Genome wide interactions of wild-type and activator bypass forms of σ54

Genome wide interactions of wild-type and activator bypass forms of σ54

Jorr Schaefer¹, Christoph Engl¹,²⁎, Nan Zhang¹, Edward Lawton¹ and Martin Buck¹⁎

¹Faculty of Natural Sciences, Division of Cell & Molecular Biology, Imperial College London, London SW7 2AZ, UK and ²Institute for Global Food Security, Queen’s University Belfast, Belfast BT9 5BN, UK

Received April 8, 2015; Revised May 7, 2015; Accepted May 25, 2015

ABSTRACT

Enhancer-dependent transcription involving the promoter specificity factor σ54 is widely distributed amongst bacteria and commonly associated with cell envelope function. For transcription initiation, σ54 RNA polymerase yields open promoter complexes through its remodelling by cognate AAA+ ATPase activators. Since activators can be bypassed in vitro, bypass transcription in vivo could be a source of emergent gene expression along evolutionary pathways yielding new control networks and transcription patterns. At a single test promoter in vivo bypass transcription was not observed. We now use genome-wide transcription profiling, genome-wide mutagenesis and gene over-expression strategies in Escherichia coli, to (i) scope the range of bypass transcription in vivo and (ii) identify genes which might alter bypass transcription in vivo. We find little evidence for pervasive bypass transcription in vivo with only a small subset of σ54 promoters functioning without activators. Results also suggest no one gene limits bypass transcription in vivo, arguing bypass transcription is strongly kept in check. Promoter sequences subject to repression by σ54 were evident, indicating loss of rpoN (encoding σ54) rather than creating rpoN bypass alleles would be one evolutionary route for new gene expression patterns. Finally, cold-shock promoters showed unusual σ54-dependence in vivo not readily correlated with conventional σ54 binding-sites.

INTRODUCTION

Amongst the strategies used to regulate transcription initiation in bacteria, the emergence of the enhancer-dependent system that uses the σ54 RNA polymerase is of special interest because it deviates from the transcription factor recruitment mechanisms commonly seen (1,2). Rather, binding and hydrolysis of aenosine triphosphate (ATP) by cognate AAA+ ATPase activator proteins remodel σ54 containing closed promoter complexes to yield open promoter complexes (3,4). Potentially, σ54 initially acted as a global repressor of RNA polymerase (RNAP) activity since it effectively road blocks the DNA entry path used for delivering melted DNA into the DNA binding cleft of RNAP. This silenced state of RNAP may then have become a target for regulation. Intriguingly, to date no bacteria have been found where the gene encoding σ54 (rpoN) is present in the absence of any cognate AAA+ activator (4,5). This suggests forms of σ54 functioning without activators are uncommon. Yet variants of σ54 bypassing activator requirements for making open promoter complexes exist and have been characterized extensively in vitro (6,7). Single amino acid substitutions in σ54 can yield activator bypass forms. Quite why σ54 has seemingly not (or rarely) evolved to activator independence is rather unclear. Here we have investigated the in vivo global actions of wild-type σ54 in Escherichia coli bacteria and one bypass variant to explore potential restrictions on evolving bypass gene regulation systems.

We found that only a very limited number of knock-outs (KOs) of non-essential genes in E. coli increased activator bypass transcription suggesting multifactorial global constraints prevent unregulated transcription by the σ54 containing RNAP holoenzyme (σ54RNAP). Intriguingly, some genes with σ54 binding sites were found to be repressed in a σ54-dependent manner. In some cases the putative σ54 binding site overlapped that of a putative σ70 binding site suggesting a competition for promoter sites, which we confirmed by in vitro experiments. In other cases, σ54 and σ70 RNAP appear to act antagonistically through e.g. convergent transcription events.

MATERIALS AND METHODS

Bacterial strains and growth conditions

For the construction of the gnaAΔ52 strains (Supplementary Table S1) Plvir transduction was used to insert rpoN208::Tn10 (K1471 donor strain) into the native chromosomal rpoN locus of BW25113 (recipient). Subsequently,
transformation was used to transfer the pSB4A3 plasmid harbouring \( \Phi P_{\text{dhAp2-gfp}} \) (a gift from Baojun Wang, University of Edinburgh, UK) and pBAD18 cm (containing *Klebsiella pneumonia* rpoN, rpoN\_ARI or vector only). For the construction of the pspA strains (Supplementary Table S1) P1\_vir transduction was used to insert rpoN208::Tn10 (K1471 donor strain) into the native chromosomal rpoN locus of BW25113 (recipient). Next the \( \Phi P_{\text{pspA-lacZ}} \) (MVA4 donor strain; 8) was inserted into the att site of the BW25113 rpoN208::Tn10 chromosome, using P1\_vir transduction. Finally, transformation was used to transfer pBAD18 cm (containing *Klebsiella pneumonia* rpoN, rpoN\_ARI or vector only) into the BW25113 rpoN208::Tn10 \( \Phi P_{\text{pspA-lacZ}} \). The F3K3 plasmid containing the filamentous phage pIV secretin was also transformed into this strain. To transfer the single-gene KOs from the KEIO collection (9) and small cretin was also transformed into this strain. To transfer the SABRS recipient strain (Supplementary Table S1).

For gene expression assays *in vivo*, strains were grown on LB indicator (XGal and MacConkey) plates and in minimal Gutnick liquid media supplemented with L-glutamine (11) as the nitrogen source to allow good growth in the absence of \( \sigma^{54} \) function (since the nitrogen assimilation pathways in *E. coli* rely on \( \sigma^{54} \) for expressing appropriate levels of glutamine synthetase). The antibiotic concentrations used were 30 \( \mu \)g/ml kanamycin, 50 \( \mu \)g/ml chloramphenicol and 50–100 \( \mu \)g/ml ampicillin.

**Gene expression assays *in vivo***

To measure activity of \( \Phi P_{\text{pspA-lacZ}} \) *in vivo*, \( \beta \)-Galactosidase assays were carried out as described (12). To determine gene expression levels of the \( \Phi P_{\text{dhAp2-gfp}} \) reporter *in vivo*, cells were grown in black 96-well clear-bottom tissue culture plates. Optical density at 600 nm (OD\(_{600}\)) and green fluorescence (excitation: 485 nm; emission: 520 ± 10 nm, gain: 1000) were measured simultaneously using a BMG FLUOstar Omega microplate reader. Promoter activity was expressed as fluorescence emission at 520 nm per OD\(_{600}\) measured in triplicate.

**Inverse PCR**

Inverse polymerase chain reaction (PCR) was used to identify selected KO mutants (Supplementary Table S2). Chromosomal DNA from strains that were positive for \( \beta \)-galactosidase activity on indicator plates and in liquid culture (Miller assay) was purified and digested with NcoI, a 6-base cutter. Re-ligation of the resulting fragments into circular DNA was done in a volume of 100 \( \mu \)l. This DNA was used as a template for PCR, using kanamycin cassette-specific forward and reverse primers (9). Products were run on a gel, extracted and sequenced. NCBI Blast (http://blast.ncbi.nlm.nih.gov/Blast.cgi) was used to align sequenced regions to the *E. coli* chromosome and to determine the identity of the deleted gene.

**RNA sequencing (RNAseq)**

A total of 100 ml cell cultures were grown to exponential phase at 42 or 37°C, in minimal Gutnick media supplemented with 5 mM L-Glutamine and 0.4% glucose. Cell pellets were sent to Vertis Biotechnologie AG (Germany) for further processing. Total RNA was isolated from the cell pellets using a bead mill and the mirVana RNA isolation kit (Ambion) including DNase treatment. Ribosomal RNA molecules were depleted from the total RNA preparations using the RiboZero rRNA Removal Kit (Bacteria) (Epi-centre). The rRNA depleted RNAs were fragmented with RNase III. Then, the RNA fragments were poly(A)-tailed using poly(A) polymerase and the \( 5' \) PPP structures were removed using RNA 5' polyphosphatase (Epicentre). An RNA adapter was ligated to the 5' -phosphate of the RNA fragments. First-strand cDNA synthesis was performed using an oligo(dT)-adapter primer and M-MLV reverse transcriptase. The resulting cDNA was PCR-amplified using a high fidelity DNA polymerase. The cDNA was purified using the Agencourt AMPure XP kit (Beckman Coulter Genomics). The cDNA pools were size fractionated in the size range of 200–450 bp using a differential clean-up with the Agencourt AMPure kit. The primers used for PCR amplification were designed for TruSeq sequencing according to the instructions of Illumina and sequenced on an Illumina HiSeq 2000 system using 50 bp read length. Near equimolar amounts of cDNA were used for sequencing. The cDNA reads were analysed via the RNA-seq workflow within Partek® Genomics suite 6.6, including a QA/QC step to gauge the sequencing quality. Each sample yielded close to equivalent total reads with 100% of reads aligned to the *E. coli* K-12 reference genome (NC_000913) (Supplementary Table S4). For each gene, reads were normalized for both the number of reads and length of RNA and expressed as RPKM (reads per kilobase per million) (13; Supplementary Table S4). A one-way ANOVA statistical analysis was performed to test for significant differences in genome-wide gene expression between all 3 strains (Supplementary Table S4).

**In vitro spRNA assays**

Small primed RNAs (spRNAs) were synthesized in 10 \( \mu \)l containers: 0.125 \( \mu \)M \( \sigma^{70} / \sigma^{54} \)RNAP holoenzyme, 20 \( \mu \)M linear promoters, 0.5 \( \mu \)M dinucleotide primers, 0.2 mg/ml heparin and 0.2 \( \mu \)Ci/\( \mu \)l \( ^{32} \)P-NTP in STA buffer (2.5 mM Tris-acetate pH8, 8 mM Mg-acetate, 10 mM KCl, 1 mM DTT, 3.5% (w/v) PEG 8000) at 37°C for 10 min. For the \( \sigma^{70} / \sigma^{54} \) competition assays, 0.4 \( \mu \)M \( \sigma^{54} \) were either added before or with \( \sigma^{70} \) for binding competition. The reactions were quenched by formamide stop dye and run on a 20% sequencing gel. Signals were detected by Fuji PhosphorImager and quantified using the AIDA Image Analyzer software (raytest).

**RESULTS**

Genome-wide screens for increased bypass transcription suggest constraints exist

A form of \( \sigma^{54} \) lacking the repressive Region I, \( \sigma^{54} \)ARI, can bypass activator requirement for transcription *in vitro* (Fig-
Figure 1. (A) The Region I of $\sigma^{54}$ acts to inhibit RNA polymerase activity by blocking promoter DNA entry into the major DNA binding cleft. (B) Removal of Region I enables in vitro transcription from the late-melted nifH promoter ($-10-1/WT$) without the AAA+ activator PspF$_{1-275}$.

Figure 2. The $\sigma^{54}$ dependent pspA promoter is active in vivo in the presence of stress inducing signals (here pIV secretin) and non-bypass wild-type $\sigma^{54}$, but is inactive with the $\Delta$RI $\sigma^{54}$ form. Experiments were performed with three biological replicates.

Using a lacZ fusion to the pspA promoter ($P_{pspA}$lacZ) in single copy integrated into the att site of the E. coli chromosome we sought bypass activity from the $\sigma^{54}$-dependent pspA promoter in vivo. Reporter strains lacked rpoN and were complemented with either the wild-type or the $\Delta$RI form of $\sigma^{54}$ expressed from pBAD18 cm (Supplementary Table S1, Figure S1). While rpoN and rpoN$_{\DeltaRI}$ were under control of the araBAD arabinose inducible promoter, leaky expression (without arabinose) yielded sufficient levels of $\sigma^{54}$ to enable pIV-dependent transcription from the pspA promoter (Supplementary Figure S2). Thus unless stated otherwise (Supplementary Table S2) the experiments were performed in the absence of arabinose. Under standard growth conditions, no clear bypass transcription was observable with the $\sigma^{54} \Delta$RI form (Figure 2). Secretin pIV-dependent activation of the pspA promoter (16) via its cognate activator PspF acting on the $\sigma^{54}$RNAP holoenzyme was clearly observed in vivo (Figure 2).

It was reported that increased temperature enhances $\sigma^{54}$ activator bypass transcription in vitro at the $ghn$Ap2 pro-
moter (6). We therefore cultured cells at 30, 37 and 42°C to favour detecting bypass transcription in vivo. Indeed, we observed a four-fold increase in $\sigma^{54}\Delta$RI-dependent $\operatorname{gln}Ap2$ promoter activity at 42°C compared to 30 or 37°C. This bypass activity accounts for almost 50% of wild-type $\sigma^{54}$-dependent transcription (Figure 4).

To seek genetic determinants potentially restricting in vivo bypass transcription from the $pspA$ promoter we introduced non-essential gene KO's into the reporter strains from the KEIO collection of *E. coli* gene KO's (9; a gift from the Carol Gross laboratory, UCSF, USA) and the small RNA/small peptide KO collection (10; a gift from Diarmuid Hughes; Uppsala University, Sweden; 17) into the *E. coli* reporter strain and screened for increased expression of $P_{pспA}$. None was observed, suggesting no single gene product limits bypass transcription in *E. coli* (Supplementary Table S2).

**RNAseq analysis of the $\sigma^{54}$ bypass allele $\Delta$RI activity reveals activator independent promoter sites**

We conducted RNAseq of cells containing the $\Delta$RI bypass form of $\sigma^{54}$ and compared outcomes to those from cells with wild-type $\sigma^{54}$ and to cells lacking $\sigma^{54}$. Cells were cultured at 42°C, which enhances bypass in vitro (6), to favour detecting bypass transcription in vivo. We confirmed using the $\operatorname{gln}Ap2$ promoter this was so in vivo (Figure 4). We chose minimal media supplemented with L-glutamine as the nitrogen source to allow good growth in the absence of $\sigma^{54}$ function (since the nitrogen assimilation pathways in *E. coli* rely on $\sigma^{54}$ for expressing appropriate levels of glutamine synthetase).

RNAseq profiles of cells with wild-type $\sigma^{54}$ compared to cells with $\sigma^{54}\Delta$RI or no $\sigma^{54}$ revealed no major global bypass transcription (Figure 5). By scoring for more RNAseq reads in presence of the bypass $\sigma^{54}\Delta$RI form compared to the no $\sigma^{54}$ we identified eight sites of low-level activator-independent transcription from sequences downstream of a canonical $\Delta12/-24\sigma^{54}$ promoter DNA recognition sites (Table 1). We conclude that a small subset of $\sigma^{54}$ binding sites can yield transcripts independently of the conventional AAA+ ATPase driven activation pathway, and that this bypassing is mediated by the loss of Region I of $\sigma^{54}$. The C residue in the GC $-12$ element is conserved in 96% of bacterial $\sigma^{54}$ promoters (18). Alignment (19,20) of the bypassing $\Delta12/-24\sigma^{54}$ motifs indicates that the C residue is not strictly conserved at these promoters (Table 1). This finding is in line with in vitro studies showing that the consensus C at $-12$ is important for keeping basal transcription in check (21).
Figure 4. Elevated temperature increases bypass transcription from the gltA p2 promoter in vivo (grey), by over 40% activity of the wild-type rpoN control at the same temperature (black). Experiments were performed with three biological replicates.

Table 1. (A) $\sigma^{54}$ promoter sites showing increased bypass transcription at 42°C in vivo when Region I of $\sigma^{54}$ is absent

<table>
<thead>
<tr>
<th>Gene</th>
<th>Fold change $(rpoN_{AR1} /$ vector)</th>
<th>Fold change $(rpoN /$ vector)</th>
<th>Promoter sequence</th>
</tr>
</thead>
<tbody>
<tr>
<td>gltK</td>
<td>1.3</td>
<td>74</td>
<td>CTGGCACACCGCTGGCA</td>
</tr>
<tr>
<td>hydN</td>
<td>1.3</td>
<td>1.5</td>
<td>CTGGCATGATTTGTGAA</td>
</tr>
<tr>
<td>prpB</td>
<td>2.5</td>
<td>4.0</td>
<td>GTGGCACACCCCTTGCT</td>
</tr>
<tr>
<td>pspG</td>
<td>1.3</td>
<td>102</td>
<td>TTGGCATGATTCCTGTGTA</td>
</tr>
<tr>
<td>norV</td>
<td>1.5</td>
<td>1.2</td>
<td>CTGGCACCGCAATCTGGCA</td>
</tr>
<tr>
<td>rutA</td>
<td>1.3</td>
<td>1.9</td>
<td>CTGGCATCCGCTGGTTCGA</td>
</tr>
<tr>
<td>yahE</td>
<td>1.7</td>
<td>1.0</td>
<td>CTGGAAAGCGATTGTGCT</td>
</tr>
<tr>
<td>yaiA</td>
<td>1.5</td>
<td>1.2</td>
<td>GTGGGCGATCGCTGGTCT</td>
</tr>
</tbody>
</table>

Consensus: YTGGCACGNNNNNTTGCW

(B) Bypassing promoters have a non-consensus −12 promoter sequence, displaying a deviation from the canonical GC motif.
Figure 5. Genome-wide transcriptional profiling by RNA-seq revealed no major global bypass transcription by RpoN\Delta1RI. Shown are MA scatterplots comparing cells with wild-type RpoN to cells with no RpoN (Top left) or RpoN\Delta1RI (Top right) as well as cells with RpoN\Delta1RI to cells with no RpoN (Bottom left). $M$ is the difference in number of reads for each gene between a pair of strains expressed as $\log_2(\text{RPKM}_{\text{strain 1}}) - \log_2(\text{RPKM}_{\text{strain 2}})$, i.e. values > 0 indicate higher expression in strain 1 while values < 0 indicate higher expression in strain 2. $A$ is the average number of reads of a gene between both strains expressed as $0.5[\log_2(\text{RPKM}_{\text{strain 1}}) + \log_2(\text{RPKM}_{\text{strain 2}})]$. Data points within the grey shaded area ($M = \log_2(\text{RPKM}) = +/−2$) depict genes less than equal to four-fold differently expressed between the strains. The scatterplot in the Bottom right corner compares the $M$-values of the Top left and Top right plot showing that the differences in gene expression between cells with wild-type RpoN and no RpoN are similar to the differences in gene expression between cells with wild-type RpoN and RpoN\Delta1RI (dotted line: $y = x$). The data is derived from single sample analysis.

the experiment. Table 2 lists these promoters and the genes under their control. The majority are promoters from several well characterized $\sigma^{54}$-dependent systems such as those controlled by the activators such as NtrC, PspF, PrpR and RtcR.

$\sigma^{54}$ is repressive at some promoters

We also compared reads from cells lacking $\sigma^{54}$ to cells with the wild-type $\sigma^{54}$ or its $\Delta$RI form. Intriguingly, in several cases reads were higher when $\sigma^{54}$ was absent, suggesting that the $\sigma^{54}$RNAP holoenzyme can be repressive for some genes, either directly or indirectly. We inspected such genes for $−12−24$ sequences and grouped them into four different classes based on the location of the $\sigma^{54}$ binding motifs relative to the active $−10−35$ promoter sequences (Figure 6 and Supplementary Table S3). In several cases (class I and II), we find that the $\sigma^{54}$ recognition site is overlapping with the likely footprint of a $\sigma^{70}$RNAP holoenzyme on these sites, suggesting a simple binding competition mechanism of repression by the $\sigma^{54}$RNAP. Using purified components, we tested four $\sigma^{70}$-regulated linear homoduplex promoters (argT, chaC, mdfA and patA) from class I and II for binding repression by $\sigma^{54}$ in vitro (Figure 7 and Supplementary Figure S3). The $\sigma^{70}$RNAP holoenzyme was allowed to generate small primed RNAs (spRNAs) using initiating dinucleotides ($−1/+1$ or $+1/+2$). The experiments were performed under two conditions. In the first, $\sigma^{54}$ and $\sigma^{70}$ were co-incubated with the RNA prior to addition of promoter DNA probes. Here $\sigma^{54}$ was in three-fold molar excess to $\sigma^{70}$ and RNAP. This competition from excess $\sigma^{54}$ reduced the amount of spRNA synthesised by 10–60% (Figure 7). In the second, $\sigma^{54}$ was pre-incubated with the promoter DNA probes prior to addition of the $\sigma^{70}$RNAP holoenzyme. Here, $\sigma^{70}$ was in more than three-fold molar excess to
Figure 6. σ^{54}-dependent repression was observed at multiple genes with σ^{54} binding sites identified through ChIP Seq (Supplementary Table S3). These genes were grouped in 4 classes based on the location of the σ^{54} motifs relative to the promoter sequences. Shown are examples of each class and consensus motifs of class I–III. Class IV consists of only one member, katE.
Figure 7. $\sigma^{54}$ represses transcription by $\sigma^{70}$ in vitro. Purified holoenzymes were incubated with linear homoduplexes from four promoters ($\text{argT}$, $\text{chaC}$, $\text{mdfA}$ and $\text{patA}$ WT/WT) and transcription was measured using dinucleotide primers ($-1/+1$ or $+1/+2$) and a radioactive rNTP corresponding to $+1$ and or $+2$ to allow initiation of spRNA synthesis at the $-10/−35$ promoter sequences.

Table 2. $\sigma^{54}$ promoter sites active at $42^\circ$C in vivo

<table>
<thead>
<tr>
<th>Gene</th>
<th>Fold change ($\sigma^{54}/\sigma^{ART}$)</th>
<th>Cognate AAA+ Activator</th>
<th>Consensus motif</th>
</tr>
</thead>
<tbody>
<tr>
<td>$\text{aslB}$</td>
<td>2</td>
<td>Unknown</td>
<td>TTGGTAGCGCAACTGGT</td>
</tr>
<tr>
<td>$\text{emrD}$</td>
<td>3</td>
<td>Unknown</td>
<td>CTGGCGTATATCTGGCT</td>
</tr>
<tr>
<td>$\text{glmY}$</td>
<td>4</td>
<td>GlrR</td>
<td>TTGGCACAGTTACTGCA</td>
</tr>
<tr>
<td>$\text{glmA}$</td>
<td>11</td>
<td>NtrC</td>
<td>TTGGCACAGATTTGCTC</td>
</tr>
<tr>
<td>$\text{glnH}$</td>
<td>8</td>
<td>NtrC</td>
<td>CTGGCACAGATTTTTTCA</td>
</tr>
<tr>
<td>$\text{glnK}$</td>
<td>6</td>
<td>NtrC</td>
<td>CTGGCACACCGCTGGCA</td>
</tr>
<tr>
<td>$\text{hyfA}$</td>
<td>4</td>
<td>HyfR</td>
<td>ATGGCATCTTTTAATGCA</td>
</tr>
<tr>
<td>$\text{ibpB}$</td>
<td>14</td>
<td>Unknown</td>
<td>AAGGCCGCCTGGGCAGCG</td>
</tr>
<tr>
<td>$\text{prpB}$</td>
<td>2</td>
<td>PrpR</td>
<td>GTGGCAACACCCCTTGGCT</td>
</tr>
<tr>
<td>$\text{pspA}$</td>
<td>201</td>
<td>PspF</td>
<td>TTGGCACGCAATTTTGA</td>
</tr>
<tr>
<td>$\text{pspG}$</td>
<td>79</td>
<td>PspG</td>
<td>TTGGCATGATTCTTTGTGA</td>
</tr>
<tr>
<td>$\text{rtcB}$</td>
<td>2</td>
<td>RtcR</td>
<td>CTGGCACAGACGTTGCA</td>
</tr>
<tr>
<td>$\text{zraP}$</td>
<td>3</td>
<td>ZraR</td>
<td>TTGGCACGGAAAGATGCA</td>
</tr>
</tbody>
</table>

Where known, the cognate AAA+ activator is listed.
Figure 8. (A) Expression of cold-shock genes \textit{in vivo} is upregulated in a $\sigma^{54}$-dependent manner at 42°C but not at 37°C. \textit{In vitro} transcription of \textit{cspA} (B) and \textit{cspH} (C) with the -10/1/WT heteroduplex promoters is $\sigma^{70}$ dependent and largely $\sigma^{54}$ independent.
σ^{54} and RNAP. Again, we observed a similar σ^{54}-dependent reduction in spRNA synthesis (Supplementary Figure S3). Since the binding affinity and dissociation constant of σ^{54} and σ^{70} for RNAP are very similar (22) and similar results were obtained in both conditions, we suggest that inhibition is observed due to a binding competition from σ^{54} containing RNAP within the −10/−35 promoter sites rather than a direct competition for RNAP by σ^{70} and σ^{54}.

In one other case the putative −12/−24 recognition site is far away from where the upregulated transcription start site is mapped to by RNAsq (Figure 6 class IV). Here, repression may occur not by simple competition for overlapping promoters, but for example through direct or indirect interferences between convergent transcribing RNA polymerases.

Interestingly, in contrast to bypass promoters (Table 1) the C −12 residue is present in all observed σ^{54} motifs correlating with local repression of transcription, consistent with the aforementioned repressive nature of GC at −12 (21).

**Cold-shock promoters have unusual σ^{54} dependencies**

Notably, some of the promoters dependent upon σ^{54} in vivo were driving the expression of cold-shock genes (Figure 8A). ChIP Seq identified a σ^{54} binding site in two of these genes (cspA and cspH) despite lacking any clear consensus −12/−24 sequences (Joe Wade, University of Albany, USA; personal communication). However, in vitro transcription from cspA (Figure 8B) and cspH (Figure 8C) promoters was found to be σ^{70}-dependent and largely unresponsive to σ^{54}. Thus, the basis of their σ^{54} dependence in *E. coli* remains unclear, however in *Bacillus subtilis* the σ^{54} homologue (σ^{L}) has been shown to be essential for mounting a cold shock response through the transcription of σ^{54}-dependent cold shock transcriptional regulators BkdR and YplP (23). Intriguingly, both cspA and cspH promoters contain putative heat shock sigma factor (σ^{32}) binding motifs (24), consistent with the transcriptional start site mapped via the RNAsq reads (Figure 8B and C). Moreover, we only observe this σ^{54}-dependent upregulation of cold shock genes at 42°C and not 37°C (Figure 8A). In all RNAsq experiments cells were cold harvested. The expression of cold shock promoters may therefore reflect the difference in the temperature shift from 42°C to ice and from 37°C to ice. We note however that σ^{32} activity has been suggested to be regulated by σ^{54} (25), raising the possibility that σ^{54} may be required for the expression of cold shock genes cspA and cspH by modulating the activity of σ^{32}.

**DISCUSSION**

The σ^{54}-dependent gene expression paradigm is widely distributed amongst bacteria and is responsible for the expression of stress-induced genes important in many bacterial driven processes, especially the nitrogen cycle (26). Bioinformatics suggests the σ^{54} system is commonly associated with cell envelope function (27). Recent structural studies highlight how σ^{54} interacts with the core RNA polymerase to yield a holoenzyme unable to manipulate promoter DNA for making open promoter complexes until re-organised by its cognate AAA+ activator proteins (28). Studied here, the loss of the σ^{54} Region I from the holoenzyme frees up RNAP for promoter DNA delivery within and the subsequent templated synthesis of RNA (6,29–33). Our analysis of the properties of the bypass form of σ^{54} in vivo suggests such DNA delivery into the RNAP once Region I is missing is rarely occurring in vivo. *In vitro* the activator bypass phenotype of the ΔRI σ^{54} form is enhanced by using pre-opened DNA templates and conditions favouring DNA opening such as DNA supercoiling (6,32). The lack of an appropriate state of the promoter DNA available to the ΔRI form of the σ^{54} RNAP holoenzyme may well be limiting for bypass transcription in vivo. The single gene KO enhancing bypass transcription encodes a nucleoid-associated protein (TtdR), and may influence DNA structure to normally repress bypass transcription.

Earlier work from the Gralla lab established that with single promoter studies (the gltA promoter) bypass transcription could be elevated by altering the TGCA sequence around the −12 GC element (21). Where we did observe evidence for bypass transcription in vivo we noted a non-consensus −12 promoter sequence, the place where the Region I of σ^{54} interacts with DNA (34,35). We propose that the repressive interactions made by Region I may be compensated for by some promoter sequences associated with DNA delivery into RNAP. Indeed destroying the repressive fork junction structure formed near the −12 promoter element yields bypass transcription *in vitro* (6,21,36). Overall we conclude that to increase bypass transcription *in vivo* appropriate changes in local DNA structure favoring DNA opening and diminishing the repressive contributions of the −12 promoter sequences would need to occur. This view is congruent with the following observations: (i) correlation between low-level bypass transcription with a less conserved C −12 nucleotide and (ii) an apparent lack of *in trans* encoded repressors of bypass transcription.

Overlapping promoters and competition between sigma factors are recognized as being important regulatory features of a range of genes (37–41), although rather few cases are documented for σ^{54} (40). Here, we provide further evidence that, in addition to being a target of activators for regulated gene expression, σ^{54} can act to repress gene transcription by other forms of RNA polymerase through a binding competition for promoter sites. Our analysis of any repressive capacity is unlikely to be comprehensive, since a single growth condition was studied. Given more σ^{54} binding sites than conventional activator dependent σ^{54} promoters have been identified using a range of genomic methods, we propose that the repressive function is pervasive. Indeed, the σ^{54} factor is designed to inhibit RNAP by penetrating and blocking DNA entry routes (27,42,43), and it may well be that the earliest function of σ^{54} or some of its domains was to globally repress transcription. The unusual behaviours of the cold-shock promoters with respect to σ^{54} dependence is intriguing, and suggests some non-canonical functions of σ^{54} and or its holoenzyme remain to be discovered. These may involve factors outside of the well-recognized AAA+ activators which reorganize σ^{54} at its classical promoter sites.
SUPPLEMENTARY DATA

Supplementary Data are available at NAR Online.

ACKNOWLEDGEMENTS

We thank the lab of Dr Joseph Wade (University of Albany, NY, USA) for sharing their data on genomewide ε54 ChIP binding. We also thank Goran Jovanovic, Patricia Burrows, Simone Wiesler and Joerg Schumacher for their advice and comments.

FUNDING

Biotechnology and Biological Sciences Research Council, UK Research Grant [BB/J00717X/1]. Funding for open access charge: Biotechnology and Biological Sciences Research Council, UK Research Grant [BB/J00717X/1]. Conflict of interest statement. None declared.

REFERENCES


