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Significance of *tagI* and *mfd* genes in the virulence of non-typeable *Haemophilus influenzae*

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Summary. Non-typeable *Haemophilus influenzae* (NTHi) is an opportunist pathogen well adapted to the human upper respiratory tract and responsible for many respiratory diseases. In the human airway, NTHi is exposed to pollutants, such as alkylating agents, that damage its DNA. In this study, we examined the significance of genes involved in the repair of DNA alkylation damage in NTHi virulence. Two knockout mutants, *tagI* and *mfd*, encoding N\(^3\)methyladenine-DNA glycosylase I and the key protein involved in transcription-coupled repair, respectively, were constructed and their virulence in a BALB/c mice model was examined. This work shows that N\(^3\)-methyladenine-DNA glycosylase I is constitutively expressed in NTHi and that it is relevant for its virulence. [Int Microbiol 2014; 17(3):159-164]

Keywords: *Haemophilus influenzae* · alkylating agents · virulence · genes *tagI* and *mfd*

Introduction

Non-typeable *Haemophilus influenzae* (NTHi) is a commensal gram-negative bacterium well adapted to the human upper respiratory tract [7]. It has been implicated in the etiology of otitis media, conjunctivitis, sinusitis, pneumonia, and chronic bronchitis, and in the progression of chronic obstructive pulmonary disease (COPD) [19]. However, within its human host, this opportunistic pathogen is exposed to high levels of genotoxic stress in the form of airway pollutants. In a study based on proteomic expression profiling of *H. influenzae* grown in pooled sputum from adults with COPD, both the expression of antioxidant activity and stress responses were shown to be important for NTHi survival in the airways [13].

DNA-damaging agents are ubiquitous. They are generated endogenously during cell metabolism and are present in the environment—in air, water and foods—although generally in low concentrations. For example, tobacco smoke contains a mixture of alkylating agents, some of which act directly (alkyl halides, acrolein, crotonaldehyde, ethylene oxide, propylene oxide, acrylonitrile, and acrylamide), while others act indirectly (requiring metabolic transformation to form reactive species) [15]. Moreover, human airway pollutants such as tobacco smoke damage not only eukaryotic cells but also the DNA of the respiratory tract microbiota.

The repair of DNA alkylation damage in bacterial cells has been mainly studied in *Escherichia coli*. As in other
bacteria, *E. coli* has two specific mechanisms to remove alkyl radicals from its DNA: (i) via the constitutive expression of genes encoding the necessary repair enzymes and (ii) via the alkyl-induced expression of these proteins [16]. This adaptive response to the repair of DNA alkylation damage is regulated by the Ada protein, a positive transcriptional regulator that stimulates the expression of the *ada*, *alkA*, *alkB*, and *aidB* genes [5,16]. Bacteria also have two additional enzymes involved in the specific repair of DNA alkylation damage: Ogt (*O^\text{6}-\text{meG}-\text{DNA} \text{ methyltransferase}*) [10] and TagI (*N^\text{3}\text{meA}-\text{DNA} \text{ glycosylase I}* ) [2]. In addition, two other systems are involved in the repair of DNA alkylation damage: the nucleotide excision repair (NER) [20] and the transcription-coupled repair (TCR) [17] systems. The latter system mediates the bulk repair of DNA damage via the Mfd protein, followed by the engagement of NER.

The aim of the present work was to determine the significance of *tagI* and *mfd* genes involved in the repair of DNA alkylation damage in *NTHi* virulence. Accordingly, knockout mutants in *tagI*, specific for DNA alkylation damage, and *mfd*, involved in bulk DNA repair, were constructed and their virulence in a BALB/c mouse model was studied.

### Materials and methods

**Bacteria, media, and growth conditions.** *Haemophilus influenzae NTHi375*, an otitis media isolate [4], was grown on chocolate agar + PolyViteX plates (PVX, BioMerieux), on brain heart infusion (BHI) medium with or without agar supplemented with 10 μg hemin ml⁻¹, and 10 μg NAD ml⁻¹ (sBHI). The cultures were grown at 37 °C for 18 h in an atmosphere of 5 % CO₂. *Escherichia coli* DY380 strain was grown in LB (Luria–Bertani) broth or on agar plates at 37 °C for 18 h. When necessary, 50 μg ampicillin ml⁻¹ and 50 μg spectinomycin ml⁻¹ were added.

**Construction of *tagI* and *mfd* knockout mutants.** The *tagI* knockout mutant was constructed from strain NTHi375 using a previously described method [18]. Briefly, the entire gene targeted for deletion was PCR-amplified from the genomic DNA of NTHi375 strain (Table 1, Fig. 1A), cloned into pGEM-T (Promega), and electroporated into *E. coli* DY380. Strain DY380 harboring the plasmid with the *tagI* gene was selected by plating onto LB agar plates supplemented with 50 μg ampicillin ml⁻¹. Then, with plasmid pRSM2832 [18] as template, PCR was used to generate an amplicon from which the *tagI* gene was cloned into pGEM-T (Promega). The *mfd* knockout mutant was constructed from strain NTHi375 using a previously described method [18]. Briefly, the entire gene targeted for deletion was PCR-amplified from the genomic DNA of NTHi375 strain (Table 1, Fig. 1A), cloned into pGEM-T (Promega), and electroporated into *E. coli* DY380. Strain DY380 harboring the plasmid with the *mfd* gene was selected by plating onto LB agar plates supplemented with 50 μg ampicillin ml⁻¹. Then, with plasmid pRSM2832 [18] as template, PCR was used to generate an amplicon from which the *mfd* gene was cloned into pGEM-T (Promega).

### Table 1. Oligonucleotide primers used in this study

<table>
<thead>
<tr>
<th>Primer</th>
<th>Sequence (5'→3')</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Primers used to obtain the mutants</strong></td>
<td></td>
</tr>
<tr>
<td>TagI_F</td>
<td>cggtgtgcaacgaatca</td>
</tr>
<tr>
<td>TagI_R</td>
<td>tctgtgaaaccttatttgaaac</td>
</tr>
<tr>
<td>Mfd_F</td>
<td>tacactatgctcaaatattttatca</td>
</tr>
<tr>
<td>Mfd_R</td>
<td>acaatgatggtgctcttttatag</td>
</tr>
<tr>
<td>P1-TagI</td>
<td>ggtttgacpaacsctatttatatttatttgattcatcagacaggaaatgagctgaatttgcagacgaacgaaaccgcatttatttcggggatccgtcgacc</td>
</tr>
<tr>
<td>P2-TagI</td>
<td>aacatttattatagtgtgtgatttcggggatccgtcgacc</td>
</tr>
<tr>
<td>P1-Mfd</td>
<td>ccatttttaaagaggtactttttgcttgtgtggatccgtcgacc</td>
</tr>
<tr>
<td>P2-Mfd</td>
<td>attttttaaagaggtactttttgcttgtgtggatccgtcgacc</td>
</tr>
<tr>
<td><strong>Primers used for RT-qPCR assays</strong></td>
<td></td>
</tr>
<tr>
<td>trpA_F</td>
<td>cttcgtgcctgctggtaacc</td>
</tr>
<tr>
<td>trpA_R</td>
<td>cttcgtgcctgctggtaacc</td>
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<td>Tag_F</td>
<td>cgccaaataataaggctgtgacc</td>
</tr>
<tr>
<td>Tag_R</td>
<td>cgggtaggctgtgtggatt</td>
</tr>
<tr>
<td>recA_F</td>
<td>cagttcggcaacggtagt</td>
</tr>
<tr>
<td>recA_R</td>
<td>cagttcggcaacggtagt</td>
</tr>
</tbody>
</table>

Forward primer, up; reverse primer, dw. Underlined text corresponds to the 80 nucleotides of the 5' and 3' ends (H1 and H2) of the *NTHi* gene to be deleted.
TAGI AND MFD GENES IN NON-TYPEABLE H. INFLUENZAE

containing a cassette with both the rpsL and the spectinomycin resistance genes, flanked by FRT (FLP recombinase target) sites. In addition, the design of the primers produced an amplicon that contained 80 nucleotides (nt) sequences of the 5′ and 3′ ends of the tagI gene to be deleted, and P1 and P2 are 20-nt sequences of DNA homologous to the 5′ and 3′ ends of the cassette, respectively. (C) The inserted region of the amplicon of panel B in the chromosome of the NTHi375 knockout tagI mutant.

NTHi infection BALB/c model. To infect the mice, the bacteria were recovered with 1 ml of PBS from a chocolate-agar plate grown for 16 h, yielding a bacterial suspension of ~5 × 109 colony-forming units (CFU)/ml. Twenty microliters of bacteria (~107 CFU) were inoculated into the nares of 5- to 7-week-old female BALB/c mice (Harland Iberica). After 48 h of infection, the mice were killed by cervical dislocation and their lungs were rapidly dissected for the determination of bacterial load. The dissected lungs were homogenized on ice in 500 μl of PBS using an Ultra-Turrax T10 basic homogenizer (IKA). Bacteria from the homogenates and from serial dilutions thereof were recovered on chocolate-agar plates. The results are reported as log CFU per gram of tissue. In each case, clones recovered from the mice were confirmed by PCR.

The mice were treated in accordance with the Directive of the European Parliament and of the Council on the protection of animals used for scientific purposes (Directive 2010/63/EU) and in agreement with the Bioethical Committee of the University of the Balearic Islands. This study was approved by the Bioethical Committee of the University of the Balearic Islands under authorization number 1748.

Reverse transcription–quantitative real-time PCR. RNA from strain NTHi375 grown in sBHI and treated or not with 1.5 μg N-methyl-N′-nitro-N-nitrosoguanidine (MNNG) ml−1 for 1 h was extracted using the RNeasy minikit (Qiagen) and DNase treatment (Ambion). Reverse transcription–quantitative real-time PCR (RT-qPCR) was performed in a 20-μl reaction mixture with Lightcycler RNA Master SYBR Green I (Roche) on a
Based on these results, we considered whether tagI expression was inducible by DNA alkylation damage. Cultures of NTHi375 were treated with a sublethal concentration (1.5 μg/ml) of MNNG for 1 h after which the expression of tagI was determined by RT-qPCR. Expression of the recA gene served as a positive control. The results showed that, as in other bacteria [13], the expression of NTHi375 tagI was constitutive because it was not further induced by MNNG treatment, whilst the expression of recA gene was induced by a factor of 3.6 (Fig. 4).

These observations indicated that the 3-methyladenine DNA glycosylase I activity encoded by the tagI gene is crucial for NTHi 375 survival during lung infection. Similar to AlkA, TagI is a monofunctional glycosylase of the base excision repair system; as such, it hydrolytically cleaves the glycosidic bond of alkylated purine bases. However, unlike AlkA, TagI has a very high specificity because it almost exclusively cleaves 3-methyladenine [2,6]. This specificity probably arises from the enzyme’s unique aromatic-residue-rich 3-MeA binding pocket and the absence of the catalytic aspartate that is present in all other helix-hairpin-helix family members, including AlkA [6].

Our results contrast with those reported for Salmonella enterica, in which inactivation of the ada, ogt, tag, uvrB, and mfd genes is necessary to decrease bacterial virulence when the cells are orally inoculated in mice [1]. It has been suggested that the extensive alkylation repair system of Salmonella is involved in the survival of Salmonella cells outside the infected animal, enabling them to overcome the potentially massive...
By contrast, DNA injuries induced by alkylating agents present in the environment [1]. By contrast, H. influenzae is a human obligate pathogen well adapted to the human upper respiratory tract, and with a low persistence outside the host [8], which would explain why it does not have the full complement of repair mechanisms needed to repair alklylation-type damage. Consequently, the deletion of a key protein in the repair of alkylation injuries must be more relevant for this species than it is for Salmonella. In this context, the role of N4-meA-DNA glycosylase I in NTHi survival in human airways must be emphasized, because this enzyme catalyzes the specific removal of N4-methyladenine, a mainly lethal insult that blocks DNA replication [3,9]. In the setting of tobacco smoke and NTHi respiratory infections [10], the bacterial TagI protein would thus be critical in repairing DNA damage caused by the alkylating agents in cigarette smoke during the infective process of NTHi.

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**Competing interests.** None declared.

**References**


