 ng/μL aliquots and stored at -20°C. This was followed by RT (M-MLV Reverse Transcriptase, 200 μ/μL and Random Hexamers, 20 μg; Promega Corporation, Alexandria, New South Wales, Australia).

The resultant cDNA was amplified by using real-time PCR (Light-Cycler 3 Carousel-based system; Roche Diagnostics Australia Pty, Ltd., Castle Hill, New South Wales, Australia). The cDNA sample (1 μL) in dH<sub>2</sub>O was combined with a master mix containing magnesium chloride (MgCl<sub>2</sub>), reverse and forward primers (Sigma-Aldrich Pty, Ltd.) and a reaction medium (FastStart DNA Master SYBR Green I; Roche Diagnostics Australia Pty, Ltd.). To quantify the amount of gene transcript, an external standard (the respective PCR-amplified cDNA products) was serially diluted to produce a four-point standard curve. A gene copy number was calculated for each dilution for post hoc analysis. All real-time PCR samples were amplified in sets of triplicates to minimize handling error. PCR results were analyzed using Light-Cycler Data Analysis software (v 3.5.28; Roche Molecular Diagnostics, Pleasanton, CA).

## Data Analysis

**Photoreceptor Response.** The leading edge of the scotopic a-wave (PIII) was modeled (Equation 1),<sup>31</sup> returning a measure of photoreceptor function. The summed rod photoreceptor response (*PIII*) as a function of time (*t*, seconds) and luminous energy (*i*, log cd.s.m<sup>-2</sup>), given by a maximal amplitude (*Rm<sub>PIII</sub>*, μV), a sensitivity (*S*, log [m<sup>2</sup>.cd<sup>-1</sup>.s<sup>-3</sup>]), and a brief delay (*t<sub>d</sub>*, seconds; largely recording equipment delay),

$$PIII(i, t) = Rm_{PIII} \cdot \left[ 1 - \exp\left(-i \cdot S \cdot (t - t_d)^2\right) \right], \quad t > t_d. \quad (1)$$

This function was modeled over an ensemble of the 2 brightest luminous energies (1.95 and 2.07 log cd.s.m<sup>-2</sup>). The delay (*t<sub>d</sub>*) was 4.07 ± 0.05 and 4.15 ± 0.03 ms for citrate and STZ-treated groups, respectively. These *t<sub>d</sub>* values were then fixed; thus, only *S* and *Rm<sub>PIII</sub>* were allowed to float in the final model, which was optimized by

minimizing the sum-of-squares merit function using the Excel Solver module (Microsoft, Redmond, WA).

**Rod and Cone ON-Bipolar Cell Responses.** To isolate the ON-bipolar cell component of the ERG (PII), the modeled PIII was subtracted from the raw waveform.<sup>32</sup> PII amplitudes measured at a fixed time of 70 ms were plotted as a function of luminous energy and modeled using a saturating hyperbolic function, as given in Equation 2. The amplitude (*V*, μV) as a function of luminous energy (*i*, log cd.s.m<sup>-2</sup>) is given by a maximal amplitude (*V<sub>max</sub>*, μV), semi-saturation constant (*k*, log cd.s.m<sup>-2</sup>), where sensitivity is 1/*k* (cd<sup>-1</sup>.s<sup>-1</sup>.m<sup>2</sup>) and *n* is a dimensionless exponent that determines the slope of the function.

$$V(i) = V_{max} \frac{i^n}{i^n + k^n} \quad (2)$$

The dark-adapted ERG reflects contributions from both rod and cone pathways. Therefore, Equation 2 was fit to luminous energies below those containing cone responses. This was determined by establishing the amplitude of the “putative” rod response from a twin-flash paradigm at 2.07 log cd.s.m<sup>-2</sup>. By using a 500-ms interstimulus interval, the long refractory period of the rod system means that only cone responses are recorded in the second flash, which is then subtracted from the mixed response to return the putative rod response. Group data for rod amplitude (at 2.07 log cd.s.m<sup>-2</sup>) was compared with the amplitude of mixed responses for all other luminous energies. Amplitudes for luminous energies greater than -0.82 log cd.s.m<sup>-2</sup> were found to be significantly larger than the isolated rod PII amplitude (*P* < 0.05), indicating significant cone contribution. Thus, Equation 2 was fit to luminous energies between -6.79 to -0.82 log cd.s.m<sup>-2</sup>, as well as the isolated rod PII at 2.07 log cd.s.m<sup>-2</sup> by minimizing the sum-of-squares merit function using the Excel Solver module (Microsoft).

**Scotopic Threshold Response.** Amplitudes of the STR were evaluated for luminous energies between -6.79 and -5.25 log cd.s.m<sup>-2</sup> by taking the amplitude at a fixed time of 110 and 220 ms to return positive (pSTR) and negative (nSTR) components, respectively.

**Susceptibility to IOP Elevation.** The change in ERG or ocular blood flow at each IOP level was expressed as a percentage relative to responses collected at baseline IOP of 10 mm Hg (stress/baseline, %). As previous,<sup>20</sup> an inverse cumulative normal function (Equation 3<sup>20</sup>) was used to parametrize the relationship between relative (mean ± SEM) ERG amplitudes as a function of IOP. This function was applied assuming a maximum of 100%, to return best-fit parameters for the mean (μ) and 1/slope (σ). The mean represents the IOP level at which the response has been reduced to 50% (IOP<sub>50</sub>) and 1/slope represents the steepness of the curve.

$$f(x) = 1 - \frac{1}{\sqrt{2\pi\sigma}} e^{-\frac{(x-\mu)^2}{2\sigma^2}} \quad (3)$$

## Statistics

To compare blood glucose, baseline IOP, SBP, and ERG, as well as changes in *Nos* expression, the following statistics were employed. A Kolmogorov-Smirnov normality test and Barlett's test for equal variance were performed before comparing treatments. Data showing normality and equal variances were analyzed using parametric analysis, such as unpaired or paired *t*-test, repeated measures ANOVA (RM-ANOVA), one-way ANOVA, and two-way ANOVA. For one-way ANOVA analysis, a Dunnett's post hoc test was applied to determine significant departures from baseline. For data failing criteria for parametric statistics, two approaches were used. First, for comparisons involving multiple factors (treatment and IOP), a more conservative  $\alpha$ -value of 0.035 was used to protect against false positives. Second, for comparison of a single factor between STZ-diabetic and citrate animals, a nonparametric Mann-Whitney test was adopted.

As previous,<sup>20</sup> to compare the susceptibility of citrate and STZ-diabetic ERG and blood flow to IOP elevation, a bootstrap method was used to generate estimates of the mean and 95% confidence limits (CLs) for  $IOP_{50}$  ( $\mu$ ) and  $1/\text{slope}$  ( $\sigma$ ). This approach has the advantage that it does not make any assumptions regarding the underlying distribution of the data and provides a more robust estimate of parameter variability in transition functions, such as in Equation 3.<sup>33,34</sup> As previous,<sup>35</sup> each iterative bootstrap sample consisted of  $n$  random selections taken (with replacement) from each group. The bootstrap sample was then modeled as a function of IOP using Equation 3, to return  $IOP_{50}$  and  $1/\text{slope}$  for this particular sample. After 1000 iterations, the 25th and 97th percentiles were taken as the "bootstrap" bounds of the 95% CL. This approach allows a robust comparison of the relative sensitivity of various citrate and STZ-diabetic eyes to IOP elevation. When the best-fit parameter of the STZ-diabetic group falls outside the 95% CL of the respective parameter for the citrate group, then  $P$  is less than 0.05.

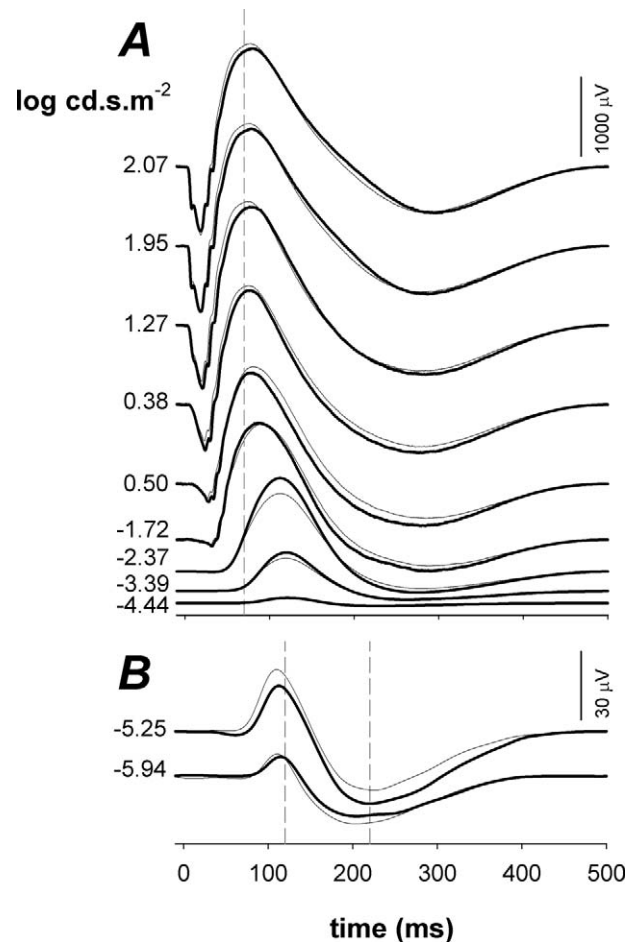
## RESULTS

Table 1 shows that at 1 ( $P < 0.001$ ) and 2 weeks following STZ injection ( $P < 0.001$ ), there was significant elevation of blood glucose in STZ-treated animals compared with citrate controls. It has been reported that systemic hyperglycemia can modify IOP and blood pressure,<sup>36</sup> which may modify ocular perfusion pressure (OPP), an important determinant of ocular blood flow. Noninvasive measurement of baseline IOP revealed no significant difference between citrate and diabetic groups ( $P = 0.932$ ; Table 1). SBP was similar when measured noninvasively at the beginning (citrate versus STZ,  $P = 0.991$ ; BP 1) and end of the IOP challenge protocol ( $P = 0.116$ ; BP 2). Although there was a small decrease in SBP from the beginning to the end of the experimental protocol, this was not statistically significant for citrate ( $P = 0.073$ ) and STZ-diabetic animals ( $P = 0.325$ ).

### Baseline ERG: STR and b-Wave

Figure 1 shows retinal function across a range of luminous energies recorded at baseline and 3 weeks following citrate (thin traces) or STZ (thick traces) injection. Three weeks of hyperglycemia had little obvious effect on the ERG in citrate and STZ-diabetic animals.

Saturated phototransduction amplitude ( $R_{mPIII}$ , Fig. 2A) and sensitivity ( $S$ ) (Fig. 2B) were not significantly different between citrate and diabetic eyes ( $R_{mPIII}$ :  $P = 0.70$ ,  $S$ :  $P = 0.76$ ). The ERG PII (reflecting ON-bipolar cell function) was assessed by fitting the luminous energy-response function for amplitudes taken at a fixed time of 70 ms with a saturating hyperbolic function (Equation 2). The saturated ON-bipolar cell amplitude was not significantly different between STZ-diabetic and citrate eyes ( $V_{max}$ ,  $P = 0.657$ ; Fig. 2C). However, there was a significant reduction in the PII sensitivity in the STZ group when compared with citrate controls ( $1/k$ :  $P = 0.006$ ; Fig. 2D).

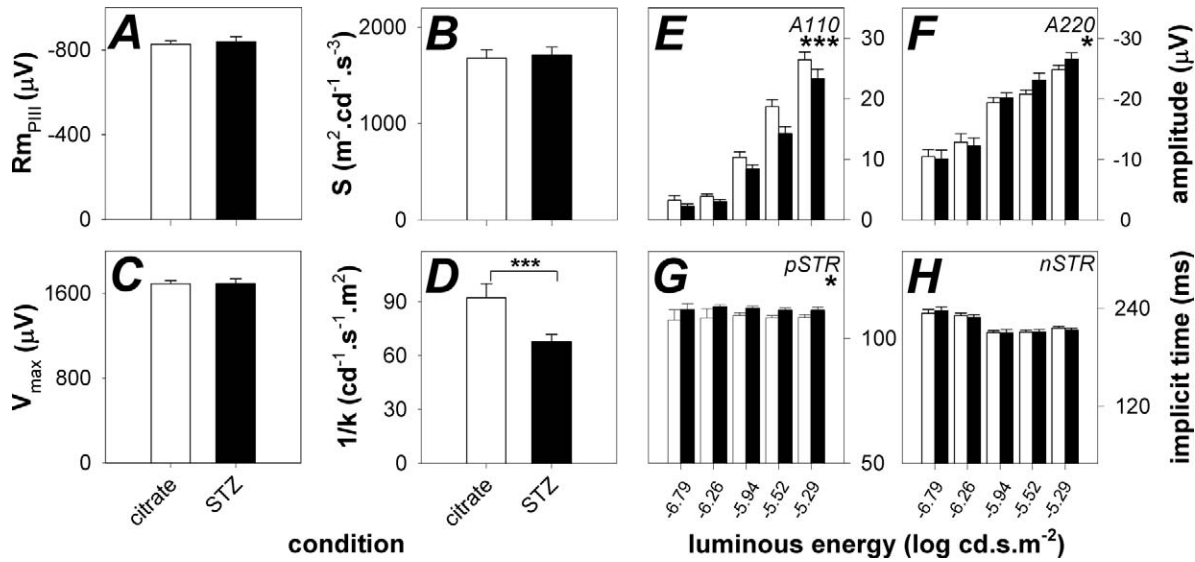


**FIGURE 1.** Comparison of retinal function between normal and diabetic groups at 3 weeks post STZ-induction. (A) Representative dark-adapted ERG waveforms for normal (thin trace) and diabetic (thick trace) groups elicited across a wide range of luminous energies ( $-4.44$  to  $2.07$  log cd.s.m<sup>-2</sup>). (B) Representative scotopic threshold responses elicited at dim light levels ( $-6.79$  to  $-5.25$  log cd.s.m<sup>-2</sup>).

The retinal ganglion cell response as given by STR amplitudes at a fixed time of 110 ms (Fig. 2E) was significantly reduced in the diabetic group ( $P < 0.001$ ). On the other hand, the amplitude at 220 ms was larger in the STZ group ( $P = 0.024$ ; Fig. 2F). The STZ-treated group also showed a delay in the pSTR ( $P = 0.016$ ; Fig. 2G); however, nSTR implicit time ( $P = 0.847$ ; Fig. 2H) was similar to citrate control eyes.

### Effect of Acute IOP on Retinal Function in STZ-Induced Diabetes

Figure 3 shows that both the ERG b-wave (Fig. 3A) and STR (Fig. 3B) responses in STZ-diabetic eyes are more sensitive to IOP elevation than the citrate group. Quantification using an inverse cumulative normal function confirmed that STR and b-wave amplitudes in STZ-diabetic eyes were significantly more susceptible ( $\mu$ , IOP at 50% response reduction,  $IOP_{50}$ ) to IOP elevation, as summarized in Table 3. Specifically, the STZ-diabetic b-wave IOP-response function was 4.8 mm Hg ( $-7.7\%$ ) more susceptible ( $IOP_{50}$ : 60.3 mm Hg, confidence interval [CI]<sub>2.5-97.5</sub>: 58.0–62.8 mm Hg; Fig. 4A) than the citrate group ( $IOP_{50}$ : 65.1 mm Hg CI<sub>2.5-97.5</sub>: 61.9–68.6 mm Hg). The STR in



**FIGURE 2.** Comparison of parameters reflecting retinal cell function for normal ( $n = 14$ , unfilled bars/symbols) and diabetic eyes ( $n = 20$ , filled bars/symbols). Averaged ( $\pm$ SEM) phototransduction saturated amplitude (A)  $R_{mPIII}$  ( $\mu V$ ) and (B) sensitivity ( $S$ ,  $m^2 \cdot cd^{-1} \cdot s^{-3}$ ). These parameters are derived from an ensemble fit of the ERG a-wave elicited at 1.95 and 2.07  $\log cd \cdot s \cdot m^{-2}$  (Equation 1). ON-bipolar cell function parameters (C) maximal amplitude,  $V_{max}$  ( $\mu V$ ) and (D) sensitivity,  $1/k$  ( $cd^{-1} \cdot s^{-1} \cdot m^2$ ) returned from a hyperbolic function (Methods, Equation 2) fitted to the averaged ERG b-wave amplitudes taken at a fixed time of 70 ms, across a range of luminous energies ( $-6.79$  to  $2.07 \log cd \cdot s \cdot m^{-2}$ ), anchored to the “putative” rod ON-bipolar cells response ( $2.07 \log cd \cdot s \cdot m^{-2}$ ). Averaged inner retinal function ( $\mu V$ , scotopic threshold response, STR) measured at fixed times of (E) 110 and (F) 220 ms. Implicit times of (G) peak (pSTR) and (H) trough (nSTR) amplitudes of STR ( $-6.79$  to  $-5.29 \log cd \cdot s \cdot m^{-2}$ ). Asterisks denote significance (\* $P < 0.05$ , \*\*\* $P < 0.001$ ).

STZ animals was 7.4 mm Hg (−11.6%) more susceptible (IOP<sub>50</sub>: 60.1 mm Hg, CI<sub>2.5-97.5</sub>: 57.0–62.0 mm Hg) in STZ-diabetic animals than the citrate group (IOP<sub>50</sub>: 67.5, CI<sub>2.5-97.5</sub>: 62.1–72.4 mm Hg; Table 3). No difference was found between the slope of the inverse cumulative normal function for the b-wave or STR (Table 3).

**Effect of Acute IOP Elevation on Ocular Blood Flow in STZ-Diabetes**

To consider whether the increased functional susceptibility observed in STZ-diabetic eyes was mediated by an impaired blood flow response to stress, ocular blood flow was measured in response to the same IOP challenge.

Similar to retinal function, ocular blood flow (Fig. 4A) in STZ-diabetic eyes was almost 10 mm Hg more susceptible to

acute IOP challenge than the citrate group (STZ, IOP<sub>50</sub>: 43.9, CI<sub>2.5-97.5</sub>: 40.8–46.8 mm Hg, filled symbols versus citrate, IOP<sub>50</sub>, 53.4, CI<sub>2.5-97.5</sub>: 50.7–56.1 mm Hg, unfilled symbols; Table 3). Table 3 also shows that no change was found in the spread of the function (or sensitivity) between the STZ-diabetic ( $\sigma$ , 24.0, CI<sub>2.5-97.5</sub>: 20.6–27.1) and the citrate group ( $\sigma$ , 26.1, CI<sub>2.5-97.5</sub>: 21.9–30.0).

Figure 4B examines the relationship between retinal function and ocular blood flow by taking the difference between the STR and blood flow. These data show that for IOPs below 80 mm Hg, blood flow is more affected than the STR amplitude. Comparison between diabetic and citrate groups suggests that for IOPs between 55 and 100 mm Hg ( $P < 0.001$ ), the gap between the STR and blood flow was smaller in STZ-diabetic eyes than citrate control eyes. This finding suggests that the relationship between blood flow

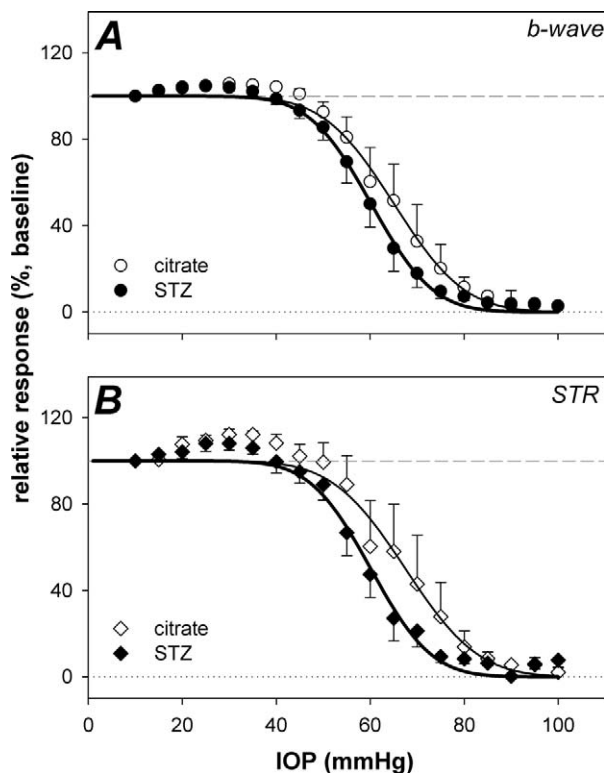
**TABLE 3.** Inverse Cumulative Normal Function Parameters (Susceptibility,  $\mu$ , and Spread,  $\sigma$ ) for Citrate and STZ-Diabetic Eyes

	Inverse Cumulative Normal Function				
	Susceptibility, Functional IOP <sub>50</sub> , $\mu$ , mm Hg			1/Slope, $\sigma$	
	Best Fit, IOP <sub>50</sub> , mm Hg	Difference	95% CI, 2.5 to 97.5%	Best Fit	95% CI, 2.5 to 97.5%
ERG					
Citrate b-wave	65.1		61.9 to 68.6	11.8	8.1 to 14.1
STZ b-wave	60.3*	4.8 (−7.7%)	58.0 to 62.8	10.4	7.4 to 12.5
Citrate STR	67.5		62.1 to 72.4	12.0	5.0 to 15.1
STZ STR	60.1†	7.4 (−11.6%)	57.0 to 62.0	10.4	5.4 to 12.3
OBF					
Citrate	53.4		50.7 to 56.1	26.1	21.9 to 30.0
STZ	43.9‡		40.8 to 46.8	24.0	20.6 to 27.1

\*  $P = 0.0038$  compared with the lower limit citrate controls.

†  $P = 0.016$  compared with the lower limit citrate controls.

‡  $P < 0.001$  compared with the lower limit citrate control.

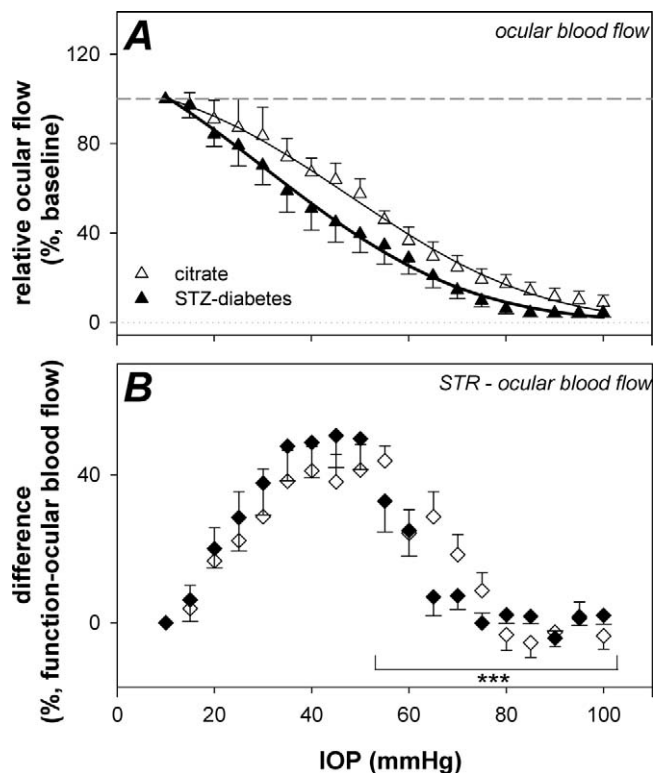


**FIGURE 3.** Comparison of relative (A) b-wave (ON-bipolar cells) and (B) STR (inner retina) averaged ( $\pm$ SEM) amplitudes as a function of IOP in citrate (unfilled symbols) and diabetic groups (filled symbols). IOP-response relationships were described with use of an inverse cumulative normal function (equation 3; citrate, *thin curve*, versus diabetic, *thick curve*) to quantify change in amplitude as a function of IOP. This function returns a best-fit  $\mu$  and  $\sigma$ , which represents sensitivity and spread, respectively. CIs for these parameters were derived from a nonparametric bootstrap method (Table 3). *Dotted* and *dashed lines* denote 0% and 100% relative amplitude, respectively.

and retinal function during stress is altered in STZ-diabetic eyes.

### Effect of STZ-Diabetes and Acute IOP on NOS mRNA Expression

Abnormal levels of Nos isoenzymes (eNos, iNos, and nNos) are thought to contribute to vascular, neuronal, and glial dysfunction.<sup>37-40</sup> Thus, we considered if the increased vulnerability to IOP challenge observed in STZ-diabetic eyes was associated with changes in nitric oxide synthase activity. Combining both control and IOP eyes, the STZ-diabetic group (Fig. 5A) for mRNA expressions for *eNos* (citrate:  $17.4 \pm 4.8$  versus STZ:  $13.3 \pm 3.9$ ;  $P = 0.153$ ), *nNos* (citrate:  $7.2 \pm 2.2$  versus STZ:  $7.2 \pm 3.1$  [ $\times 10^3$  copies];  $P = 0.463$ ), and *iNos* (citrate:  $0.6 \pm 0.2$  versus STZ:  $1.2 \pm 0.7$  [ $\times 10^2$  copies];  $P = 0.640$ ) was not significantly different between diabetic and citrate eyes. Interestingly, IOP challenge on mRNA expression (%; Fig. 5B) increased *eNos* mRNA expression by 49.7% in citrate control retinas, an effect significantly greater than the STZ-diabetic group ( $-5.7\%$ ,  $P = 0.034$ ). However, no significant difference was found for *nNos* ( $P = 0.789$ ) and *iNos* ( $P = 0.859$ ) expression between citrate and STZ-diabetic groups. In addition, the IOP-induced change in *eNos* mRNA expression was significantly correlated with the susceptibility of ocular blood flow to IOP challenge (IOP<sub>50</sub>,  $y = 0.131x +$



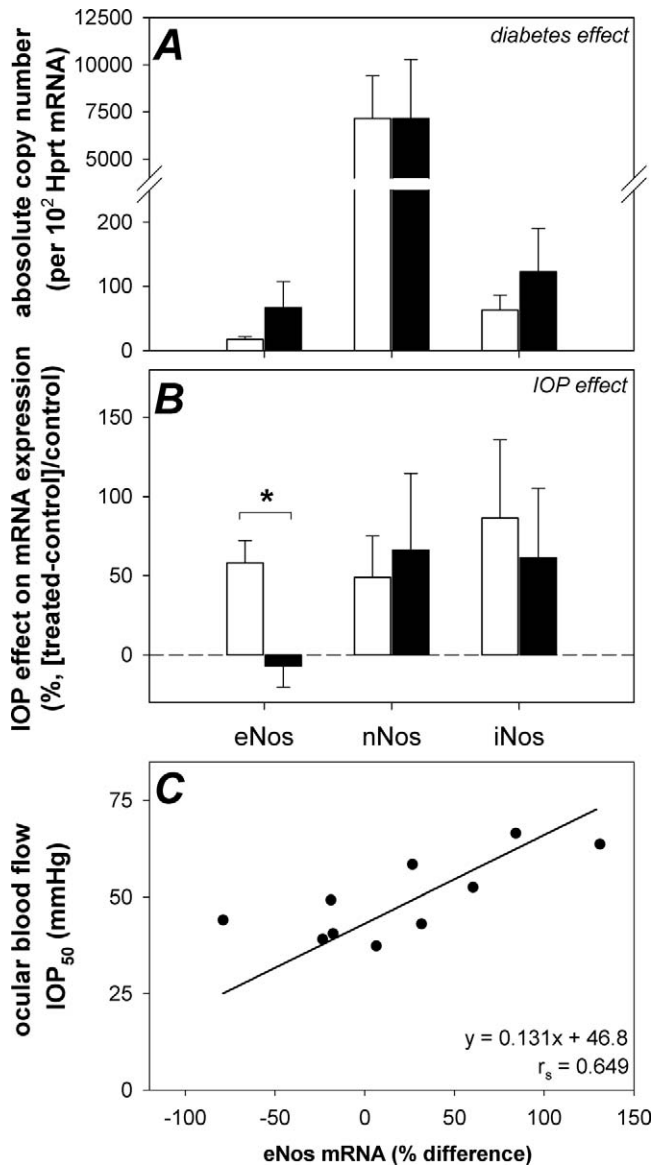
**FIGURE 4.** Effect of acute IOP challenge on ocular blood flow in normal ( $n = 8$ , unfilled symbols) and diabetic animals ( $n = 10$ , filled symbols). (A) Averaged ocular blood flow ( $\pm$ SEM) relative to each eye's own baseline (100%, *dashed line*) for normal and diabetic groups. Each *curve* was modeled with an inverse cumulative normal function (equation 3) to quantify the effect between treatment groups. CIs for these parameters were derived from a nonparametric bootstrap method (Table 3). *Dotted* and *dashed lines* denote 0% and 100% relative amplitude, respectively. (B) Difference (%) between relative inner retinal function and ocular blood flow responses across increasing IOP. *Asterisks* denote significance ( $***P < 0.001$ ).

46.8,  $r_s = 0.649$ , Spearman correlation,  $r_s = 0.649$ ,  $P < 0.05$ , Fig. 5C).

### DISCUSSION

The key finding of this study was that as early as 4 weeks following the onset of chronic hyperglycemia, before changes in the major components of the ERG,<sup>18,41-43</sup> retinal function (Fig. 3) shows greater susceptibility to acute IOP challenge. Although this is consistent with the work of Kanamori et al.,<sup>17</sup> it is inconsistent with the study of Casson and colleagues.<sup>26</sup> These authors reported that 5 days or 6 weeks post-STZ injection (60 mg/kg; blood glucose: approximately  $25 \pm 3.2$  mM), b-wave recovery at 3 to 7 days following IOP elevation to ischemic level (110 mm Hg) was better in diabetic eyes. It may be that hyperglycemia is protective against ischemic levels of IOP elevation of 110 mm Hg (as long as 60 minutes). Indeed, a "protective" effect of hyperglycemia may be important for neuronal viability under acute anaerobic stress.<sup>44</sup> The IOP used in this study only reached 100 mm Hg for 3 minutes.

The increase in ERG susceptibility to IOP elevation was associated with an increase in susceptibility in ocular blood flow (Figs. 3B, 4A). Indeed, ocular blood flow in diabetic eyes was 10 mm Hg more sensitive to IOP elevation than in the citrate group (Fig. 4), which is somewhat similar to the



**FIGURE 5.** Effect of STZ-diabetes and acute IOP challenge on *eNOS*, *nNOS*, and *iNOS* mRNA levels for citrate (unfilled bars) and STZ-diabetic eyes (filled bars). (A) Diabetes effect on absolute copy number relative to *HPRT* (averaged  $\pm$  SEM), showing combined control and IOP-treated eyes. (B) Difference in effect of IOP treatment on mRNA expression expressed relative to control eyes (%; treated-control/control) for both conditions. Dashed line denotes 0%. Asterisk denotes significance ( $*P < 0.05$ ). (C) Relationship between susceptibility of ocular blood flow to IOP elevation (IOP<sub>50</sub>) and IOP-induced changes in *eNOS* mRNA expression (%). A Deming linear regression, which takes into account variability in both *y*- and *x*-axes and is thus more robust for false positives and Spearman correlations (coefficient,  $r_s$ ) were employed to describe the relationship (black line) between parameters.

increase in STR susceptibility of 7.4 mm Hg. This outcome is consistent with a previous study<sup>45</sup> using the blue field entoptic technique to show that macular blood flow was compromised with smaller IOP elevation in patients with mild or moderate retinopathy (increase of 11 and 8 mm Hg, respectively) compared with healthy individuals (15 mm Hg). This study suggested that the effective autoregulatory range is compromised with increasing severity of diabetic retinopathy. It is worth noting that the LDF measure of blood flow receives

contributions from both retinal and choroidal circulations, thus care must be taken when attributing the observed changes to retinal autoregulation.

Impaired autoregulation, as evidenced by diminished retinal vessel reactivity in the eye in response to perturbations, such as oxygen breathing<sup>11</sup> and flickering light,<sup>9,12,13,46</sup> have been reported in patients with diabetes. Impaired autoregulation may be associated with vascular changes, such as loss of pericytes and endothelial cells (as reviewed by Fletcher et al.<sup>47</sup> and Hammes<sup>48</sup>),<sup>49</sup> which are important in structural support and autoregulation, respectively. Endothelial cells are an important source of both endothelin and nitric oxide; thus, their absence will impair autoregulation.

Several studies suggest that the bioavailability of nitric oxide is altered by hyperglycemia. This change in nitric oxide production has been attributed to disruption of *eNOS* and *nNOS* activity.<sup>50</sup> In this study, acute IOP challenge produced a substantial increase in *eNos* mRNA expression in citrate control animals but not STZ-rats ( $-5.7\%$ ). It appears that an IOP-induced increase in *eNos* is associated with better blood flow resistance to acute IOP challenge (Fig. 5C). Previous studies have reported that well before the onset of endothelial cell loss, and as early as 1 and 2 weeks after treatment, there is detectable elevation of retinal *eNos* and *nNos* levels in STZ-treated rat eyes compared with healthy controls.<sup>51,52</sup> Although the data were not significant, there was a trend for increased *eNos* expression in STZ-diabetic eyes (Fig. 5A). It has been suggested that hyperglycemia-induced elevation of protein kinase C activity could lead to the overproduction of nitric oxide,<sup>53</sup> possibly via upregulation of *eNos*. It has also been demonstrated that in diabetic mice lacking *iNos* there was a significant reduction in the number of acellular capillaries and pericyte ghosts.<sup>49</sup> Thus, *iNOS*<sup>54</sup> is also likely to play an important role in the increased sensitivity to IOP challenge observed in our study.

Although our data argue that the functional susceptibility arises from a vascular impairment, we cannot rule out the possibility that in the STZ-diabetic eyes there is (1) reduced neuronal redundancy due to cell loss or (2) hyperglycemia-induced inherent neuronal susceptibility to IOP-related stress. Although retinal cell loss has been described in laboratory and clinical studies,<sup>55-57</sup> such changes are usually noted 6 weeks after STZ-injection.<sup>17,56</sup> Kanamori et al.<sup>17</sup> reported an 8- to 9-fold increase in apoptotic cell death after 1 month of STZ-diabetes in rats, of which the majority of these TUNEL (terminal transferase dUTP nick end labeling)-positive cells localized to the inner retinal layers. Thus, reduced neuronal redundancy could contribute to our observations. Consistent with this possibility, the pSTR was reduced at baseline in our STZ-diabetic rats, as has been reported elsewhere.<sup>41,43</sup> Moreover, as IOP increased above 55 mm Hg, the buffer between the STR amplitude and blood flow was reduced in STZ-diabetic eyes compared with citrate controls (Fig. 4B). This might suggest an additional neuronal susceptibility above the observed impairment in blood flow autoregulation. An inherent neuronal susceptibility might arise from increased oxidative stress as evidenced by the reversal of axonal transport and ion regulation abnormalities in mice overexpressing superoxide dismutase-2<sup>58</sup> and superoxide dismutase-1,<sup>59</sup> respectively.

## CONCLUSIONS

Our study shows that retinal function in STZ-diabetic eyes is more vulnerable to acute IOP challenge. This increased susceptibility occurs very early in the course of STZ-induced



diabetes, prior to severe retinal dysfunction. This was associated with increased vulnerability of ocular blood flow to IOP elevation, as well as a reduced capacity for diabetic eyes to upregulate eNos in response to IOP stress.

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