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RNAi screen identifies Jarid1b as a major regulator of mouse HSC activity

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Running title: Jarid1b negatively influences self-renewal

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Key Point: Jarid1b knockdown promotes enhanced HSC activity

**ABSTRACT**

Histone methylation is a dynamic and reversible process proposed to directly impact on stem cell fate. The Jumonji (JmjC) domain-containing family of demethylases comprises 27 members which target mono-, di- and tri-methylated lysine residues of histone (or non-histone) proteins. To evaluate their role in regulation of hematopoietic stem cell (HSC) behaviour we performed an *in vivo* RNAi-based functional screen and demonstrated that Jarid1b and Jhdm1f play opposing roles in regulation of HSC activity. Decrease in *Jarid1b* levels correlated with an *in vitro* expansion of HSCs with preserved long term *in vivo* lympho-myeloid differentiation potential. Through RNA sequencing analysis *Jarid1b* knockdown was associated with increased expression levels of several HSC regulators (*Hoxa7, Hoxa9, Hoxa10, Hes1, Gata2*) and reduced levels of differentiation associated genes. shRNA against *Jhdm1f*, in contrast, impaired hematopoietic reconstitution of bone marrow cells. Together, our studies identified Jarid1b as a negative, and Jhdm1f as a positive regulator of HSC activity.
INTRODUCTION

Multipotent hematopoietic stem cells (HSCs) ensure sustained production of lineage committed blood cells throughout life. The pool of long-lived HSCs is preserved due to their inherent capacity to undergo self-renewal divisions. Cell fate decisions result from integrated influences of various nuclear factors, including chromatin modifiers that regulate access of transcriptional machinery to specified genomic loci. Chromatin, consisting of intertwined DNA, histone and non-histone proteins, can be covalently modified by epigenetic effectors. DNA methyltransferases (DNMTs) modify carbon-5 of cytosine bases within cytosine-guanosine dinucleotides (CpG) frequently located proximal to or in promoters. In mammalian cells, the mechanism and effectors of DNA demethylation are less well understood, but recent reports suggest this process likely requires the activity of the TET enzyme family. Post-translational modifications of histones include methylation, acetylation, phosphorylation, ubiquitination, sumoylation or ADP-ribosylation and introduce changes in local chromatin topography that subsequently alter gene expression patterns in developmental stage and cell context-specific manners.

Methylation of histone residues represents a classical paradigm linking epigenetics to cell fate and identity, best illustrated by the antagonistic forces of Polycomb (PcG) and Trithorax (Trx) group protein complexes on regulation of key developmental loci such as Hox gene clusters. In general, PcG protein repressor complexes PRC2 and PRC1 are associated with gene silencing, and Trx complexes (compass-like or MLL, Mixed Lineage Leukemia, complexes) with gene activation. The histone methyltransferase (HMT) Enhancer of Zeste Homolog 2 (EZH2) of the PRC2 complex catalyzes tri-methylation of lysine 27 on histone H3 (H3K27me3). This covalent mark serves as a docking site for the PRC1 complex which mono-ubiquitinates lysine 119 of histone H2A (H2AK119Ub) resulting in gene silencing. MLL complexes counteract these epigenetic marks via tri-methylation of lysine 4
on histone H3 (H3K4me3) at transcription start sites, a mark associated with active gene transcription\textsuperscript{6} and recruitment of H3K27me3 demethylases UTX and JMJD3\textsuperscript{7,9}. Additional Trx-mediated modifications such as acetylation of H3K27 (H3K27Ac) and di-methylation of H3K36 (H3K36me2) further oppose the PcG mediated gene repression.

Histone methylation status on lysine (K) or arginine (R) residues is reported to evolve through highly dynamic and finely regulated processes (reviewed in\textsuperscript{10}). Histone demethylases (HDMs) integrate into multi-unit complexes resulting in removal of methylation marks by amine oxidation, deimination\textsuperscript{11} or hydroxylation. The lysine specific demethylase 1 (LSD1/KDM1A)-related HDM can demethylate mono- and di-methylated lysine residues. Jumonji C (JmjC) domain-containing HDMs (n=27) are capable of removing all three lysine methylation states by an oxidative reaction requiring $\alpha$-ketoglutarate and iron (Fe$^{2+}$) as cofactors. Like the LSD1 family, JmjC/JARID1 proteins act as components of multi-subunit complexes, with non-catalytic domains proposed to mediate protein-protein interactions involved in regulation of demethylase activity and/or target specificity (reviewed in\textsuperscript{12}). In addition, JMJD6 demethylates arginine residues\textsuperscript{13}, and bacterial Jumonji domain-containing AlkB protein is involved in DNA demethylation and repair\textsuperscript{14,15} suggesting that JmjC substrates include non-histone targets.

JmjC protein activity results in dynamic chromatin landscape changes which enable expression of distinct gene subsets required for self-renewal\textsuperscript{16}, proliferation\textsuperscript{17}, differentiation\textsuperscript{18-20}, cellular senescence\textsuperscript{21} and cancer development\textsuperscript{22-24}. In light of these findings, an established \textit{in vivo} RNAi based screening strategy\textsuperscript{25} was undertaken, in a targeted way, to assess the impact of JmjC gene down-regulation on adult primary HSC cell fate. We identify Jarid1b as a negative regulator of HSC self-renewal and progenitor cell activity, while Jhdm1f positively influences blood reconstitution. Results from these experiments and possible downstream functional networks involved are presented.
METHODS

Construction of shRNA retroviral vectors

For each gene target, 3-5 shRNAs were designed as single stranded oligonucleotides also incorporating miR-30 flanking arms using the RNAi Central shRNA design tool at http://cancan.cshl.edu/RNAi_central/main2.cgi and our previously established methodology25.

Mice

C57BL/6J (CD45.2+) transplant recipients and C57BL/6Ly-Pep3b (CD45.1+) congenic bone marrow donor mice were bred and manipulated in a specific pathogen-free animal facility. Experimental procedures were revised and approved by the University of Montreal animal ethics committee (Comité de Déontologie de l’Expérimentation sur les Animaux de l’Université de Montréal).

Flow cytometry

Negative selection of hematopoietic lineage marker (GR-1+, B220+, Ter119+) expressing cells (Lin-) was performed as described25. Lin- bone marrow fraction was subsequently stained with PE-Cy7-conjugated anti-cKit, PE-Cy5-conjugated anti-Sca1 (eBioscience), PE-conjugated anti-CD150 (BioLegend) and FITC-conjugated anti-CD48 (BD Biosciences) antibodies, followed by isolation of HSC-enriched PE-Cy5-Sca1+/PE-Cy7-cKit+/PE-CD150+/FITC-CD48/APC-Lin- cell population. Day E14.5d.p.c. fetal liver derived HSCs were purified from the Lin- cell populations by isolating the fraction of PE-Cy5-Sca1+/PE-CD11b+/PE-Cy7-CD150+/FITC-CD48/APC-Lin- cells. Cells were sorted using the FACSARia cell sorter (Becton-Dickinson, San Jose, CA, USA. The frequency of LT-HSC in the sorted populations (Table S1), the proportions of transduced (GFP+) transplant-derived
(CD45.1+) peripheral blood leukocytes \(^\text{26}\) and contribution of these cells to reconstitution of hematopoietic lineages \(^\text{25,26}\) were determined as described.

**HSC/progenitor cell culture, retroviral infection and transplantation**

Suspension cell cultures of HSC/progenitor cell enriched populations, generation of retrovirus-producing GP+E-86 cells and infection of the sorted HSC/progenitor cells were performed as described\(^\text{26}\). For validation assays, 1500 CD150\(^+\)CD48\(^-\) Lin\(^-\) bone marrow derived cells were introduced in co-culture with retroviral producers in 96-well plates\(^\text{26}\). After 5-day incubation (day 0), the total cell content of each well was harvested, and partitioned for transplantation and cell culture as previously reported\(^\text{25}\). Briefly, 1/8 of cell suspension was transplanted into sublethally irradiated congenic recipients (n=2 for each shRNA species). The remaining cells were cultured for additional 7 days and proportions corresponding to 1/8 of the cell input transplanted in 3 recipient mice (day 7). To compensate for the \(shJhdm1f\)-associated loss of reconstituting activity, ¼ of day 0 cell suspensions was transplanted in each of the 3 recipients, and no continuing suspension cultures were initiated.

**Clonogenic progenitor cell assays**

The total numbers and distributions of myeloid clonogenic progenitors in various cell populations recovered from the 5-day co-culture with retroviral producers (day 0), or from the subsequent suspension cultures (day 5-7) were determined as described\(^\text{25}\).

**Microphotography**

Images of Wright-stained cytospin cell preparations were acquired using a Leica DMIRB microscope with an HCXPL FluotarL 40x/0.6 numeric aperture objective (Leica) and a Retiga EX-i camera (Q-Imaging). Images were transformed directly into TIFF files using
Adobe Photoshop Version 6.0 (Adobe Systems). In situ images of colonies in semisolid media were acquired using the same set up, but using the HC Pl Fluotar 10X/0.30, Ph 1 lens.

**Competitive repopulating unit (CRU) assay**

These assays were performed as previously described\textsuperscript{25,26}.

**Q-RT-PCR assessment of JmjC gene expression in HSC enriched populations**

Gene expression was assessed by Q-RT-PCR using the Roche LightCycler\textsuperscript{®} 480 System (Roche, Basel, Switzerland). Reactions were performed in 384-well plates for 50 amplification cycles (95°C 10 s; 60°C 10 s; 72°C 10 s). Reference Taqman\textsuperscript{®} gene assay (\textit{Hprt}) was purchased from Applied Biosystems. Primer sequences in Table S2, Delta Ct values in Table S3.

**ChIP-chip analysis**

Two-colour hybridizations on NimbleGen MM8 Deluxe Promoter HX1 arrays were carried out using 34\textmu g of Cy5-labeled input and 34\textmu g of Cy3-labeled test (\textit{shLuc} or \textit{shJarid1b}) ChIP DNA using the NimbleGen hybridization kit as recommended by the manufacturer. Arrays were scanned at 5 \textmu m resolution using a GenePix4000B scanner (Molecular Devices). Data extraction and peak finding analysis was done using NimbleScan 2.5 and visualized in the UCSC genome browser.

**RNA sequencing analysis**

RNA sequencing was performed as previously described\textsuperscript{27}. RNA was extracted from 1x 10\textsuperscript{6} cells for each test (\textit{sh3Jarid1b-GFP}) and control (\textit{shLuc-GFP}) culture condition. Cultures were initiated with 24,000 CD150\textsuperscript{+}CD48\textsuperscript{−}Lin\textsuperscript{−} bone marrow cells and expanded for 4 days.
following infection with the respective constructs. Cells from 2 independent cultures were isolated for each condition, and only cell populations with >90% gene transfer were selected.

**shRNA-mediated knockdown in NUP98-Hoxa10ΔHD-transduced HSCs**

1500 purified CD150+CD48-Lin- bone marrow cells were co-cultured with irradiated (40 Gy) GP+E86 cells producing MSCV-NUP98-HOXA10HD-IRES-Puromycin (NA10HD) virus. After 4-day co-culture, cells were collected and cultured for 6 days in the presence of 10 μg/mL of Puromycin, harvested and stained with APC/Cy7-conjugated anti-CD43 (BioLegend), APC-conjugated anti-Sca1, PE/Cy7-conjugated anti-Gr1 and PE/Cy5-conjugated anti-F4/80 (eBioscience). The sorted NA10-transduced Sca1+CD43+Gr1-F4/80- cell subpopulation was co-cultured with GP+E86 cells producing shLuc, sh3Jarid1b or sh1Moz recombinant retroviruses. After 4 days, the nonadherent cells were recovered and expanded for additional 6 days. To assess the extent of differentiation in these cultures, cells were stained with PE/Cy7-conjugated anti-Gr1 and PE/Cy5-conjugated anti-F4/80-PE/Cy5 antibodies. Data were acquired using BD LSRII cytometer and FACSDiva Version 4.1 software (BD Biosciences PharMingen), and analyzed using the FlowJo Version 7.6.4 software (TreeStar).

**Statistical Analysis**

Statistical analysis was conducted using Prism (Graphpad Software version 5) or SPSS. The two-tailed Student-t-test was performed throughout unless otherwise stated. Statistical significance was calculated at a 95% confidence level.
RESULTS

JmjC gene expression in HSC-enriched populations

Expression profiles were obtained for all JmjC genes in sorted mouse bone marrow and fetal liver cell populations comprising ≥30% of functionally defined long-term-repopulating LTR-HSCs, as evaluated by competitive repopulation unit (CRU) assays 28 (Table S1). Results showed that all JmjC gene transcripts, except Hairless, were detected at relatively high levels (Fig. 1A). The histone acetyl-transferase MOZ (MYST3), reported as essential for HSC expansion29,30 was also highly expressed in this fraction and chosen as a control for the functional screen. Transcript levels of four JmjC genes, namely Jarid1b, Jmjd2d, Fbxl10 and Jmjd1c, were notably increased in the LTR-HSC-enriched fractions relative to total bone marrow cells (Fig. 1B). In contrast, Jhdm1d, Hspbap1 and Hairless transcript levels were notably higher in more mature cells. Genes expressed higher in the LTR-HSC enriched population e.g. Jarid1b, Jmjd2d, Fbxl10 and Jmjd1c were assigned a HSC expression score of one, while genes that were higher expressed in mature cells were attributed negative scores (Fig. 1B, lower panel). Increased expression in LTR-HSCs implied a HSC biological role, and these candidates were prioritized in the functional studies.

Functional in vivo RNAi-based primary screen

The shRNA based screen tested 23 of the 27 known JmjC mouse genes (Fig. 1C) for their potential functional role in HSC biology. shRNA Luciferase (shLuc) was used as a negative control, while shRNA Moz (shMoz) and Nup98Hoxa10-homeodomain (NA10hd) overexpression were used as controls for the loss and gain of HSC activity, respectively (data not shown for NA10hd, and31,32). HDM-directed shRNA sequences (n=112, Table S5) were subcloned into a LMP retroviral vector and assessed for their ability to modulate HSC/progenitor activity in vivo as previously reported 25 (summarized in Fig. 1D). The
biological impact of each shRNA was evaluated by serial sampling of peripheral blood from transplanted mice at early (3-4 weeks) and late (16-20 weeks) time points. To prioritize candidates for validation and downstream functional studies, a “selection threshold scoring system (STS),” weighted for HSC-enriched genes, was established (Fig. 2A and Table S3). HSC activity was determined by the long-term blood engraftment potential of the transduced donor cells (CD45.1\(^+\)GFP\(^+\) cells) based on their contribution to blood leucocytes, calculated as a proportion of the total donor (CD45.1\(^+\)) population, which comprises both transduced and non-transduced cells (%blood GFP= CD45.1\(^+\)GFP\(^+\) cells/total CD45.1\(^+\) cells). The percentage of blood reconstitution of the transduced cells was then compared to the respective gene transfer (GT) level on the day of transplantation (day0), and this ratio (Fig.2B, y-axis) used to document an increased (ratio>0.8072) or decreased (ratio<0.2751) contribution to the donor graft (HSC activity score). This ratio was used as GT levels for the hairpins presented in Fig.2B ranged from 20-80%, with an overall average of 49% as previously reported\(^{25}\) (see also Table S4 for complete data), and thus transduced cell expansion or attrition could be measured. Overall, six hits were identified: \(\text{Jmjd2a, Jhdm1f}\) and \(\text{Jmjd1b}\) (STS of 2); \(\text{Jarid1a, Jarid1b}\) and \(\text{Hif1an}\) (STS of 3). Four of these genes; namely \(\text{Jarid1a, Jarid1b, Jmjd2a, Hif1an}\), were identified as potential negative regulators of HSC activity (Fig. 2B, green shaded areas), while \(\text{Jhdm1f}\) and \(\text{Jmjd1b}\) (Fig. 2B, red area), as putative positive regulators.

**JmjC HDM knockdown validation experiments**

Hit validation was further restricted to proven JmjC domain containing histone demethylases (HIF1AN demethylase excluded as it targets HIF1\(\alpha\)) and for technical considerations validation of potential positive regulators was limited to \(\text{Jhdm1f}\). The increased hematopoietic reconstitution levels observed with shRNA mediated knockdown of the three remaining hits;
namely Jarid1a, Jarid1b, Jmjd2a (Fig.2B) prompted us to examine how the transduced HSCs would behave in vitro, where control HSCs normally exhaust rapidly\(^2\). Validation experiments were performed (Fig. 3A) and a gain-of-function phenotype for Jarid1a or Jmjd2a could not be confirmed upon shRNA transduction and prolonged in vitro culture (data not shown). Conversely, reducing Jarid1b levels in HSC populations by multiple shRNA moieties (shJarid1b 2, 3 and 4) clearly conferred an in vivo competitive advantage to freshly transduced cells compared to shLuc controls (Fig. 3B, left panel: n=3 different shRNAs to Jarid1b, day 0 transplanted cells). As higher GT rates were achieved in the validation experiments (>80%), blood reconstitution levels were measured as CD45.1\(^+\)GFP\(^+\) cell percentages. The positive impact of shJarid1b on HSC activity was even more noticeable for cells transplanted after one week of in vitro culture (Fig. 3B, middle and right panels, cells transplanted after 7-day culture) to better detect HSC gain-of-function activity, as our previous studies demonstrated \(^2\). Proportions of shJarid1b-transduced cells (GFP\(^+\)) in peripheral blood remained well above those determined for shLuc controls for up to 10 months (Fig. 3B). Knockdown of shJarid1b was determined for freshly transduced cells, and a >40% decrease in expression observed for shJarid1b 2, 3 and 4 (Fig. 3C). Decreased HSC activity relative to control cells was confirmed with multiple shRNA constructs against Jhdm1f (n=5), Fig. 3D,E. All five shRNAs against Jhdm1f had knockdown efficiencies >40\% (Fig. 3F). Validation experiments thus uncovered one negative (Jarid1b) and one positive (Jhdm1f) regulator of HSC activity. Further in vitro studies of shJhdm1f-transduced cells detected no significant change in proliferation or clonogenic progenitor activity of nucleated cells (Figure S1) indicating a distinct role for Jhdm1f in HSC repopulation ability. The striking impact of Jarid1b knockdown on blood reconstitution oriented research towards Jarid1b function.
**Jarid1b knockdown decreases hematopoietic cell differentiation in vitro**

Sh2- or sh3Jarid1b-transduced primitive hematopoietic cell populations had a ~1.5-2-fold proliferative advantage in vitro compared to shLuc controls, while Moz knockdown decreased expansion of the transduced cells (Fig. 4A, left panel). The expanded shJarid1b-transduced populations comprised high proportions of morphologically immature cells (Fig. 4A, right panel), and lower percentages of differentiated (Gr1+) myeloid cells than shLuc controls (Fig. 4B). Moreover, fractions of primitive (Gr1-) shJarid1b-transduced cells were comparable to that determined in response to NA10hd, a well characterized enhancer of in vitro HSC/progenitor cell expansion, while a marked elevation in differentiation was observed for shMoz loss-of-function control (Fig. 4B). During the 7-day culture period, the numbers of shJarid1b-transduced clonogenic progenitor cells (colony forming cell, CFC) increased ~3-fold compared to shLuc and shMoz controls (Fig.4C). Jarid1b knockdown enhanced the proliferative capacity of individual CFCs compared to shMoz controls (Fig.4D), and promoted expansion of the highly proliferative granulocyte-macrophage progenitors (Fig. 4E). Although the total CFC contents of shLuc and shMoz control cultures were comparable (Fig. 4C), and both cell populations differentiated into granulocytes and macrophages (Fig. 4E), the antiproliferative effect of Moz knockdown was evident from the decrease in sizes of shMoz clones compared to shLuc controls (data not shown).

Q-RT-PCR assays involving selected candidates revealed a marked downregulation of differentiation associated genes Hairless (Hr) and Pu.1 in shJarid1b-cells compared to shMoz control (Fig. 4F). No major changes in expression levels of genes regulating senescence (Cdkn2a (p16) and Cdkn2d (p19)) or apoptosis (Bcl2, McI1) were detected in shJarid1b-cells, while modest increases in c-Myc and decreases in Mad1 levels resembled those determined for Hoxb4-overexpressing cells (data not shown, and33). Moz knockdown was, however, clearly associated with upregulation of Hr, Cdkn2a and Cdkn2d expression. This suggests
that Jarid1b knockdown promotes in vitro HSC/progenitor expansion by suppressing differentiation, while cells remained permissive to cell cycle re-entry. In contrast, reduced Moz expression enforced commitment to differentiation and senescence cell fate pathways.

**Jarid1b negatively regulates HSC self-renewal**

Transplant recipients of shJarid1b-cells had normal numbers of total bone marrow mononuclear cells (MNC), and no splenomegaly was observed (Fig. 5A, 1st and 2nd panels). Contributions of the transduced (GFP+) shLuc- and shJarid1b-cells to myeloid progenitor cell compartments were comparable (Fig. 5A, 3rd panel), but an ~8-10-fold increase in the frequencies of shJarid1b-multilineage progenitors (Colony Forming Unit-Granulocyte, Erythroid, Macrophage, Megakaryocyte, CFU-GEMM) compared to controls was observed (Fig. 5A, 4th panel). This difference in immature progenitor cell content remained benign, and no hematological abnormalities could be detected during the 12 month observation period. Moreover, transplanted shJarid1b-cells generated normal proportions of myeloid (Mac1+/CD11b+), B-lymphoid (B220+/CD45R+) and T-lymphoid (CD4+, CD8+) progeny (Fig. 5B), suggesting that differentiation ability in vivo was not affected.

To examine if Jarid1b knockdown favoured self-renewal divisions, leading to expansion of HSC populations, equal numbers of HSC-enriched (CD150+CD48-Lin-) cell populations were co-cultured with shLuc or shJarid1b retroviral producers. HSC frequencies were determined in samples immediately after cell sorting and after a cumulative 12-day ex vivo culture period (Fig. 5C, top panel), using the CRU assay. CRU numbers in cell populations recovered from shLuc cultures (Fig. 5C, green bar) were comparable to those determined for the input cell population (Fig. 5C, red bar) suggesting no major loss or gain of HSCs during the in vitro incubation. In contrast, the CRU numbers in shJarid1b cultures increased 8- to 20-fold above shLuc control samples, or sorted cells prior to shJarid1b infection (Fig. 5C, blue bars),
suggesting that Jarid1b knockdown promoted the in vitro maintenance/expansion of long term-repopulating HSCs. Southern blot analyses of shJarid1b revealed a common proviral integration pattern between bone marrow (mostly myeloid, erythroid and B-), thymus (mostly T-) and spleen (B- and T-) cells of individual recipients (Fig. 5D) demonstrating multipotency and oligoclonal hematopoietic reconstitution of the expanded HSCs.

Impact of Jarid1b knockdown on gene expression

To examine downstream effects of Jarid1b knockdown, results of transcriptome next generation sequencing (RNAseq), Q-RT-PCR and ChIP-Chip assays were analyzed and compared to controls. The similarity in HSC expansion observed between Jarid1b knockdown and Hox gene overexpression \(^{31,33}\) and the recent report that JmjC members KDM6A (UTX) and KDM6B (JMJD3) are positive regulators of Hox gene expression \(^{21}\) prompted initial focus on the Hox family of transcription factors (Fig. 6A). Both RNAseq and Q-RT-PCR assays revealed that Jarid1b knockdown induced a 2 to 5-fold increase in 5’ Hoxa gene (Hoxa5 to Hoxa11) expression levels compared to controls (Fig. 6A and Figure S2). RNAseq analysis identified upregulation of genes associated with key pathways such as erythroid differentiation, notch signalling and stem/progenitor cell function (Fig. 6B and Figure S3), and downregulation of others involved in immune responses and differentiation (Fig. 6C and Figure S4). Several of the candidate downstream genes, including Hoxa7, Hes1, Hoxa10 and Hoxb2, were identified as potential direct epigenetic targets by enrichment of the associated H3K4me3 mark in 5’ loci as determined by ChIP-Chip assays (Fig. 6D and Figure S5). Supporting this possibility, non-regulated genes e.g. Hoxa2 demonstrated no proximal promoter H3K4me3 enrichment following Jarid1b knockdown (Figure S5). Jarid1b knockdown also correlated with downregulation of the previously reported putative JARID1 target Cav1\(^{34}\) (Table S7).
Study of epistasis between Jarid1b and Nup98Hoxa10-homeodomain

To directly examine potential gene interaction of Jarid1b and Hoxa overexpression, HSC/progenitor cell populations were first transduced with NUP98Hoxa10-homeodomain (NA10hd) and the HSC-enriched populations (Sca1^+CD43^+ Gr1^-F4/80^-; Keith Humphries, personal communication) subsequently transduced with shJarid1b, shMoz, or control shLuc. The proportions of mature GR1^+ F4/80^+ cell fractions in each condition were evaluated as a measure of differentiation (Fig. 6E, upper panel). Results demonstrate that the combined effects of NA10hd overexpression and Jarid1b knockdown were additive, resulting in significant suppression of in vitro differentiation (p<0.0001; Student t-test) below the levels determined for control NA10hd+shLuc-cells (Fig. 6E, lower panel). In contrast, shMoz targeted knockdown overrode the maturation arrest imposed by NA10hd overexpression, and enforced differentiation above the levels determined for controls (Fig. 6E, lower panel).

Together, the data presented support a role for Jarid1b in regulating key loci implicated in HSC cell fate and identify Jarid1b as a negative regulator of in vitro HSC expansion.

DISCUSSION

Following the established pipeline strategy from HSC isolation and infection to in vivo functional assessment of hematopoietic reconstitution, the presented RNAi screen highlighted Jarid1b as a negative and Jhmd1f as a positive modulator of HSC activity. Due to defined inclusion criteria other HDMs should not be excluded as potential HSC modifiers. Akin to Hox gene overexpression, differentiation was restrained in shJarid1b-transduced HSC cultures as evidenced by more primitive cell morphology, reduced granulocytic maturation, and greater expansion of clonogenic progenitors relative to controls. After transplantation, ex vivo expanded HSCs were able to resume normal lympho-myeloid differentiation in recipient
mice, in the absence of lineage skewing or hematological abnormalities, for up to one year. Logarithmic ex vivo expansion of shJarid1b-transduced HSCs was demonstrated by CRU assays indicating that Jarid1b modulation influences stem cell fate decisions. Oligoclonal origin of the repopulating HSC pool and inherent multipotency of shJarid1b-transduced parental stem cells was shown by proviral insertion patterns in long-term recipient mice. Mechanistically, the competitive advantage conferred to shJarid1b-transduced HSCs could be attributed in part to the selective up-regulation of 5’ Hoxa genes. Segmental transcription of this chromosomal region is well documented and particularly targeted by epigenetic regulators such as MLL or its derived fusion oncoproteins both in normal and leukemic stem cells. Interestingly, the 5’ Hoxa cluster is also targeted by the fusion oncoprotein NUP98-JARID1A in a mouse model of myeloid leukemia.

In agreement with the proposed substrate specificity of JARID1B for H3K4me3, we noted enrichment for this epigenetic mark on 5’ Hoxa genes indicating that Jarid1b contributes to the negative regulation of Hoxa gene expression which when relieved following knockdown, leads in part to increased HSC activity. Enhancement of NA10hd-induced maturation arrest in the presence of reduced Jarid1b levels argues for regulation of additional cell fate determinants to account for the HSC phenotype seen. Comprehensive transcriptome analysis by RNAseq identified additional downstream genes, some of which are associated with key hematopoietic or stem cell associated pathways. Recent reports further support a role for Jarid1b in transcriptional regulation of cell fate associated genes. PU.1 induction of transcription factor EGR2 was reported to recruit Jarid1b to the miR-17-92 promoter site, resulting in H3H4 demethylation and transcriptional silencing of the cluster required for monocyte maturation in a mouse model. Schmitz et al., recently reported that Jarid1b depletion prevents neuronal differentiation of ESCs by indirectly preventing H3K4
demethylation and silencing of pluripotency and germ cell-associated gene loci. Similarly, H3K4me2/1 monoamine oxidase LSD1 was also deemed essential for proper hematopoietic progenitor differentiation.

We demonstrate that transcript levels of Jarid1b are increased in HSCs suggesting either a role in preventing unrestricted self-renewal divisions or in enabling downregulation of cell fate associated genes upon lineage commitment. Interestingly, 1q32 anomalies, which include Jarid1b, are common genetic mutations found in cells of chronic myeloid leukemia (CML) patients during disease progression characterized by a block in myeloid differentiation. This observation supports the hypothesis that low Jarid1b levels maintain stem cell fate, which combined with BCR-ABL induced proliferation, could result in development of overt leukemia. In our studies, transplant recipients of shJarid1b-transduced cells never developed leukemia. However, all Jarid1b hairpins tested achieved similar gene knockdown (≤50%), and more drastic outcomes following complete Jarid1b depletion cannot be ruled out. Two Jarid1b null alleles, one (exon 1 deletion) embryonic lethal, the other (exon 6 deletion) no gross abnormalities, have recently been described. Precise elucidation of the role Jarid1b gene dosage plays in HSCs and leukemia development will thus likely require analyses of HSC-specific Jarid1b deletion.

Overall, the data presented supports a cellular and developmental stage specific effect of Jarid1b levels in modulation of HSC cell fate. To sustain stem cell fate, integrated influences from various epigenetic effectors promote an active state of transcription at multipotency loci (Fig.7 upper panel). Activity of chromatin modifiers, including JARID1B, should culminate in maintenance of the H3K4me3 epigenetic mark on these loci, and exclusion of repressive marks, such as H3K9me3 or H3K27me3. Jhdm1f/PHF8 has recently been identified as a key regulator of ATRA response in acute promyelocytic leukemia cells. Whether JHDM1F
activity influences the methylation status of H3K9 or normal HSC fate decisions remains to be explored. In contrast, removal of the active H3K4me3 epigenetic mark, potentially by JARID1B, could repress transcription of “stemness” genes and favor lineage commitment (Fig. 7, lower panel). Other JmjC candidates (e.g. Hif1an) may also be involved in this process, underscoring the importance of further characterization of these enzymes in the context of HSC regulation.
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AUTHORSHIP CONTRIBUTIONS

S.C. and G.S. established the gene candidate list. M.S., J.C and S.C. contributed to isolation and functional assessment of highly purified HSC populations used for expression profile studies of gene candidates. S.C., K.J.H. and G.S. planned and performed the initial screen. S.C., K.J.H. and N.M. designed and performed validation experiments. S.C., N.M., K.J.H., S.B.T., J.C., E.D. J.K. and M.S. contributed to HSC isolation experiments required for initial screen, validation experiments and subsequent experiments. S.C. and N.M. performed confirmation experiments for shJarid1b-transduced cells including cell culture, progenitor assays and FACS. T.M. and S.C. performed RNA extraction and Q-RT-PCR analyses involving shJarid1b transduced cells. S.C., N.M. and J.C. performed LDA experiments and

CONFLICT OF INTEREST

The authors declare no competing financial interests.
REFERENCES


FIGURE LEGENDS

Figure 1. JmjC gene expression in HSC/progenitor cell populations and selection for RNAi screen.  
A) Transcript levels of histone demethylases (HDM) in HSC enriched cell populations. Results show ΔCT values determined by quantitative (Q) RT-PCR assays (with respect to endogenous Hprt expression levels, Ct~22) and represent average ± SEM of 5 independently sorted HSC populations (bone marrow, n=3; E14.5 d.p.c. fetal liver, n=2). Frequencies of long term-repopulating HSCs in these populations are shown in Table S1.  
B) Comparison of HDM transcript levels detected in HSCs and total bone marrow cell populations. Relative transcript quantities (RQ) are shown in log2 scale, and represent the ΔCT (HSC)/ΔCT(bone marrow) ratio determined by quantitative RT-PCR assays (average ± SEM, n=3). An HSC expression score was implemented based on expression levels (ΔCT) and differential expression (RQ) of individual HDM, to rank gene candidates according to relative expression in HSC vs mature cells: gene not expressed in HSC, -2; less (<) expressed in HSC vs mature cells, -1; more (> ) expressed in HSC, +1; equally expressed, 0.  
C) List of the 23 HDM candidates tested in primary screen. HDMs sub-families sharing similarities outside the catalytic domain are denoted by different shading. Left column, the revised current terminology; central column, synonyms; right column, proposed substrate specificity10,42. Four HDM genes were excluded from the screen: Jarid1d and Uty map to chromosome Y and are thus likely not required for regulation of HSC activity; Pla3g4b belongs to the cytosolic phospholipase A2 family; Jhdm1e knockdown could not be achieved by any hairpin in two independent experiments.  
D) Schema of shRNA retroviral vector backbone (top), and experimental outline (bottom) of the primary screen at the bottom. At 16 week after transplantation, increase in the proportion of GFP+ peripheral blood leukocytes above their input levels reflects knockdown of a negative regulator of HSC activity (green box), and the inverse outcome denotes a positive HSC regulator (red box).
Figure 2. HDM hit identification. A) The selection threshold score for genes was calculated from the expression score (Fig. 1B) and HSC activity score or biological score, (Fig. 2B). Biological score represents the number of shRNAs per HDM which modulate HSC activity in recipients above or below the 95% confidence interval range established for control shLuc cells (dotted blue lines in Fig. 2B). Selection threshold score of 2 and above identifies hits selected for validation experiments. B) Contributions of GFP* (shRNA-transduced) cells to peripheral blood reconstitution of recipients at 20 weeks after transplantation. Results are presented as proportions GFP* cells within the transplant-derived (CD45.1+) peripheral blood cells and are normalized for the gene transfer efficiency determined on the day of transplantation (day 0). Green shaded areas, suppressors of HSC activity; red shaded areas, enhancers of HSC activity. Raw data for gene transfer rates and blood reconstitution levels for all recipients are provided in Table S4.

Figure 3. Validation assays for identified hits. A) Schema of experimental design. B) Long-term contribution of GFP* (shLuc+, shJarid1b+, or NA10hd+) cells to peripheral blood reconstitution of recipients; gene transfer > 80% for all conditions. Left panel: recipients of day 0 cells; central and right panel, recipients of day 7 cells. NA10hd, cells engineered to overexpress NUP98Hoxa10-homeodomain fusion protein and green fluorescent protein (GFP). C) Evaluation of Jarid1b knockdown in GFP* shJarid1b-transduced cells compared to shLuc controls. Results represent average ± SEM (n=4) RQ values determined for 6 different hairpins. Bars with dashed lines correspond to shRNA constructs (sh5,6 and 1) that did not achieve significant (ie >30%) knockdown of Jarid1b in BM HSC. D) Long-term contribution of GFP* (shLuc+, or shMoz+, or shJhdm1f+) cells to peripheral blood reconstitution in recipients of day 0 cells. Each recipient received a ¼ of the transduced cell population, or twice the number of input cells transplanted for validation experiment shown
in Fig. 3A, E). Short-term (3 weeks) contribution of GFP\(^+\) (\textit{shLuc}\(^+\) or \textit{shJhdm1f}\(^+\)-transduced) cells to peripheral blood reconstitution in recipients of day 0 cells. GT=99\% for all conditions. Experiment (as in Fig. 4D) was repeated to include all hairpins against \textit{Jhdm1f}. F) Evaluation of \textit{Jhmdm1f} knockdown in GFP\(^+\) \textit{shJhdm1f}-transduced cells compared to \textit{shLuc} controls. Results represent average ± SEM (n=3). RQ values determined for 5 different hairpins. G) Summary of screen results.

**Figure 4. Jarid 1b knockdown decreases hematopoietic cell differentiation in vitro.** A) Left panel: \textit{Jarid1b} knockdown increases yields of mononuclear cells in cultures initiated with shJarid-transduced HSC/progenitor cell populations (mean ±SD, n=2). Right panel: Wright-stained cytopsin preparations of cells on day 9 of culture, 40x magnification, white arrows indicate primitive cells. B) \textit{shJarid1b} suppresses \textit{in vitro} differentiation of HSC/progenitor cell populations. Proportions of Gr1\(^+\) cells on day 9 of culture were determined by flow cytometry. Each dot represents an independent culture. C) \textit{Jarid1b} knockdown enhances \textit{in vitro} expansion of myeloid colony forming cells (CFC). The increase in CFC numbers was calculated from MNC and CFC numbers determined on days 2 and 9 (mean ± SD, n=4). D) Images of the predominant colony types. \textit{shJarid1b}, colony forming unit granulocyte-macrophage (CFU-GM), high proliferative potential; \textit{shMoz}, colony forming unit macrophage (CFU-M), low proliferation. Left panels, bright field; right panels, epifluorescence. E) Proportions of the highly proliferative CFU-GM in cultures of shRNA-transduced cells (mean ± SD, n=4). F) Q-RT-PCR-based comparison of cell fate-associated transcript levels in day 7 (Fig. 3A) \textit{shJarid1b} and \textit{shMoz}-cells compared to \textit{shLuc} controls.

**Figure 5. In vitro-expanded shJarid1b-HSC retain long-term in vivo multipotency.** A) Analysis of hematopoietic tissues in recipients of day 7 cells (Fig. 3B) at 1 year after
transplantation. From left to right: first panel, spleen weight; second panel, the total numbers of bone marrow cells pooled from pelvis, femur and tibia; third panel, CFC frequency in the GFP+ bone marrow cell populations; fourth panel, proportions of GFP+ multilineage progenitors ( Colony Forming Unit-Granulocyte, Erythrocyte, Monocyte, Megakaryocyte, CFU-GEMM). Dots in first plot represent the numbers of individual mice for which all the described parameters were analyzed. B) Contribution of day 7 (Fig. 3A) shJarid 1b (GFP+) cells to reconstitution of myeloid (Mac1+), B-lymphoid (B220+) and T-lymphoid lineage (CD4+, CD8+) at 1 year after transplantation. An example of typical reconstitution observed in all recipients (n>10) is shown. C) Jarid1b knockdown promotes the in vitro expansion of LTR-HSCs. Upper panel, experimental outline. Lower panel, CRU numbers in freshly sorted (i.e., input) and day 7 shRNA-transduced cell populations (mean ± SE). shJarid1b CRUs were determined in 2 independent experiments (see Table S6). D) Clonal analysis of proviral integrations in DNA isolated from hematopoietic tissues of mouse from shJarid1b cohort introduced in Fig. 5C. DNA was digested with EcoRI which cuts once within the provirus such that each DNA fragment recognized by the 32P-labelled Gfp probe represents a unique integration event. Mouse ID#, the total dose of transplanted cells, and the estimated number of transplanted CRU are shown on top. T, thymus; S, spleen; BM, bone marrow.

Figure 6. Jarid1b knockdown modulates molecular mechanisms implicated in maintenance of stemness. A) Quantification of Hox gene transcripts in shJarid1b-cells and shLuc controls as assessed by RNA sequencing (RNAseq) analysis. For each Hox cluster (a and b) genes, FPKM (fragments per kilobase per million reads) expression values are shown for both conditions. Error bars indicate SD. RNA was isolated from HSC enriched cells in culture (4 days following retroviral infection), and only cultures with gene transfer rates >90% were selected. For each condition, two biological replicates were sequenced. B)
Average FPKM and fold-change expression values of the 40 most upregulated (FPKM >1 for shJarid1b-cells) and C) downregulated (FPKM >1 for shLuc controls) genes from the RNAseq experiment described in Fig. 6A. Genes annotated to specific functions are specified by a cross mark in respective columns. Complete data for all differentially expressed genes (q-value < 0.05; all FPKM values included) shown in Table S7. D) Enrichment for H3K4me3 marks (black peaks) at the Hoxa7 and Hes1 loci in shJarid1b cells. Chromatin immunoprecipitation was carried out using day 7 (Fig. 5C) shJarid1b or shLuc-cells. Total H3K4me3 levels are presented in Fig. S6. E) Top panel: Experimental strategy for generation of Nup98Hoxa10-homeodomain (NA10hd) plus shRNA overexpressing cells. Following puromycin selection, the Sca1+CD43− Gr1−F4/80− NA10hd-transduced cells were infected with shLuc−, shMoz−, and shJarid1b 1b-carrying retroviruses. Lower panel: Jarid1b knockdown suppresses differentiation of NA10hd overexpressing cells. Proportions of Gr1+F4/80+ (i.e. differentiated) cells in cultures were determined by flow cytometry on day 7 after shRNA transduction. Each dot represents individual culture comprising the transduced progeny of 1,500 CD150+CD48-Lin− bone marrow cells. ¹: Manual curation 2,3 : 3.4- and 3.7-fold enrichment with false discovery rate of 4.6E-24 and 4.2E-11 in Gorilla bioinformatic tool. *Denotes high H3K4me3 densities, refer to Fig. 6D.

**Figure 7. Proposed model for JARID1B activity in modulation of HSC fate.** Grey shaded area: JARID1B erases the tri-methyl marks of H3K4 at stemness loci and represses activity of multipotency genes. Pink area: Decrease in JARID1B activity shifts balance in favour of histone methylases, preserving the active H3K4me3 mark at stemness loci to sustain multipotency.
KDM2A | JHDM1A / FBXL11 | H3K36me2 / 1
KDM2B | JHDM1B / FBXL10 | H3K36me2 / 1, H3K4me3
JHDM1D / KIAA1718 | H3K9me2 / 1, H3K27me2 / 1
JHDM1F / PHF8 | H3K9me2 / 1, H4K20me
KDM3A | JMJD1A / JHDM2A / TSGA | H3K9me2 / 1
KDM3B | JMJD1B | H3K9me2 / 1
KDM3C | JMJD1C | H3K9me2 / 1
KDM4A | JMJD2A / JHDM3A | H3K9me3 / 2, H3K36me2 / 1
KDM4B | JMJD2B / JHDM3B | H3K9me3 / 2, H3K36me2 / 1
KDM4C | JMJD2C / JHDM3C / GASC1 | H3K9me3 / 2, H3K36me2 / 1
KDM4D | JMJD2D / JHDM3D | H3K9me3 / 2, H3K36me2 / 1
KDM5A | JARID1A | H3K4me3 / 2
KDM5B | JARID1B | H3K4me3 / 2
KDM5C | JARID1C | H3K4me3 / 2
KDM6A | UTX | H3K27me3 / 2
KDM6B | JMJD3 | H3K27me3 / 2
JMJD4
JMJD5
JMJD6 / PTDSR / PSR | H3R2me2, H4R3me
HSPBAP1
HIF1AN
HIF1a-N803
Puro
IRES
GFP
5-day infection
CD150+ CD48- Lin-
Bone marrow cells
CD45.1 cells
+ve HSC regulator
-ve HSC regulator
Gene transfer evaluation (%GFP)
Transplantation (% well)
RNA
CD45.1 : CD45.2
1:1
5-day infection
shRNA histone demethylase
+5′ miR30
+3′ miR30
Pggk
Puro
IRE
GFP
LTR
CD150+ CD48- Lin-
Bone marrow cells
CD45.1 cells
+ve HSC regulator
-ve HSC regulator
Gene transfer evaluation (%GFP)
Transplantation (% well)
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CD45.1 : CD45.2
1:1
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+5′ miR30
+3′ miR30
Pggk
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CD150+ CD48- Lin-
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CD45.1 : CD45.2
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shRNA histone demethylase
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+3′ miR30
Pggk
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GFP
LTR
CD150+ CD48- Lin-
Bone marrow cells
CD45.1 cells
+ve HSC regulator
-ve HSC regulator
Gene transfer evaluation (%GFP)
Transplantation (% well)
RNA
CD45.1 : CD45.2
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LTR
CD150+ CD48- Lin-
Bone marrow cells
CD45.1 cells
+ve HSC regulator
-ve HSC regulator
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+3′ miR30
Pggk
Puro
IRE
GFP
LTR
CD150+ CD48- Lin-
Bone marrow cells
CD45.1 cells
+ve HSC regulator
-ve HSC regulator
Gene transfer evaluation (%GFP)
Transplantation (% well)
Selection Threshold Score (STS) for hit identification

\[ \text{STS} = \text{IF} \left( (\text{HSC activity score: Fig. 2B}) + (\text{Expression score: Fig. 1B}) \right) \geq 2 \rightarrow \text{HIT} \]

Cellot S. et al, Fig. 2
A) CD150<sup>+</sup> CD48<sup>-</sup> Lin<sup>-</sup> Bone marrow cells

5-day infection

B) % GFP blood

in vitro

Day 0

Day 7

4 mo post Tx

3 mo post Tx

10 mo post Tx

C) % knockdown

Jarid1b

Gene

D) % GFP blood

E) % GFP blood

F) % knockdown

G) 23 gene candidates

1<sup>st</sup> screen

6 hits

validation

2 hits

1 +ve regulator

Jhdm1f

1 -ve regulator

Jarid1b

Cellot S. et al, Fig. 3
A. Hox genes

B. Upregulated genes

C. Downregulated genes

D. Hoxa Cluster

E. Hes1 Cluster

For personal use only.
MULTIPOTENCY

ON

DIFERENTIATION

OFF

“stemness” genes
(Hoxa, others?)

JARID1B

Cellot S. et al, Fig. 7