



**QUEEN'S
UNIVERSITY
BELFAST**

Macrophage Inducible C-Type Lectin As a Multifunctional Player in Immunity.

Patin, EC., Orr, SJ., & Schaible, UE. (2017). Macrophage Inducible C-Type Lectin As a Multifunctional Player in Immunity. *Frontiers in immunology*, 8, [861]. <https://doi.org/10.3389/fimmu.2017.00861>

Published in:
Frontiers in immunology

Document Version:
Publisher's PDF, also known as Version of record

Queen's University Belfast - Research Portal:
[Link to publication record in Queen's University Belfast Research Portal](#)

Publisher rights

Copyright © 2017 Patin, Orr and Schaible. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms

General rights

Copyright for the publications made accessible via the Queen's University Belfast Research Portal is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy

The Research Portal is Queen's institutional repository that provides access to Queen's research output. Every effort has been made to ensure that content in the Research Portal does not infringe any person's rights, or applicable UK laws. If you discover content in the Research Portal that you believe breaches copyright or violates any law, please contact openaccess@qub.ac.uk.



Macrophage Inducible C-Type Lectin As a Multifunctional Player in Immunity

Emmanuel C. Patin^{1,2†}, Selinda Jane Orr² and Ulrich E. Schaible^{1*}

¹Priority Area Infections, Department Cellular Microbiology, Forschungszentrum Borstel, and German Center for Infection Research, TTU-TB, Borstel, Germany, ²Division of Infection and Immunity, School of Medicine, Cardiff University, Cardiff, United Kingdom

OPEN ACCESS

Edited by:

Uday Kishore,
Brunel University London,
United Kingdom

Reviewed by:

Bernd Lepenies,
University of Veterinary Medicine,
Germany
Taruna Madan,
National Institute for Research
in Reproductive Health, India

*Correspondence:

Ulrich E. Schaible
uschaible@fz-borstel.de

[†]Present address:

Emmanuel C. Patin,
Retroviral Immunology, The
Francis Crick Institute,
London, United Kingdom

Specialty section:

This article was submitted to
Molecular Innate Immunity,
a section of the journal
Frontiers in Immunology

Received: 28 April 2017

Accepted: 07 July 2017

Published: 25 July 2017

Citation:

Patin EC, Orr SJ and Schaible UE
(2017) Macrophage Inducible
C-Type Lectin As a Multifunctional
Player in Immunity.
Front. Immunol. 8:861.
doi: 10.3389/fimmu.2017.00861

The macrophage-inducible C-type lectin (Mincle) is an innate immune receptor on myeloid cells sensing diverse entities including pathogens and damaged cells. Mincle was first described as a receptor for the mycobacterial cell wall glycolipid, trehalose-6, 6'-dimycolate, or cord factor, and the mammalian necrotic cell-derived alarmin histone deacetylase complex unit Sin3-associated protein 130. Upon engagement by its ligands, Mincle induces secretion of innate cytokines and other immune mediators modulating inflammation and immunity. Since its discovery more than 25 years ago, the understanding of Mincle's immune function has made significant advances in recent years. In addition to mediating immune responses to infectious agents, Mincle has been linked to promote tumor progression, autoimmunity, and sterile inflammation; however, further studies are required to completely unravel the complex role of Mincle in these distinct host responses. In this review, we discuss recent findings on Mincle's biology with an emphasis on its diverse functions in immunity.

Keywords: macrophage inducible C-type lectin, trehalose dimycolate, anti-inflammatory, phagocytosis, innate immunity

INTRODUCTION

Upon initial encounter with infectious invaders or cellular stress conditions, the host defense immune system has to rapidly recognize pathogens or danger signals as potentially harmful. For this purpose, innate immune cells, such as macrophages, dendritic cells (DCs), and neutrophils (PMN) use a limited number of pattern recognition receptors (PRRs), including toll-like receptors (TLRs), nucleotide-binding oligomerization domain-like receptors, and C-type lectin receptors (CLRs), which activate immediate anti-microbial effectors or other defense mechanisms. These PRRs can sense conserved structural motifs of both microbial and endogenous danger signals, i.e., pathogen-associated molecular patterns (PAMPs) and damage-associated molecular patterns (DAMPs), respectively (1, 2). The macrophage-inducible C-type lectin (Mincle) belongs to the CLR family, along with Dectin-1 (CLEC7A), Dectin-2 (CLEC6A), macrophage C-type lectin (MCL) (CLEC4D), macrophage mannose receptor (MMR, CD206) as well as dendritic cell-specific intercellular adhesion molecule-3 grabbing non-integrin (DC-SIGN, CD209) (1–3). Initial reports demonstrated that Mincle, similar to Dectin-2 and MCL, associates with the Fc receptor gamma (FcγR) signaling chain, which contains an immunoreceptor tyrosine-based activation motif (ITAM). Phosphorylation of the associated FcγR ITAM leads to recruitment of spleen tyrosine kinase (Syk), which activates Card9-Bcl10-Malt1-mediated NF-κB signaling pathway and subsequent expression of pro-inflammatory cytokines (2, 4, 5).

Over recent years, Mincle gained growing interest as shown by an exponentially increasing number of publications on its complex biology. Excellent reviews have already emphasized the

beneficial role of Mincle during infection in triggering pro-inflammatory responses as well as its recognition of the mycobacterial ligand, trehalose-6,6'-dimycolate (TDM or cord factor), and the putative utilization of synthetic homologs as better adjuvants for vaccination (5–9). Based on recent studies, we discuss herein Mincle's expression patterns, rapidly expanding ligand diversity, and interactions with ligand relevant for adjuvant design as well as novel facets of Mincle's functions in immune modulation, phagocytosis, cancer surveillance, and autoimmunity.

MINCLE'S FINE-TUNED EXPRESSION IN IMMUNE CELLS

Macrophage inducible C-type lectin was first identified in the late 1990s by Matsumoto and colleagues who observed increased expression of the Mincle encoding CLEC4E gene in macrophages following stimulation with inflammatory agents such as TLR agonists, including LPS, but also pro-inflammatory cytokines such as IFN γ , TNF- α , and IL-6 (10). Importantly, the increase in Mincle expression was dependent on the nuclear factor NF-IL6 and MyD88 (10–12). To date, Mincle has been shown to be mainly expressed on myeloid cells, including macrophages, monocytes, DCs, and PMN (5, 10, 13–15) although one report also described Mincle expression on human B cell subsets (16). Induction of Mincle expression was observed upon infection with mycobacteria (17, 18), *Leishmania major* (*L. major*) (19) and treatment with the *Agrocybe aegerita* lectin (AAL) (20). In addition to TLR-mediated signals, induction of Mincle expression was also shown to depend on MCL (CLEC4D). In contrast to Mincle, which is weakly expressed in the absence of inflammatory stimuli, myeloid cells were shown to express MCL constitutively in macrophages and DCs (21), whereas MyD88 was critical for its surface translocation (12). Notably, MCL and Mincle share ligands such as TDM, which strongly suggests synergy between both receptors and a two-step control of the downstream signaling pathway (21). Recent investigations indicate that MCL and Mincle form a complex heterodimer that is translocated to the cell surface during microbial challenge (12, 22, 23). One could speculate that a certain threshold of MCL ligands further enhances sensing of those triggers by increasing Mincle expression to better regulate host responses. Of note, Schoenen and co-workers recently demonstrated the critical function of the early growth response transcription factor C/EBP β for the expression of Mincle in macrophages, which failed to respond to TDB/TDM stimulation in the absence of this factor (11).

Little is known about the molecular mechanisms involved in the negative modulation of Mincle expression. However, one group recently showed that a cocktail of GM-CSF and IL-4 can downregulate Mincle mRNA transcripts in human DCs (24). Follow-up investigations from the same group extended previous observations by showing that IL-4 inhibits Mincle expression during *in vitro* differentiation of murine bone marrow-derived macrophages and DCs in a STAT-6-dependent manner (25). Interestingly, stimulation by the TLR4 ligand, LPS, counteracts IL-4-mediated suppression of Mincle expression in myeloid cells.

MINCLE, A PROMISCUOUS SENSOR OF DIVERSE STIMULI

Diverse structures originating from both the mammalian host as well as microbes have been identified as ligands for Mincle. Mincle thereby acts as a PRR for infectious as well as endogenous inflammatory conditions. All currently described putative ligands of Mincle are listed in **Table 1**. However, some of these ligands most likely require further biochemical evaluation in order to pivotally clarify whether these are true Mincle ligands or whether their observed reactivities were due to minor contaminations with other PAMPs, a lesson learned from studies on putative TLR2 and TLR4 ligands (26, 27).

Macrophage-inducible C-type lectin has been shown to sense mammalian cell components or DAMPs such as the histone deacetylase complex unit SAP130 alarmin, a protein released from damaged and necrotic cells (4, 28). Recombinant SAP130 triggers pro-inflammatory cytokine secretion such as MIP-2 from peritoneal macrophages in a Fc γ R chain-dependent manner (4). The relevant role of Mincle for endogenous inflammatory conditions is corroborated by concomitant upregulation of Mincle, SAP130, and phospho-Syk expression in ischemia (29), pancreatic ductal adenocarcinoma (PDA) (28), as well as ethanol-induced liver injury (30). More recently, Nagata and coworkers identified β -glucosylceramide, an ubiquitous intracellular metabolite also released from damaged cells, as a ligand for Mincle (31). Accordingly, this endogenous component was able to induce immunostimulatory functions in myeloid cells, which was abrogated in the absence of Mincle. In addition, crystalline cholesterol present in atherosclerotic plaques, inflammatory foci associated with macrophage infiltrates, was shown to bind to human, but not mouse Mincle, thereby inducing pro-inflammatory cytokines (32). Furthermore, it was recently reported that Mincle could sense cholesterol sulfate in a sterile skin inflammation model in mice (33).

In addition to binding DAMPs, Mincle also interacts with PAMPs from various microbes. Initial studies reported Mincle as the receptor for the mycobacterial cell wall glycolipid TDM (6).

TABLE 1 | Microbial or endogenous ligands of macrophage inducible C-type lectin (Mincle).

Mincle ligand	Origin	Selected publications
Sin3-associated protein 130	Damaged and necrotic cells	Yamasaki et al. (4)
β -glucosylceramide	Damaged cells	Nagata et al. (31)
Cholesterol crystals	Bovine liver	Kiyotake et al. (32)
Cholesterol sulfate	Skin epithelium	Kostarnoy et al. (33)
Trehalose-6,6'-dimycolate	Mycobacteria	Ishikawa et al. (47); Schoenen et al. (48)
Glycerol monomycolate (GroMM)	Mycobacteria	Hattori et al. (49)
β -gentiobiosyl diacylglycerides	<i>M. tuberculosis</i>	Richardson et al. (50)
Glucosyl-diacylglycerol	<i>Streptococcus pneumoniae</i>	Behler-Janbeck et al. (44)
α -glucosyl diglyceride	<i>Lactobacillus plantarum</i>	Shah et al. (46)
Brautemycin	<i>Nonomuraea</i> spp.	Jacobsen et al. (92)
<i>Agrocybe aegerita</i> lectin	<i>Agrocybe aegerita</i>	Zhang et al. (20)

Consequently, Mincle was also found to be involved in protective immunity to both, *Mycobacterium bovis* bacillus Calmette–Guérin (BCG) and *M. tuberculosis* Erdman. However, an independent study demonstrated that Mincle is dispensable for the control of *M. tuberculosis* H37Rv in mice (15, 17, 34). The different routes of administration, i.e., systemic versus airway, might explain the discrepancies between those reports. Differences in TDM content or localization between the mycobacterial strains used in those studies may also affect Mincle sensing and subsequent innate responses. In addition to Mincle, TDM can also engage MCL (21). Recent investigations have revealed a critical function of MCL in *M. tuberculosis* infection, which was demonstrated by enhanced mortality rates and aggravated inflammation in MCL-deficient mice when compared to their WT counterparts (35).

Macrophage inducible C-type lectin has also been described as important player in immunity to fungal pathogens such as *Candida* (*C.*) *albicans* and *Malassezia* species (36, 37). However, the fungal cell wall component acting as a ligand for Mincle has still not yet been fully identified although two distinct glycolipids isolated from *Malassezia* were shown to bind Mincle (38) (Table 2).

Over recent years, the list of pathogens recognized by Mincle has expanded tremendously, including *Streptococcus* (*S.*) *pneumoniae*, *Fonsecaea* (*F.*) *monophora*, *Helicobacter* (*H.*) *pylori*, *L. major*, *Pneumocystis carinii* as well as different *Corynebacterium* strains (19, 39–43). From many of these pathogens, the specific ligands that bind Mincle are yet unidentified. However, one group recently reported Mincle as receptor for glucosyl-diacylglycerol from *S. pneumoniae*, and this interaction was demonstrated to determine the outcome of experimental pneumococcus infection in mice (44). It was also shown that a galectin isolated from *Agrocybe aegerita* mushroom could act as ligand for Mincle (45). In addition, Mincle has also been described as a PRR for cyclopropane-fatty acid α -glucosyl diglyceride, a product of the commensal probiotic *Lactobacillus* (*L.*) *plantarum* (46). Therefore, Mincle is not only involved in sensing infectious or endo-inflammatory conditions but may also act as sensor for changes in microbiota community compositions and may, therefore, contribute to microbiota associated immunomodulation. Taken together, Mincle recognizes a heterogeneous array

of ligands most of them not identified yet. Whether Mincle's promiscuous ligand interactions are based on common structural analogies is awaiting further studies (Table 2).

As supported by numerous investigations, TDM is the best Mincle ligand characterized. Interaction between Mincle and TDM triggers pro-inflammatory cytokine secretion in macrophages (11, 47). Consequently, Mincle-deficient mice showed impaired production of inflammatory cytokines and chemokines upon infection with *M. bovis* BCG or exposure to oil-in-water emulsion containing TDM (15, 48). *In vitro*, deletion of Mincle impairs TDM-mediated induction of pro-inflammatory cytokine release by murine macrophages (47). Another mycolic acid derivatized compound, glycerol monomycolate (GroMM), derived from mycobacteria was recently identified as a Mincle ligand (49). Interestingly, further structurally unrelated *M. tuberculosis* metabolites, β -gentiobiosyl diacylglycerides, are sensed by murine but not human Mincle. However, a synthetic truncated β -glucosyl diglyceride emerged as a superior agonist of murine as well as human Mincle (50).

To summarize, the list of endogenous and exogenous ligands recognized by Mincle is steadily growing supporting the critical role of this innate immune receptor in detecting various stimuli under stress conditions.

MINCLE ACTIVATION: A PROMISING PATH FOR THE DESIGN OF NEW ADJUVANTS

Macrophage inducible C-type lectin ligands including various DAMPs and PAMPs, but also synthetic ones such as TDB, are promising adjuvants for vaccine therapies (24, 51–53). Complete Freund's adjuvant (CFA), which was extensively used for optimal immunization in animal models, also contains mycobacterial cell wall glycolipids including TDM but is to inflammatory for usage in humans.

As critical step in designing new adjuvants, the mechanism of ligand recognition by Mincle has been investigated intensively over recent years. The ultimate aim is to design synthetic Mincle ligands, which retain their robust adjuvancy while their exacerbated inflammatory properties are reduced for safer usage.

The structure–function relationships during molecular Mincle–ligand interactions have already been the subject of excellent reviews (5, 54, 55). To date, structural studies have identified various ligand-binding sites in Mincle (56–59). A glycolipid ligand has been shown to interact with Mincle *via* three binding sites: (I) a primary canonical Ca^{2+} based C-type binding site with similarity to sites in other C-type lectins; (II) a proximal secondary binding site for a glucose residue in the trehalose headgroup; (III) a shallow hydrophobic region, which can bind acyl chains and is in close proximity to the putative sugar binding site in a similar manner as in MCL (56, 59). The latter two binding sites likely act in concert to interact with both, the hydrophilic sugar head group as well as the fatty acid chains of glycolipid ligands, which may modulate affinity and/or specificity. Recently, Feinberg and co-workers demonstrated that the affinity of Mincle for hydrophobic ligands correlates with the number of carbon atoms. This was irrespective of the ligand's fatty acid

TABLE 2 | Future questions on macrophage inducible C-type lectin (Mincle)'s immune functions.

1. Which are the physicochemical properties of *Helicobacter pylori*, *Fonsecaea monophora*, *Leishmania major*, *Candida albicans*, and *Pneumocystis carinii* ligands recognized by Mincle?
2. Can the promiscuity of Mincle's ligand interactions be explained by structural analogies between ligands?
3. Is Mincle recruited to the phagocytic synapse together with other receptors?
4. Are Mincle SNPs associated with increased susceptibility to cancers and autoimmune diseases?
5. Is Mincle involved in autoimmunity in the absence of respective ligands during the induction phase?
6. Is targeting Mincle a strategy to cure infections, cancers, and autoimmune diseases?
7. Is Mincle playing a role in sterile inflammation?

side chain structures, i.e., linear, branched, aromatic rings, which could be responsible for a certain level of promiscuity in Mincle's interactions with hydrophobic ligands. In addition, the structural conformation of Mincle displays a certain degree of flexibility, which might influence the structural properties of the hydrophobic groove and, therefore, ligand binding (58). However, further studies are required to fully understand the structural basis of Mincle's promiscuity in ligand binding.

Several studies have investigated the potential of synthetic ligands to induce Mincle-mediated immune responses *in vitro* and *in vivo*. Ostrop and co-workers demonstrated strong Mincle-mediated cytokine production from human primary macrophages and DCs in response to TDB/TDM stimulation *in vitro* (24). Synthetic mycolic acids, such as trehalose, glucose, and arabinose monomycolates, were shown to induce production of cytokines and reactive oxygen species as well as surface expression of costimulatory molecules in DCs through Mincle binding (60). Immunization of mice with liposome-based antigen preparations containing synthetic Mincle ligands, including trehalose diesters and monoesters led to robust antigen-specific Th1/Th17 responses (51). Rationally designed Mincle ligands, namely 2-tetradecyloctadecanoic acid and (GlcC14C18) mannose 2-tetradecyloctadecanoic (ManC14C18), have been shown to possess potent immunoprotective activity against *M. tuberculosis* infection (61).

MINCLE MODULATES INFLAMMATORY RESPONSES

The role of Mincle as a receptor for pathogenic stimuli, which subsequently triggers innate pro-inflammatory responses, is well established [reviewed in Ref. (5)]. However, recent reports indicate that Mincle rather than purely inducing pro-inflammatory responses is an immune modulator as its engagement also promotes expression of anti-inflammatory cytokines and counter regulates pro-inflammatory signaling pathways (62).

Enhanced secretion of IL-10 has been reported to increase susceptibility of mice to infections by mycobacterial and fungal pathogens (63–67). Moreover, blocking IL-10 production enhanced the protective efficacy of BCG vaccination against *M. tuberculosis* infection (68). Zhang and colleagues demonstrated the ability of neutrophils to secrete IL-10 in response to TDM and concomitant TLR signals (69) and IL-10 production by DCs was also reported upon stimulation with TDB (70). Recently, we revealed the critical role of Mincle and FcγR-Syk signaling pathway for induction of IL-10 secretion *in vitro* and subsequent downregulation of IL-12p40 in response to TDM or *M. bovis* BCG (18). It should be noted that IL-10 secretion as induced by TDM requires a concomitant TLR2 signal, which is provided by whole mycobacteria containing lipoproteins.

Other studies have also revealed an important role for Mincle in the induction of IL-10 expression in BMDMs or BMDCs upon infection with *Malassezia* spp. (37, 38). Moreover, Mincle was shown to induce anti-inflammatory IL-10 responses in human THP-1 cells challenged with *H. pylori* (40). Considering the role of Mincle in regulating IL-10 production, it would be

interesting to analyze whether inhibition of IL-10 at the same time of vaccination using TDB as an adjuvant can increase protection to the respective infections. Finally, it remains to be elucidated whether engaging Mincle signaling can trigger expression of other anti-inflammatory cytokines other than IL-10 such as TGF-β and IL-27. However, our preliminary studies failed to reveal production of IL-27 upon TDB stimulation *in vitro* (71).

Besides induction of anti-inflammatory cytokine secretion, Mincle has recently been implicated in the downregulation of pro-inflammatory signals. Investigations from Wevers and colleagues revealed that *F. monophor*a engagement of Mincle inhibits Dectin-1-mediated IL-12p35 responses in human DCs (41, 72). Similar results were obtained with TDB, which was shown to interfere with LPS-mediated induction of IL-12p35. Interestingly, *F. monophor*a-mediated inhibition of IL-12p35 was abrogated upon treatment of human DCs with wortmannin, a pharmacological inhibitor of phosphoinositide-3-kinase (PI3K). Inhibition of the PI3K effector molecule PKB (or Akt) also blocked Mincle suppression. The authors further concluded that Mincle-mediated activation PI3K/PKB signaling leads to impaired protective Th1 immunity *via* proteasomal degradation of interferon regulatory transcription factor (IRF)-1. Interestingly, another study revealed Mincle-mediated suppression of Dectin-2/FcγR/Card9-induced Th17 response during *F. monophor*a infection (73). Indeed, while Dectin-2-deficient mice displayed impaired IL-17 production in lymph nodes of infected mice, this was abrogated in Mincle-deficient animals, thereby revealing a dual function of Mincle and Dectin-2 during fungal infections. More recently, an alternative inhibitory Mincle/FcγR/SH2-domain-containing inositol polyphosphate 5' phosphatase (SHP-1) signaling pathway was described in *L. major* infection (19). It was demonstrated that Mincle could impair DC activation through SHP-1 recruitment. As a consequence, Iborra and colleagues observed less inflammatory lesions and more robust IFNγ-producing-CD4⁺ T cell responses in the absence of Mincle. Finally, Mincle-deficient mice were less susceptible to experimental Leishmaniasis as shown by decreased parasite burden when compared to their WT counterparts (19).

Other studies have reported Mincle-mediated suppression of TLR2 and TLR4 signaling. Indeed, it was shown that interference with Mincle signaling significantly increased the production of pro-inflammatory cytokines such as TNF-α and IL-6 in splenocytes, BMDCs, and BMDMs upon LPS stimulation. Mincle-deficient mice were likewise more susceptible to an endotoxic shock syndrome-associated pro-inflammatory cytokine storm when compared to their WT counterparts (74). While we observed that TDM-Mincle-mediated IL-10 production in macrophages downregulated TLR2-induced IL-12p40 secretion (18), Mincle-mediated suppression of TLR4-induced cytokine secretion was independent of IL-10. Furthermore, Mincle-deficient splenocytes were shown to express less inhibitory proteins such as SOCS1, A20, and ABIN3, but increased amounts of the TLR4 co-receptor, CD14, in response to LPS. Increased CD14 expression in the absence of Mincle signaling was responsible for enhanced LPS-induced cytokine production (74).

Finally, Lee and co-workers recently revealed a contribution of Mincle to enhanced NOS-2 expression, which involves p38-mediated

eIF5A hypusination (45). This mechanism subsequently inhibits the NLRP3 inflammasome and caspase-1-dependent IL-1 β secretion by murine macrophages through elevated nitric oxide (NO) production (45). Upon TDM injection, NOS-2-deficient mice developed larger granulomas than wild type ones.

Thus, as depicted in **Figure 1**, recent investigations have shown that Mincle can act as immune modulator in different models by either triggering anti-inflammatory responses or downregulating pro-inflammatory signals.

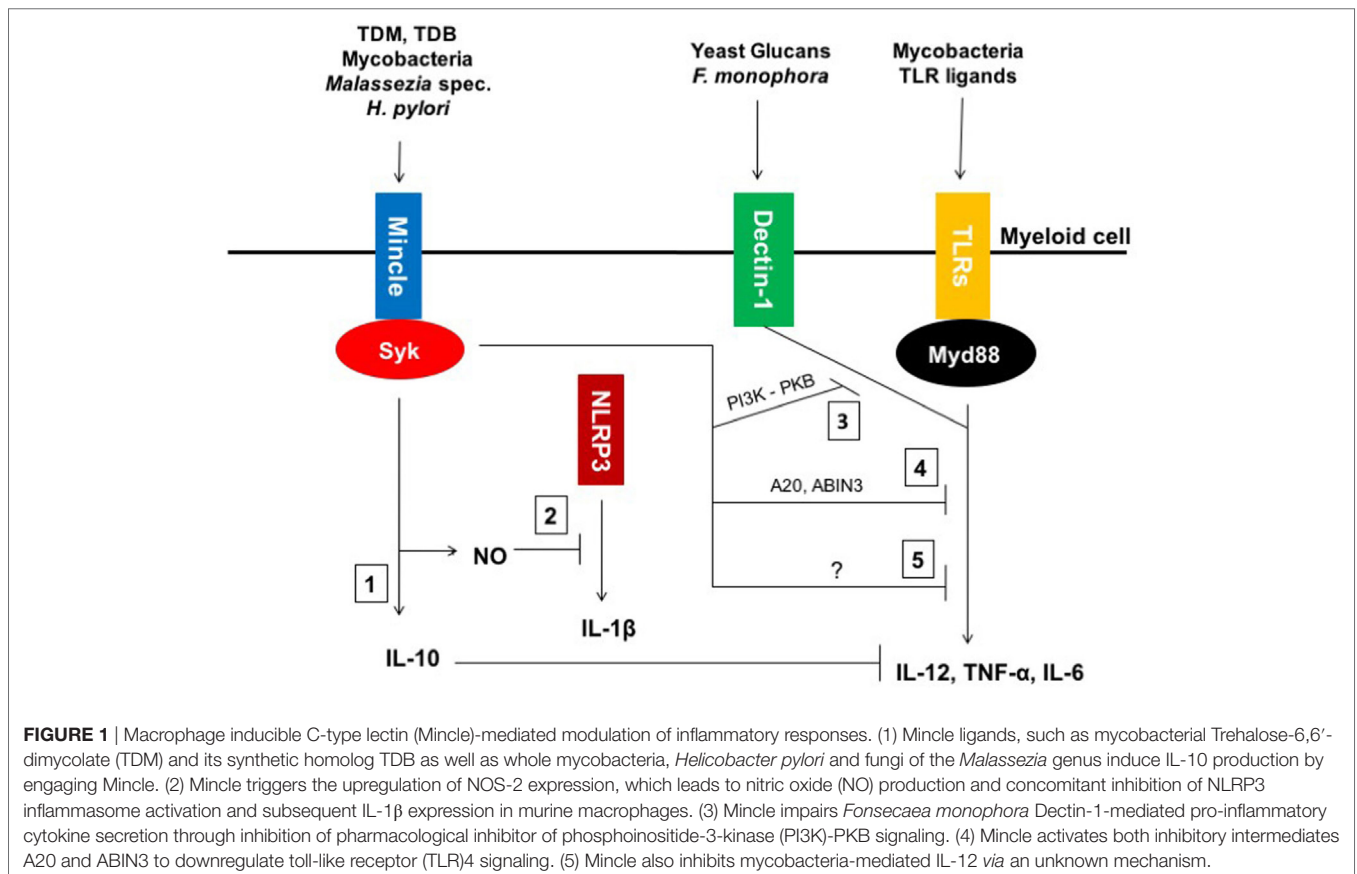
IS MINCLE A MODULATOR DURING PHAGOCYTOSIS?

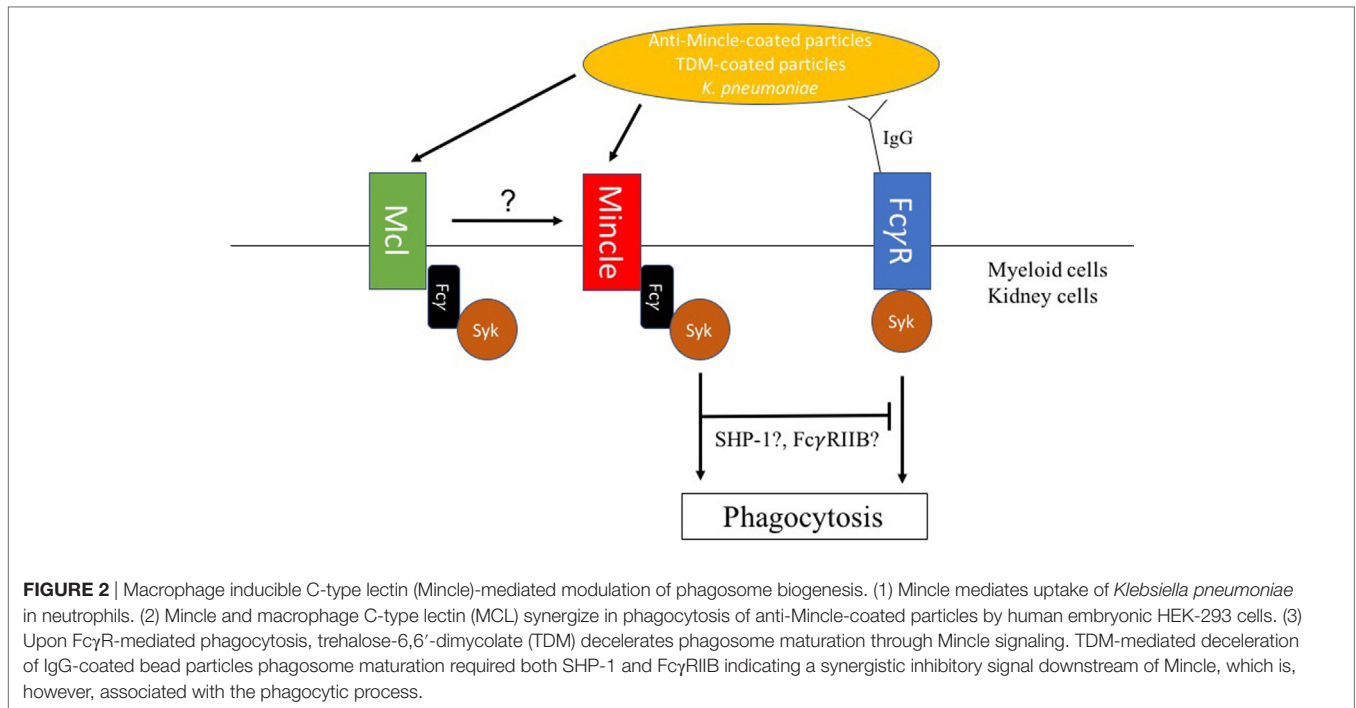
Phagocytosis is an essential effector mechanism in innate immunity to eliminate pathogens or infected cells. Moreover, in multicellular organisms, phagocytosis is part of the maintenance system to assure tissue homeostasis by clearing necrotic and apoptotic cells (75). Several studies raised the question whether Mincle could be involved in phagocytosis similar to other Syk-coupled CLRs such as Dectin-1 (76, 77). However, this question has not yet been satisfactorily answered.

An early report from Wells and colleagues suggested Mincle as a non-phagocytic receptor in *C. albicans* infection (36). Although the authors described significant recruitment of Mincle to yeast-containing phagosomes in murine macrophages, Mincle was not required for fungal uptake although confocal

microscopy analysis demonstrated localization of Mincle around *C. albicans* at the newly formed macrophage phagocytic cup (36). However, neither treatment of macrophages with an anti-Mincle blocking antibody nor using Mincle-deficient macrophages altered the number of phagocytosed yeasts when compared with untreated or control macrophages, respectively (36). From those observations, the authors concluded that the role of Mincle was restricted to sensing *C. albicans* concomitantly with the phagocytic process. An exclusive sensing function of Mincle was also observed in the *S. pneumoniae*–macrophage interaction. Using a recombinant Mincle–Fc fusion protein, Rabes et al. reported that Mincle was capable of binding heat-inactivated *S. pneumoniae* but was dispensable for pneumococcus internalization by professional phagocytes, including macrophages and neutrophils, and for inducing either pro-inflammatory cytokine secretion (39).

In contrast, Mincle was implicated as a receptor for non-opsonic phagocytosis of *Klebsiella pneumoniae* (78). Indeed, lower phagocytosis rates of non-opsonized *K. pneumoniae* were observed by flow cytometry in Mincle-deficient neutrophils when compared with WT control neutrophils (**Figure 2**). Lobato-Pascual and colleagues investigated the role of Mincle in phagocytosis of anti-Mincle coated particles by 293T cells transfected with Mincle (**Figure 2**) (23). Phagocytosis of anti-Mincle-coated beads was detected in cells expressing Mincle, but interestingly, phagocytosis was synergistically increased





in cells expressing both MCL and ITAM-containing Fc γ R in addition to Mincle. Those results showed the importance of close association between Mincle, MCL, and Fc γ R for optimal uptake of anti-Mincle-coated particles. Importantly, MCL has also been reported to be involved in the uptake of mycobacteria by neutrophils, which was taken as explanation for the higher susceptibility to experimental tuberculosis of MCL-deficient mice (12).

Cell wall glycolipids have been shown to interfere with phagocytosis through modulation of phagosome maturation following PRR surface recognition. For instance, mannose-capped lipoarabinomannan (ManLAM) engaging the MMR during phagocytosis was suggested as a mechanism to direct and maintain ManLAM-coated particle in early endosomal phagosomes in human macrophages (79). Similarly, higher-order phosphatidylinositol mannosides (PIMs) such as PIM₆ or glycopeptidolipids were shown to inhibit phagosome maturation by binding to the MMR (80, 81). Importantly, TDM has been proposed as one of the main mycobacterial glycolipids responsible for interference with phagosome maturation. Indrigo and colleagues showed that reconstitution of delipidated mycobacteria with purified TDM restored the ability of the bacteria to inhibit trafficking of phagosomes to lysosomes (82). We and others described delayed maturation of phagosomes containing TDM-coated beads when compared with uncoated beads (82, 83). Phagosomes containing TDM-coated beads retained early endosomal characteristics for a longer time period than phagosomes containing beads coated with the TDM precursor, trehalose monomycolate, or non-related lipids. This indicates that the fatty acid chain is a determinant for TDM-mediated interference with phagosome maturation. Notably, IFN- γ -activation of macrophages overcomes TDM-induced delay of phagosome maturation through NO-mediated

alteration of TDM's hydroxyl residues (83). Those observations suggested a potential role of Mincle during phagocytosis of TDM-coated particles. However, a recent study utilizing several fluorescent trehalose glycolipid reporter systems showed that Mincle was not involved in the uptake of TDM or TDM-coated particles (84), which reinforced the hypothesis of a non-phagocytic role of Mincle. However, Mincle was involved in decelerating bead phagosome maturation, when TDM beads were additionally opsonized with specific IgG (**Figure 2**). Therefore, parallel engagement of Mincle interferes with Fc γ R receptor initiated phagosome biogenesis (85). A recent study by Iborra and colleagues demonstrated that Mincle employs a SHP-1-coupling cellular signaling pathway to dampen adaptive immunity to *L. major* infection (19). Interestingly, our investigations showed that TDM-mediated deceleration of phagosome maturation also required SHP-1 suggesting an inhibitory signal downstream of Mincle during the phagocytic process (**Figure 2**). TDM-mediated inhibition of IgG bead phagosome maturation additionally required the inhibitory Fc γ RIIB, which suggests collaboration between Mincle and Fc γ RIIB receptor signaling pathways in modulating phagocytosis of IgG-opsonized TDM-containing particles (**Figure 2**) (85). Corroborating this notion, association between Dectin-1 and Fc γ RIIB was previously shown to be required for the inhibition of neutrophil function by IgG1 immune complexes (86). In the context of Fc γ RIIB's role in delaying IgG-TDM bead phagosome maturation, it should be noted that mice lacking Fc γ RIIB were less susceptible to *M. tuberculosis* infection than wild-type ones (87). However, whether Mincle can also modulate phagosome biogenesis upon phagocytosis of particles, which expose other ligands on their surface requires further studies. As mycobacteria triggers strong antibody responses (88), Mincle function upon BCG vaccination would need to be reassessed.

MINCLE AS AN ENDOGENOUS SENSOR IN CANCER AND AUTOIMMUNITY

Macrophage inducible C-type lectin can be exploited as a target for TDB adjuvant to boost vaccine efficacy but also for antitumor immunotherapies (89). However, recent studies also indicate a “natural” role of Mincle in sensing tumors and promoting their progression.

Pioneering studies by Roperto and colleagues revealed increased Mincle expression in bovine urothelial tumor cells when compared to healthy individuals (90). The authors concluded that upregulation of Mincle in bladder cancer cells can influence interactions between tumor and immune cells. The anti-TB vaccine strain BCG is successfully used clinically as an immunotherapy against bladder cancer. The underlying immune mechanisms are not well understood yet, but BCG derived TDM might be involved in this mechanism. The natural trehalose-derived metabolite, brartermicin, a new pharmacological inhibitor of murine colon carcinoma cells has been also described as a Mincle ligand (91, 92). Furthermore, the likely Mincle ligand AAL was described to exert significant inhibitory activities toward various murine and human tumor cell lines such as mouse sarcoma S-180 and HeLa, respectively (20, 93).

A recent study demonstrated a detrimental role of Mincle in pancreatic tumorigenesis in a model of PDA in mice (28). Seifert and co-workers revealed that Mincle expression in tumor-infiltrating myeloid-derived suppressor cells, macrophages, and DCs can promote necrosome-induced accelerated oncogenesis through ligation of the cellular alarmin SAP130. Notably, pancreatic oncogenesis was decelerated in Mincle-deficient mice, and treatment of mice with TDB can drive tumorigenesis, thereby confirming the deleterious effect of Mincle in PDA.

Besides promoting tumor progression, Mincle also seems to be detrimentally involved in exacerbation of certain autoimmune diseases. In T cell-mediated human autoimmune hepatitis as well as its experimental murine counterpart, SAP130 was strongly enhanced and hepatic innate inflammatory cells overexpressed Mincle triggering exacerbated inflammation, whereas interference with Mincle signaling protected against autoimmune hepatitis (94). Experimental autoimmune uveoretinitis (EAU) develops in mice immunized with the endogenous retinal protein interphoto-receptor retinoid binding protein in CFA. Consequently, mice deficient for Mincle but no other C-type lectins were protected against EAU. This observation indicates that Mincle and its Syk/Card9 signaling pathway can promote the development of experimentally induced autoimmune reactions

(95). However, it should be noted that CFA, which is frequently used for immunization with autoantigens in order to break tolerance, contains mycobacterial cell wall constituents including TDM. Therefore, it is questionable whether Mincle is involved in triggering autoimmunity under natural conditions when TDM or other Mincle ligands are absent.

CONCLUDING REMARKS

Over the last decade, great progress has been achieved to understand the role of Mincle in immunity. Recent investigations have demonstrated that immune activation using rationally designed synthetic Mincle ligands represents an interesting strategy to shape immune responses toward robust protection against infectious diseases. However, originally categorized as a pro-inflammatory receptor involved in sensing pathogens and necrotic cells, Mincle is now also considered as important regulatory element in inflammation by promoting anti-inflammatory cytokines and subsequent downregulating pro-inflammatory responses. Moreover, Mincle can also determine signaling events controlling phagocytosis and downstream phagosome maturation. Thereby, Mincle can be exploited by mycobacteria through their virulence-associated glycolipid, TDM, to decelerate phagosome maturation. The multifunctional aspect of Mincle in immune responses becomes even more evident by recent observations that it can promote progression of certain tumors, autoimmune reactions, as well as sterile inflammation. The detrimental role of Mincle in these pathological conditions opens up new possibilities of targeting Mincle signaling as potential immune modulatory therapies.

AUTHOR CONTRIBUTIONS

EP, US, and SO participated in conceptualization, writing, review, and editing of the manuscript. US and SO contributed to funding acquisition.

FUNDING

This work was supported by grants to U.S. from the Wellcome Trust UK (WT082825) and the German “Bundesministerium für Bildung und Forschung” (BMBF) program “Medical Infection Genomics” (0315834C-D), the Deutsche Zentrum für Infektionsforschung (TTU-TB), and to SO. through the Sir Henry Dale Fellowship jointly funded by the Wellcome Trust and the Royal Society (Grant Number 099953/Z/12/Z) (SO).

REFERENCES

- Brown GD. Sensing necrosis with Mincle. *Nat Immunol* (2008) 9:1099–100. doi:10.1038/ni1008-1099
- Kingeter LM, Lin X. C-type lectin receptor-induced NF-kappaB activation in innate immune and inflammatory responses. *Cell Mol Immunol* (2012) 9:105–12. doi:10.1038/cmi.2011.58
- Hardison SE, Brown GD. C-type lectin receptors orchestrate antifungal immunity. *Nat Immunol* (2012) 13:817–22. doi:10.1038/ni.2369
- Yamasaki S, Ishikawa E, Sakuma M, Hara H, Ogata K, Saito T. Mincle is an ITAM-coupled activating receptor that senses damaged cells. *Nat Immunol* (2008) 9:1179–88. doi:10.1038/ni.1651
- Richardson MB, Williams SJ. MCL and Mincle: C-type lectin receptors that sense damaged self and pathogen-associated molecular patterns. *Front Immunol* (2014) 5:288. doi:10.3389/fimmu.2014.00288
- Matsunaga I, Moody DB. Mincle is a long sought receptor for mycobacterial cord factor. *J Exp Med* (2009) 206:2865–8. doi:10.1084/jem.20092533
- Graham LM, Brown GD. The dectin-2 family of C-type lectins in immunity and homeostasis. *Cytokine* (2009) 48:148–55. doi:10.1016/j.cyto.2009.07.010
- Drummond RA, Saijo S, Iwakura Y, Brown GD. The role of Syk/CARD9 coupled C-type lectins in antifungal immunity. *Eur J Immunol* (2011) 41:276–81. doi:10.1002/eji.201041252

9. Lang R. Recognition of the mycobacterial cord factor by Mincle: relevance for granuloma formation and resistance to tuberculosis. *Front Immunol* (2013) 4:5. doi:10.3389/fimmu.2013.00005
10. Matsumoto M, Tanaka T, Kaisho T, Sanjo H, Copeland NG, Gilbert DJ, et al. A novel LPS-inducible C-type lectin is a transcriptional target of NF-IL6 in macrophages. *J Immunol* (1999) 163:5039–48.
11. Schoenen H, Huber A, Sonda N, Zimmermann S, Jantsch J, Lepenies B, et al. Differential control of Mincle-dependent cord factor recognition and macrophage responses by the transcription factors C/EBPbeta and HIF1alpha. *J Immunol* (2014) 193:3664–75. doi:10.4049/jimmunol.1301593
12. Kerscher B, Dambuza IM, Christofi M, Reid DM, Yamasaki S, Willment JA, et al. Signalling through MyD88 drives surface expression of the mycobacterial receptors MCL (Clec5f8, Clec4d) and Mincle (Clec4e) following microbial stimulation. *Microbes Infect* (2016) 18:505–9. doi:10.1016/j.micinf.2016.03.007
13. Flornes LM, Bryceson YT, Spurkland A, Lorentzen JC, Dissen E, Fossum S. Identification of lectin-like receptors expressed by antigen presenting cells and neutrophils and their mapping to a novel gene complex. *Immunogenetics* (2004) 56:506–17. doi:10.1007/s00251-004-0714-x
14. Behler F, Maus R, Bohling J, Knippenberg S, Kirchhof G, Nagata M, et al. Macrophage-inducible C-type lectin Mincle-expressing dendritic cells contribute to control of splenic *Mycobacterium bovis* BCG infection in mice. *Infect Immun* (2015) 83:184–96. doi:10.1128/IAI.02500-14
15. Lee WB, Kang JS, Yan JJ, Lee MS, Jeon BY, Cho SN, et al. Neutrophils promote mycobacterial trehalose dimycolate-induced lung inflammation via the Mincle pathway. *PLoS Pathog* (2012) 8:e1002614. doi:10.1371/journal.ppat.1002614
16. Kawata K, Illarionov P, Yang GX, Kenny TP, Zhang W, Tsuda M, et al. Mincle and human B cell function. *J Autoimmun* (2012) 39:315–22. doi:10.1016/j.jaut.2012.04.004
17. Behler F, Steinwede K, Balboa L, Ueberberg B, Maus R, Kirchhof G, et al. Role of Mincle in alveolar macrophage-dependent innate immunity against mycobacterial infections in mice. *J Immunol* (2012) 189:3121–9. doi:10.4049/jimmunol.1201399
18. Patin EC, Willcocks S, Orr S, Ward TH, Lang R, Schaible UE. Mincle-mediated anti-inflammatory IL-10 response counter-regulates IL-12 in vitro. *Innate Immun* (2016) 22(3):181–5. doi:10.1177/1753425916636671
19. Iborra S, Martinez-Lopez M, Cueto FJ, Conde-Garrosa R, Del Fresno C, Izquierdo HM, et al. *Leishmania* uses Mincle to target an inhibitory ITAM signaling pathway in dendritic cells that dampens adaptive immunity to infection. *Immunity* (2016) 45:788–801. doi:10.1016/j.immuni.2016.09.012
20. Zhang Z, He L, Hu S, Wang Y, Lai Q, Yang P, et al. AAL exacerbates pro-inflammatory response in macrophages by regulating Mincle/Syk/Card9 signaling along with the Nlrp3 inflammasome assembly. *Am J Transl Res* (2015) 7:1812–25.
21. Miyake Y, Toyonaga K, Mori D, Kakuta S, Hoshino Y, Oyamada A, et al. C-type lectin MCL is an FcRgamma-coupled receptor that mediates the adjuvanticity of mycobacterial cord factor. *Immunity* (2013) 38:1050–62. doi:10.1016/j.immuni.2013.03.010
22. Miyake Y, Masatsugu OH, Yamasaki S. C-type lectin receptor MCL facilitates Mincle expression and signaling through complex formation. *J Immunol* (2015) 194:5366–74. doi:10.4049/jimmunol.1402429
23. Lobato-Pascual A, Saether PC, Fossum S, Dissen E, Daws MR. Mincle, the receptor for mycobacterial cord factor, forms a functional receptor complex with MCL and FcepsilonRI-gamma. *Eur J Immunol* (2013) 43:3167–74. doi:10.1002/eji.201343752
24. Ostrop J, Jozefowski K, Zimmermann S, Hofmann K, Strasser E, Lepenies B, et al. Contribution of MINCLE-SYK signaling to activation of primary human APCs by mycobacterial cord factor and the novel adjuvant TDB. *J Immunol* (2015) 195:2417–28. doi:10.4049/jimmunol.1500102
25. Hupfer T, Schick J, Jozefowski K, Voehringer D, Ostrop J, Lang R. Stat6-dependent inhibition of Mincle expression in mouse and human antigen-presenting cells by the Th2 cytokine IL-4. *Front Immunol* (2016) 7:423. doi:10.3389/fimmu.2016.00423
26. Ebid R, Lichtnekert J, Anders HJ. Hyaluronan is not a ligand but a regulator of toll-like receptor signaling in mesangial cells: role of extracellular matrix in innate immunity. *ISRN Nephrol* (2014) 2014:714081. doi:10.1155/2014/714081
27. Zahringer U, Lindner B, Inamura S, Heine H, Alexander C. TLR2 – promiscuous or specific? A critical re-evaluation of a receptor expressing apparent broad specificity. *Immunobiology* (2008) 213:205–24. doi:10.1016/j.imbio.2008.02.005
28. Seifert L, Werba G, Tiwari S, Gao Ly NN, Allothman S, Alqunaibit D, et al. The necrosome promotes pancreatic oncogenesis via CXCL1 and Mincle-induced immune suppression. *Nature* (2016) 532:245–9. doi:10.1038/nature17403
29. Suzuki Y, Nakano Y, Mishiro K, Takagi T, Tsuruma K, Nakamura M, et al. Involvement of Mincle and Syk in the changes to innate immunity after ischemic stroke. *Sci Rep* (2013) 3:3177. doi:10.1038/srep03177
30. Zhou H, Yu M, Zhao J, Martin BN, Roychowdhury S, McMullen MR, et al. IRAKM-Mincle axis links cell death to inflammation: pathophysiological implications for chronic alcoholic liver disease. *Hepatology* (2016) 64:1978–93. doi:10.1002/hep.28811
31. Nagata M, Izumi Y, Ishikawa E, Kiyotake R, Doi R, Iwai S, et al. Intracellular metabolite beta-glucosylceramide is an endogenous Mincle ligand possessing immunostimulatory activity. *Proc Natl Acad Sci U S A* (2017) 114(16):E3285–94. doi:10.1073/pnas.1618133114
32. Kiyotake R, Oh-Hora M, Ishikawa E, Miyamoto T, Ishibashi T, Yamasaki S. Human Mincle binds to cholesterol crystals and triggers innate immune responses. *J Biol Chem* (2015) 290:25322–32. doi:10.1074/jbc.M115.645234
33. Kostarnoy AV, Gancheva PG, Lepenies B, Tukhvatulin AI, Dzharullaeva AS, Polyakov NB, et al. Receptor Mincle promotes skin allergies and is capable of recognizing cholesterol sulfate. *Proc Natl Acad Sci U S A* (2017) 114: E2758–65. doi:10.1073/pnas.1611665114
34. Heitmann L, Schoenen H, Ehlers S, Lang R, Holscher C. Mincle is not essential for controlling *Mycobacterium tuberculosis* infection. *Immunobiology* (2013) 218:506–16. doi:10.1016/j.imbio.2012.06.005
35. Wilson GJ, Marakalala MJ, Hoving JC, van Laarhoven A, Drummond RA, Kerscher B, et al. The C-type lectin receptor CLECSF8/CLEC4D is a key component of anti-mycobacterial immunity. *Cell Host Microbe* (2015) 17:252–9. doi:10.1016/j.chom.2015.01.004
36. Wells CA, Salvage-Jones JA, Li X, Hitchens K, Butcher S, Murray RZ, et al. The macrophage-inducible C-type lectin, Mincle, is an essential component of the innate immune response to *Candida albicans*. *J Immunol* (2008) 180:7404–13. doi:10.4049/jimmunol.180.11.7404
37. Yamasaki S, Matsumoto M, Takeuchi O, Matsuzawa T, Ishikawa E, Sakuma M, et al. C-type lectin Mincle is an activating receptor for pathogenic fungus *Malassezia*. *Proc Natl Acad Sci U S A* (2009) 106:1897–902. doi:10.1073/pnas.0805177106
38. Ishikawa T, Itoh F, Yoshida S, Saijo S, Matsuzawa T, Gono T, et al. Identification of distinct ligands for the C-type lectin receptors Mincle and Dectin-2 in the pathogenic fungus *Malassezia*. *Cell Host Microbe* (2013) 13:477–88. doi:10.1016/j.chom.2013.03.008
39. Rabes A, Zimmermann S, Reppe K, Lang R, Seeberger PH, Suttorp N, et al. The C-type lectin receptor Mincle binds to *Streptococcus pneumoniae* but plays a limited role in the anti-pneumococcal innate immune response. *PLoS One* (2015) 10:e017022. doi:10.1371/journal.pone.0117022
40. Devi S, Rajakumara E, Ahmed N. Induction of Mincle by *Helicobacter pylori* and consequent anti-inflammatory signaling denote a bacterial survival strategy. *Sci Rep* (2015) 5:15049. doi:10.1038/srep15049
41. Wevers BA, Kaptein TM, Zijlstra-Willems EM, Theelen B, Boekhout T, Geijtenbeek TB, et al. Fungal engagement of the C-type lectin Mincle suppresses dectin-1-induced antifungal immunity. *Cell Host Microbe* (2014) 15:494–505. doi:10.1016/j.chom.2014.03.008
42. Kottom TJ, Hebrink DM, Jenson PE, Nandakumar V, Wuthrich M, Wang H, et al. The interaction of pneumocystis with the C-type lectin receptor Mincle exerts a significant role in host defense against infection. *J Immunol* (2017) 198(9):3515–25. doi:10.4049/jimmunol.1600744
43. Schick J, Etschel P, Bailo R, Ott L, Bhatt A, Lepenies B, et al. Toll-like receptor 2 and Mincle cooperatively sense corynebacterial cell wall glycolipids. *Infect Immun* (2017) 85(7):e00075–17. doi:10.1128/IAI.00075-17
44. Behler-Janbeck F, Takano T, Maus R, Stolper J, Jonigk D, Tort Tarres M, et al. C-type lectin Mincle recognizes glucosyl-diacylglycerol of *Streptococcus pneumoniae* and plays a protective role in pneumococcal pneumonia. *PLoS Pathog* (2016) 12:e1006038. doi:10.1371/journal.ppat.1006038
45. Lee WB, Kang JS, Choi WY, Zhang Q, Kim CH, Choi UY, et al. Mincle-mediated translational regulation is required for strong nitric oxide production and inflammation resolution. *Nat Commun* (2016) 7:11322. doi:10.1038/ncomms11322

46. Shah S, Nagata M, Yamasaki S, Williams SJ. Total synthesis of a cyclopropane-fatty acid alpha-glucosyl diglyceride from *Lactobacillus plantarum* and identification of its ability to signal through Mincle. *Chem Commun* (2016) 52:10902–5. doi:10.1039/C6CC05631H
47. Ishikawa E, Ishikawa T, Morita YS, Toyonaga K, Yamada H, Takeuchi O, et al. Direct recognition of the mycobacterial glycolipid, trehalose dimycolate, by C-type lectin Mincle. *J Exp Med* (2009) 206:2879–88. doi:10.1084/jem.20091750
48. Schoenen H, Bodendorfer B, Hitchens K, Manzanero S, Werninghaus K, Nimmerjahn F, et al. Cutting edge: Mincle is essential for recognition and adjuvant activity of the mycobacterial cord factor and its synthetic analog trehalose-dibehenate. *J Immunol* (2010) 184:2756–60. doi:10.4049/jimmunol.0904013
49. Hattori Y, Morita D, Fujiwara N, Mori D, Nakamura T, Harashima H, et al. Glycerol monomycolate is a novel ligand for the human, but not mouse macrophage inducible C-type lectin, Mincle. *J Biol Chem* (2014) 289:15405–12. doi:10.1074/jbc.M114.566489
50. Richardson MB, Torigoe S, Yamasaki S, Williams SJ. *Mycobacterium tuberculosis* beta-gentiobiosyl diacylglycerides signal through the pattern recognition receptor Mincle: total synthesis and structure activity relationships. *Chem Commun* (2015) 51:15027–30. doi:10.1039/C5CC04773K
51. Huber A, Kallerup RS, Korsholm KS, Franzky H, Lepenies B, Christensen D, et al. Trehalose diester glycolipids are superior to the monoesters in binding to Mincle, activation of macrophages in vitro and adjuvant activity in vivo. *Innate Immun* (2016) 22:405–18. doi:10.1177/1753425916651132
52. Lang R, Schoenen H, Desel C. Targeting Syk-Card9-activating C-type lectin receptors by vaccine adjuvants: findings, implications and open questions. *Immunobiology* (2011) 216:1184–91. doi:10.1016/j.imbio.2011.06.005
53. Levitz SM, Golenbock DT. Beyond empiricism: informing vaccine development through innate immunity research. *Cell* (2012) 148:1284–92. doi:10.1016/j.cell.2012.02.012
54. Smith DG, Williams SJ. Immune sensing of microbial glycolipids and related conjugates by T cells and the pattern recognition receptors MCL and Mincle. *Carbohydr Res* (2016) 420:32–45. doi:10.1016/j.carres.2015.11.009
55. Drickamer K, Taylor ME. Recent insights into structures and functions of C-type lectins in the immune system. *Curr Opin Struct Biol* (2015) 34:26–34. doi:10.1016/j.sbi.2015.06.003
56. Feinberg H, Jegouzo SA, Rowntree TJ, Guan Y, Brash MA, Taylor ME, et al. Mechanism for recognition of an unusual mycobacterial glycolipid by the macrophage receptor Mincle. *J Biol Chem* (2013) 288:28457–65. doi:10.1074/jbc.M113.497149
57. Jegouzo SA, Harding EC, Acton O, Rex MJ, Fadden AJ, Taylor ME, et al. Defining the conformation of human Mincle that interacts with mycobacterial trehalose dimycolate. *Glycobiology* (2014) 24:1291–300. doi:10.1093/glycob/cwu072
58. Feinberg H, Rambaruth ND, Jegouzo SA, Jacobsen KM, Djurhuus R, Poulsen TB, et al. Binding sites for acylated trehalose analogs of glycolipid ligands on an extended carbohydrate recognition domain of the macrophage receptor Mincle. *J Biol Chem* (2016) 291:21222–33. doi:10.1074/jbc.M116.749515
59. Furukawa A, Kamishikiryo J, Mori D, Toyonaga K, Okabe Y, Toji A, et al. Structural analysis for glycolipid recognition by the C-type lectins Mincle and MCL. *Proc Natl Acad Sci U S A* (2013) 110:17438–43. doi:10.1073/pnas.1312649110
60. Tima HG, Al Dulayymi JR, Denis O, Lehebel P, Baols KS, Mohammed MO, et al. Inflammatory properties and adjuvant potential of synthetic glycolipids homologous to mycolate esters of the cell wall of *Mycobacterium tuberculosis*. *J Innate Immun* (2017) 9:162–80. doi:10.1159/000450955
61. Decout A, Silva-Gomes S, Drocourt D, Barbe S, Andre I, Cueto FJ, et al. Rational design of adjuvants targeting the C-type lectin Mincle. *Proc Natl Acad Sci U S A* (2017) 114:2675–80. doi:10.1073/pnas.1612421114
62. Ostrop J, Lang R. Contact, collaboration, and conflict: signal integration of Syk-coupled C-type lectin receptors. *J Immunol* (2017) 198:1403–14. doi:10.4049/jimmunol.1601665
63. Turner J, Gonzalez-Juarrero M, Ellis DL, Basaraba RJ, Kipnis A, Orme IM, et al. In vivo IL-10 production reactivates chronic pulmonary tuberculosis in C57BL/6 mice. *J Immunol* (2002) 169:6343–51. doi:10.4049/jimmunol.169.11.6343
64. Feng CG, Kullberg MC, Jankovic D, Cheever AW, Caspar P, Coffman RL, et al. Transgenic mice expressing human interleukin-10 in the antigen-presenting cell compartment show increased susceptibility to infection with *Mycobacterium avium* associated with decreased macrophage effector function and apoptosis. *Infect Immun* (2002) 70:6672–9. doi:10.1128/IAI.70.12.6672-6679.2002
65. Lang R, Rutschman RL, Greaves DR, Murray PJ. Autocrine deactivation of macrophages in transgenic mice constitutively overexpressing IL-10 under control of the human CD68 promoter. *J Immunol* (2002) 168:3402–11. doi:10.4049/jimmunol.168.7.3402
66. Beamer GL, Flaherty DK, Assogba BD, Stromberg P, Gonzalez-Juarrero M, de Waal Malefyt R, et al. Interleukin-10 promotes *Mycobacterium tuberculosis* disease progression in CBA/J mice. *J Immunol* (2008) 181:5545–50. doi:10.4049/jimmunol.181.8.5545
67. Haraguchi N, Kikuchi N, Morishima Y, Matsuyama M, Sakurai H, Shibuya A, et al. Activation of murine invariant NKT cells promotes susceptibility to candidiasis by IL-10 induced modulation of phagocyte antifungal activity. *Eur J Immunol* (2016) 46:1691–703. doi:10.1002/eji.201545987
68. Pitt JM, Stavropoulos E, Redford PS, Beebe AM, Bancroft GJ, Young DB, et al. Blockade of IL-10 signaling during bacillus Calmette-Guerin vaccination enhances and sustains Th1, Th17, and innate lymphoid IFN-gamma and IL-17 responses and increases protection to *Mycobacterium tuberculosis* infection. *J Immunol* (2012) 189:4079–87. doi:10.4049/jimmunol.1201061
69. Zhang X, Majlessi L, Deriaud E, Leclerc C, Lo-Man R. Coactivation of Syk kinase and MyD88 adaptor protein pathways by bacteria promotes regulatory properties of neutrophils. *Immunity* (2009) 31:761–71. doi:10.1016/j.immuni.2009.09.016
70. Strasser D, Neumann K, Bergmann H, Marakalala MJ, Guler R, Rojowska A, et al. Syk kinase-coupled C-type lectin receptors engage protein kinase C-sigma to elicit Card9 adaptor-mediated innate immunity. *Immunity* (2012) 36:32–42. doi:10.1016/j.immuni.2011.11.015
71. Patin EC, Jones AV, Thompson A, Clement M, Liao CT, Griffiths JS, et al. IL-27 induced by select *Candida* spp. via TLR7/NOD2 signaling and IFN-beta production inhibits fungal clearance. *J Immunol* (2016) 197:208–21. doi:10.4049/jimmunol.1501204
72. Rivera A. When PRRs collide: Mincle meddles with dectin and toll. *Cell Host Microbe* (2014) 15:397–9. doi:10.1016/j.chom.2014.03.013
73. Wuthrich M, Wang H, Li M, Lerkuthirat T, Hardison SE, Brown GD, et al. *Fonsecaea pedrosoi*-induced Th17-cell differentiation in mice is fostered by dectin-2 and suppressed by Mincle recognition. *Eur J Immunol* (2015) 45:2542–52. doi:10.1002/eji.201545591
74. Greco SH, Mahmood SK, Vahle AK, Ochi A, Batel J, Deutsch M, et al. Mincle suppresses toll-like receptor 4 activation. *J Leukoc Biol* (2016) 100:185–94. doi:10.1189/jlb.3A0515-185R
75. Weiss G, Schaible UE. Macrophage defense mechanisms against intracellular bacteria. *Immunol Rev* (2015) 264:182–203. doi:10.1111/imr.12266
76. Kerrigan AM, Brown GD. C-type lectins and phagocytosis. *Immunobiology* (2009) 214:562–75. doi:10.1016/j.imbio.2008.11.003
77. Dambuzza IM, Brown GD. C-type lectins in immunity: recent developments. *Curr Opin Immunol* (2015) 32:21–7. doi:10.1016/j.coi.2014.12.002
78. Sharma A, Steichen AL, Jondle CN, Mishra BB, Sharma J. Protective role of Mincle in bacterial pneumonia by regulation of neutrophil mediated phagocytosis and extracellular trap formation. *J Infect Dis* (2014) 209:1837–46. doi:10.1093/infdis/jit820
79. Kang PB, Azad AK, Torrelles JB, Kaufman TM, Beharka A, Tibesar E, et al. The human macrophage mannose receptor directs *Mycobacterium tuberculosis* lipoarabinomannan-mediated phagosomal biogenesis. *J Exp Med* (2005) 202:987–99. doi:10.1084/jem.20051239
80. Torrelles JB, Azad AK, Schlesinger LS. Fine discrimination in the recognition of individual species of phosphatidyl-myo-inositol mannosides from *Mycobacterium tuberculosis* by C-type lectin pattern recognition receptors. *J Immunol* (2006) 177:1805–16. doi:10.4049/jimmunol.177.3.1805
81. Sweet L, Singh PP, Azad AK, Rajaram MV, Schlesinger LS, Schorey JS. Mannose receptor-dependent delay in phagosome maturation by *Mycobacterium avium* glycopeptidolipids. *Infect Immun* (2010) 78:518–26. doi:10.1128/IAI.00257-09
82. Indrigo J, Hunter RL Jr, Actor JK. Cord factor trehalose 6,6'-dimycolate (TDM) mediates trafficking events during mycobacterial infection of

- murine macrophages. *Microbiology* (2003) 149:2049–59. doi:10.1099/mic.0.26226-0
83. Axelrod S, Oschkinat H, Enders J, Schlegel B, Brinkmann V, Kaufmann SH, et al. Delay of phagosome maturation by a mycobacterial lipid is reversed by nitric oxide. *Cell Microbiol* (2008) 10:1530–45. doi:10.1111/j.1462-5822.2008.01147.x
 84. Kodar K, Eising S, Khan AA, Steiger S, Harper JL, Timmer MS, et al. The uptake of trehalose glycolipids by macrophages is independent of Mincle. *Chembiochem* (2015) 16:683–93. doi:10.1002/cbic.201402506
 85. Patin EC, Geffken AC, Willcocks S, Leschczyk C, Haas A, Nimmerjahn F, et al. Trehalose dimycolate interferes with FcγR-mediated phagosome maturation through Mincle, SHP-1 and FcγRIIB signalling. *PLoS One* (2017) 12:e0174973. doi:10.1371/journal.pone.0174973
 86. Karsten CM, Pandey MK, Figge J, Kilchenstein R, Taylor PR, Rosas M, et al. Anti-inflammatory activity of IgG1 mediated by Fc galactosylation and association of FcγRIIB and dectin-1. *Nat Med* (2012) 18:1401–6. doi:10.1038/nm.2862
 87. Maglione PJ, Xu J, Casadevall A, Chan J. Fc γ receptors regulate immune activation and susceptibility during *Mycobacterium tuberculosis* infection. *J Immunol* (2008) 180:3329–38. doi:10.4049/jimmunol.180.5.3329
 88. Perley CC, Frahm M, Click EM, Dobos KM, Ferrari G, Stout JE, et al. The human antibody response to the surface of *Mycobacterium tuberculosis*. *PLoS One* (2014) 9:e98938. doi:10.1371/journal.pone.0098938
 89. Korsholm KS, Hansen J, Karlens K, Filskov J, Mikkelsen M, Lindenstrom T, et al. Induction of CD8+ T-cell responses against subunit antigens by the novel cationic liposomal CAF09 adjuvant. *Vaccine* (2014) 32:3927–35. doi:10.1016/j.vaccine.2014.05.050
 90. Roperto S, Russo V, Esposito I, Ceccarelli DM, Paciello O, Avallone L, et al. Mincle, an innate immune receptor, is expressed in urothelial cancer cells of papillomavirus-associated urothelial tumors of cattle. *PLoS One* (2015) 10:e0141624. doi:10.1371/journal.pone.0141624
 91. Igarashi Y, Mogi T, Yanase S, Miyanaga S, Fujita T, Sakurai H, et al. Brartemicin, an inhibitor of tumor cell invasion from the actinomycete *Nonomuraea* sp. *J Nat Prod* (2009) 72:980–2. doi:10.1021/np9000575
 92. Jacobsen KM, Keiding UB, Clement LL, Schaffert ES, Rambaruth ND, Johannsen M, et al. The natural product brartemicin is a high affinity ligand for the carbohydrate-recognition domain of the macrophage receptor Mincle. *Medchemcomm* (2015) 6:647–52. doi:10.1039/c4md00512k
 93. Zhao C, Sun H, Tong X, Qi Y. An antitumour lectin from the edible mushroom *Agrocybe aegerita*. *Biochem J* (2003) 374:321–7. doi:10.1042/bj20030300
 94. Greco SH, Torres-Hernandez A, Kalabin A, Whiteman C, Rokosh R, Ravirala S, et al. Mincle signaling promotes con A hepatitis. *J Immunol* (2016) 197:2816–27. doi:10.4049/jimmunol.1600598
 95. Lee EJ, Brown BR, Vance EE, Snow PE, Silver PB, Heinrichs D, et al. Mincle activation and the Syk/Card9 signaling axis are central to the development of autoimmune disease of the eye. *J Immunol* (2016) 196:3148–58. doi:10.4049/jimmunol.1502355

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2017 Patin, Orr and Schaible. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.