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Activated lignin–chitosan extruded blends for efficient adsorption of methylene blue

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Abstract
This work investigates the production of activated lignin-chitosan extruded (ALiCE) pellets with controlled particle size distribution (almost spherical: \(dp \sim 500–1000\mu m\)) for efficient methylene blue adsorption. The novel preparation method employed in this study successfully produced activated lignin-chitosan pellets. Structural and morphological characterizations were performed using BET, FTIR and SEM-EDX analyses. The influence of contact time, solution pH, ionic strength, initial adsorbate concentration and desorption studies was investigated. The experimental data fitted well with the Langmuir isotherm \(R^2 = 0.997\), yielding a maximum adsorption capacity of 36.25mg/g. The kinetic data indicated that methylene blue (MB) adsorption onto ALiCE can be represented by the pseudo second-order-model with intraparticle processes initially controlling the process of MB adsorption. Overall, these results indicate that the novel ALiCE offers great potential for removing cationic organic pollutants from rivers and streams.

Keywords: Adsorption; Lignin; Chitosan; Dyes; Blends Extrusion; Activated Carbon.

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1. Introduction

The elimination of organic dyes from rivers and streams is a key environmental challenge [1]. Adsorption processes offer a mechanism for the removal and recovery of metals, dyes, oils, organic mixtures and pharmaceuticals from water [2, 3]. The advantages of liquid phase adsorption include: low cost, simplicity of design and operation [4], absence of sludge and harmful by-product formation. This has been acknowledged by the United States Environmental Protection Agency (USEPA) by classifying adsorption as one of the top control methods [5]. Adsorption rates and adsorption quantities are two of the most significant performance indices of any adsorption system. Hence, it is essential to design and prepare efficient adsorbents which are able to remove large amount of pollutants quickly using small amounts of adsorbent. Various adsorbent materials have been tested for the effective removal of dye such as methylene blue (MB) include spent olive stone [6] and raw dolomite [2]. Table 1 provides a comparison of the adsorption capacities of various adsorbents for the removal of MB dye [7-11]. Methylene blue has been explored in various medical applications including antimicrobial chemotherapy, biomedical dyes for cell staining, phototherapy and cancer research. Compared to other toxic materials and dyes, MB is not a particularly hazardous compound, though, large doses of MB (>7.0 mg/kg) can cause high blood pressure, mental disorder, nausea, abdominal pain, etc. [12]. Also, MB can be freely photo-sensibilized by white light producing very reactive singlet species i.e. oxygen ($^{1}\text{O}_2$). These singlet oxygens can cause destruction to the DNA structures, especially at high concentrations and, ultimately, spread to the food chain. Pure lignin and chitosan have been employed individually and blended to remove dyes from aqueous solutions. Chitosan is a biopolymer with many advantages: non-toxicity, biocompatibility and biodegradability that can be obtained from the N-deacetylation of chitin. However, chitosan is expensive, chemically unstable and can easily agglomerate. Lignin is an amorphous aromatic polymer with complex structure formed by phenylpropane.
units. Lignin is well known as a cheap waste by-product from paper and wood mills. The structural units of lignin vary depending on the source of biomass from which lignin is extracted. The development of bio-composites and blends containing chitosan and lignin by-product as adsorption materials will decrease the total cost of the adsorbent due to decrease in the usage of costly chitosan, and offer a chance for exploiting the renewable by-products produced in industries. Generally, composite/blended adsorption materials are prepared by using many organic solvents, acid/basic and ionic solutions.

This paper investigates the development of a novel adsorbent, activated lignin-chitosan extruded (ALiCE) pellets, for dye adsorption. In this study, the preparation method used very few solvents and was easily operated in a continuous mode (extrusion). Also, the produced material had a targeted particle size, effectively attracted the methylene blue dye with high adsorption capacity and exhibited superior physical and chemical stabilities. A targeted particle size means that the adsorbent can be explicitly manufactured to suit specific applications of water treatment; pilot plant packed columns. The lignin-chitosan materials were blended and extruded with acetic acid before thermal activation under N₂ gas to enhance the mechanical and physicochemical properties of the extrudates. Then, the application of the prepared activated lignin-chitosan extrudates (ALiCE), as an adsorption material for methylene blue, was investigated. The influence of experimental conditions such as contact time, initial pH, ionic strength and dye concentration on the adsorption were elucidated and the adsorption kinetics were calculated to establish the efficiency of these novel materials.

2. Material and procedure

Lignin (Alkali low sulfonate content), chitosan (95% deacetylated), methylene blue (C₁₆H₁₈ClN₃S: MW 319.85 g/mol) and acetic acid ≥99% were obtained from Sigma–Aldrich, UK. Deionized water (resistivity 18.24 Ω·cm) was used to prepare synthetic dye solutions.
2.1. Preparation of activated lignin-chitosan extrudates (ALiCE)

A Caleva mini mixer (Multi Lab: serial number 800106) with a 30cm³ stainless steel bowl, within which are two contra-rotating blades with a 0–100rpm rotational speed, was used to prepare the lignin–chitosan blends. An accurately weighed 20g of lignin and chitosan (10g of each) was mixed for 3min at 60rpm to obtain a uniform powdered mixture. Then, the mixture was further granulated with 20cm³ of aqueous acetic acid (10% v/v) solution for 10min at 60rpm, followed by extruding the wet mass at 60rpm using a Caleva single extruder (MSE: serial number 1037) fitted with a round-shaped screen (8 holes: perforation diameter: 1mm & thickness = 0.2mm). The extrudates were then spheronised for 5 mins in a Caleva spheroniser (MBS: serial number 251101) containing a friction plate with cross-hatched geometry to produce pellets with a size range of ~500–1000µm. Finally, the wet pellets were dried overnight at 80°C, followed by activation in a three zone tabular furnace (TZF 12/65/550) under nitrogen atmosphere. Activation conditions were as follows: material mass = 5g; 2 hours @ 200, 400 and 600°C (ramp rate of 20ºC/min); N₂ at a flow rate of 56cm³/min. The activated lignin–chitosan extruded (ALiCE) material was finally rinsed with deionized water after which it was dried at 80°C for 12 hours before use.

2.2. Characterisation of ALiCE adsorbent

The surface area of the ALiCE was calculated from N₂ adsorption measurements completed at liquid N₂ temperature, 77 K, using the Brunauer, Emmett and Teller (BET) equation. Fourier transform infrared spectroscopy (FT-IR), energy dispersive X-ray analysis (EDX) and scanning electron microscopy (SEM) were employed to examine the surface characteristics before and after adsorption using a Perkin Elmer Spectrum 100 within the range of 400–4000 cm⁻¹ and JEOL-JSM 6400 scanning microscope, respectively. The zeta potential measurements were carried out using a Malvern zetasizer. The oxygenated acidic and basic surface groups were determined using the Boehm's titration method [13]. Single pellet diametric compression
measurements were performed using a TA.XT2i Texture analyser materials testing machine with a PC for real time data logging and analysis.

2.3. Batch studies

All the experiments were performed with artificial wastewater stock solution of 1000mg/dm$^3$ prepared by dissolving a precise amount of methylene blue in deionized water. Experiments were performed by mixing 0.05g of ALiCE adsorbent with 25cm$^3$ of MB solutions in a 50cm$^3$ screw type glass jar at room temperature (~20°C). The pH of MB solutions is obtained using a pH meter (Orion 3-Star). Good contact is made between ALiCE adsorbent and MB solutions by agitating at 110rpm on a mechanical shaker (Gerhardt type LS5). The MB adsorption was investigated at pH (2–9) with $C_o$ of 43.65mg/dm$^3$ and dosage 2.0g/dm$^3$, initial MB concentrations (10.15–82.01mg/dm$^3$). After equilibrium, the adsorbent was isolated from the MB solution by centrifugation at 5000rpm for 5min. MB concentrations were obtained spectrophotometrically by detecting the absorbance at $\lambda_{max} = 664$nm using a Perkin Elmer LAMBDA 25, UK. For the contact time experiments at different MB concentrations, 10.15–82.01mg/dm$^3$, the same conditions were applied (optimal pH = 7 and room temperature ~20°C), and the solution volume used was 250cm$^3$. The $q_e$ values obtained from the contact time experiment were used to construct the $q_e$ vs $C_e$ plots and the isotherm analysis at 20°C. In order to study the effect of ionic strength: LiCl, KCl and NaCl salts were added to MB solutions at pH = 7, $C_o = 43.65$mg/dm$^3$ and 20°C. Adsorption experiments were performed in duplicate and average values were reported. The standard error was less than 5%. Desorption experiments were carried out in order to investigate the regeneration of the adsorbent and evaluate the mechanism involved in MB adsorption. The loaded ALiCE sample from the pH experiment (pH = 7) was used for the desorption study. Loaded ALiCE was immersed in 25cm$^3$ of desorbent solutions: 0.11M HCl, deionised water and 0.1M NaOH and the suspensions were shaken for 12h. After that, the ALiCE material was separated and the amount of MB desorbed
was computed. Finally, it is worth mentioning that after adsorption-desorption process, the adsorbent pellets remained intact.

2.4. Theory of adsorption, isotherms and kinetics

Methylene blue uptake, $q$ (mg/g), and percentage of removal (%) were obtained according to Eq. (1) and (2), respectively:

$$ q = \left[ \frac{C_o - C_e}{M} \right] \times V $$

(1)

The percentage removal = $\left[ 1 - \frac{C_e}{C_o} \right] \times 100\%$  

(2)

where $C_o$ is the initial and $C_e$ is the equilibrium concentration of methylene blue (MB) in mg/dm$^3$, $M$ is the amount of adsorbent in grams and $V$ is the volume of the MB solution in dm$^3$.

The Lagergren’s pseudo-first and second-order [14, 15] and intraparticle diffusion [16] models were used to describe the kinetic data tested:

$$ q_t = q_e \left( 1 - e^{-k_1 t} \right) $$

(3)

$$ q_t = \frac{k_2 q_e^2}{(1 + k_2 q_e t)} t $$

(4)

$$ q_t = k_{id} t^{1/2} + C $$

(5)

where $k_1$ (1/min) and $k_2$ (g/mg min) are the pseudo-first-order and the pseudo-second-order rate constants, respectively. $k_{id}$ is the intraparticle diffusion rate constant (mg/(g min$^{1/2}$)) and $C$ the intercept related to the boundary layer effect. The adsorption initial rates were calculated from pseudo-first and second-order models by using the equations:

$$ h_{0,1} = k_1 \times q_e $$

(6)

$$ h_{0,2} = k_2 \times q_e^2 $$

(7)
The isotherm adsorption data was represented by a number of isotherm equations namely: Langmuir [17], Freundlich [18], Redlich-Peterson [19] and Sips [20] adsorption isotherm models:

4. Langmuir isotherm: \( q_e = q_{\text{max}} \left( \frac{bC_e}{1 + bC_e} \right) \) \hspace{1cm} (8)

5. Freundlich isotherm: \( q_e = K_F C_e^{1/n} \) \hspace{1cm} (9)

6. Redlich-Peterson isotherm: \( q_e = \frac{K_R C_e}{1 + a_R C_e^\beta} \) \hspace{1cm} (10)

7. Sips isotherm: \( q_e = q_{\text{max}} \left( \frac{K_s C_e^{n_s}}{1 + K_s C_e^{n_s}} \right) \) \hspace{1cm} (11)

where \( q_{\text{max}} \) (mg/g) and \( b \) (dm\(^3\)/mg) are the Langmuir isotherm constants; \( K_F \) (mg/g (dm\(^3\)/mg\(^{1/n}\)) and \( 1/n \) are related to the adsorption capacity of the adsorbent and adsorption intensity, respectively; \( K_R \) (dm\(^3\)/g), \( a_R \) and \( \beta \) are the Redlich-Peterson isotherm constants, where: \( 0 \leq \beta \leq 1 \). The Sips isotherm: \( K_s \) (dm\(^3\)/mmol) is associated with the energy of adsorption and \( n_s \) indicates the system heterogeneity.

The fundamental characteristics of Langmuir isotherm, defined by Webber and Chakkravorti [21], can be expressed by a dimensionless constant called separation factor, \( R_L \):

\[ R_L = \frac{1}{1 + bC_o} \] \hspace{1cm} (12)

\( R_L \) indicates the shape of isotherm as follows: and \( R_L = 0 \) (irreversible); \( R_L > 1 \) (unfavourable); \( R_L = 1 \) (linear); \( 0 < R_L < 1 \) (favourable).

3. Results and discussion

3.1. ALiCE characteristics

One of the main objectives of this study is to prepare activated carbon with targeted size and high mechanical strength. The final ALiCE pellets were semi spherical with diameters from
400‒800µm and approx. 1mm long. The char yields after 200, 400 and 600°C carbonization were 67.7, 61.3 and 56.4 wt%, respectively. However, the prepared ALiCE adsorbent particles at 600°C are the most robust with the highest mechanical strength and lowest dust content even in comparison to commercial activated carbon, AC (Filtrasorb FS-400). The calculated failure loads for the ALiCE at 200, 400, 600°C and for AC are 5.463, 7.332, 9.597 and 8.218N, respectively. Based on that, the ALiCE prepared at 600°C was used in the adsorption experiments. The specific surface area of ALiCE was calculated as 80.77 m²/g, which is larger than that for chemically prepared lignin-chitosan blends (2.44 m²/g) [22], pZ4 activated sample (25.19 m²/g) and smaller than pP6 activated sample (1041.43 m²/g) prepared in [23]. Adsorbent surfaces could comprise chemical functional groups of acidic or basic nature. Such groups often have an effect on the adsorption process of pollutants in wastewater. The acidity and basicity of ALiCE surface values were calculated as: acidity = 0.279 mmol/g and basicity = 0.047 mmol/g. It is obvious that the acidic characteristic of ALiCE is higher than its basic characteristic. It is known that, carbons with high oxygen content show acidic surface properties and cation exchange behaviour.

3.2. Effect of solution pH and ionic strength

The adsorption removal capacity depends on the adsorbent surface properties and adsorbate structure which are greatly affected by pH. The measurements of zeta potential and influence of the pH solution value on adsorption of the MB was investigated and the results are illustrated in Figure 1. At natural pH, the zeta potential measured for ALiCE adsorbent is approx. –20 mV. This value indicates that ALiCE surface is negatively charged. Starting from the ALiCE natural pH, altering the pH of MB solutions indicates the difference of the zeta potential against the pH of the aqueous phase in the range 2–9. The MB removal efficiency increases slightly with an increase with the pH value from 2 to 4, and then the efficiency to some extent rises when the pH elevated from 4 to 7. The maximum adsorption percentage removal was 87.69% at pH
7. Additional increase in the pH beyond 7 consequences in a small reduction in the removal effectiveness. At high pH values, the presence of high concentrations of OH$^-$ ions increases the negative charge on the ALiCE surface by deprotonating the positively charged sites. These phenomena are attributed to the value of the measured zeta potential. The negative charge value was sharply enhanced when the initial solution pH was increased from 5 to 7. At pH values > 7, the ALiCE surface has more anionic charges, and the surface would participate in cation attraction and cation exchange reactions. Thus, the barrier to dye molecules diffusion decreases and results in a high adsorption capacity due to the significantly high electrostatic attraction which occurs between the positive dye molecules and the negatively charged ALiCE surface. Finally, at low pH, while the ALiCE less negative surface does not favour the adsorption process as a result of the electrostatic repulsion, a substantial amount of dye adsorption onto ALiCE still takes place at low pH values. This indicates that other mechanisms other than electrostatic attraction are operative for the MB adsorption onto ALiCE. The effect of ionic strength was investigated by adding LiCl, KCl and NaCl salts to the MB solutions. It was found that the cations can change the surface property of ALiCE in the order $K^+ > Na^+ > Li^+$. The adsorption efficiency slightly decreased from 87.69 to 81.12, 83.42 and 84.07 due to the addition of $K^+$, $Na^+$, $Li^+$, respectively. The hydrated radius of $K^+ = 2.32$ Å which is smaller than $Na^+ = 2.76$ Å and $Li^+ = 3.4$ Å. This explains the stronger competition between $MB^+$ and $K^+$ and as a results the more obvious influence on MB adsorption compared with the other two cations. However, the small reduction in the MB adsorption efficiency indicates the selective adsorption of ALiCE for MB. Similar trends have been reported for Adsorption of methylene blue onto multiparous palygorskite modified by ion beam bombardment [24].

3.3. Effect of contact time and initial concentration

The effect of exposure time on the performance of ALiCE in adsorbing MB was investigated at different concentrations, 10.15 to 82.01 mg/dm$^3$. As shown in Figure 2, the MB adsorption
uptake values increased from 4.60 to 27.90mg/g, due to the increase in the driving force from the concentration gradient. The adsorption process was fast in the initial phase ~150min and the adsorption capacities increased quickly. Subsequently the adsorption of MB sustained at a slower rate until the adsorption rate attained a constant value at its maximum level after approx. 40h of reaction; maximum levels were found for all concentrations as shown in Figure 2. The fast initial adsorption rates are attributed to the high number of accessible free functional groups e.g. –OH and –NH₂ on the surface of the ALiCE adsorbent at the start of the adsorption process and less steric hindrance for the approaching dye molecules. The slow adsorption process is a result of the reduction in the available adsorption positions and the build-up of MB molecules on the surface of the ALiCE adsorbent; this hinders the diffusion of more MB molecules into the ALiCE pores. Slow diffusion rates are related to pores which are of a similar size to the diffusing molecules [25]. Furthermore, in order to demonstrate how near the processes at any time are to equilibrium, the values of the fractional uptake $f(q_t/q_e)$ were plotted against time $t$ (Figure 2B). Figure 2B illustrates that the fractional uptakes $f$ decrease and the time necessary to achieve equilibrium increases with increased initial MB concentrations.

3.4. Kinetic modelling
The kinetic results were fit to Eqs. (3) and (4) using SigmaPlot 11.0 nonlinear regression software; the fitting results and the values of the calculated constants are shown in Figure 2A and Table 2, respectively. It was found that $k_1$ and $k_2$ of the pseudo-first and second-order rate constants decrease with an increase in the MB concentration ($C_o$) values. This is attributed to competition between higher levels of MB molecules for the adsorbent active sites. The tendency that $k_1$ declines with increasing $C_o$ suggests that it is quicker for a system with a lower $C_o$ to attain a definite fractional uptake. This phenomenon is supported by the trends in plots in Figure 2B. From Figure 2A, the first-order equation of Lagergren has described the kinetic data over the early
stage of the adsorption processes only, especially at high MB concentrations but not over the
enter contact time. Conversely, kinetics of the adsorption of MB onto ALiCE best fit the
pseudo-second-order model for the whole contact time range with correlation coefficient, $R^2$,
values close to unity and agreeing well with calculated adsorption capacities, $q_e$, with the
experimental data $q_{exp}$. This indicates that the adsorption mechanism depends on the adsorbent
and adsorbate, and the chemisorption is the rate-controlling step containing valence forces
through the exchange/sharing of electrons. The calculated values of $h_{0,1}$, $h_{0,2}$ as well as the rate
constant of external mass transfer, $k_s$, at the start of adsorption (determined from the plots of
$C_t/C_o$ against time at various $C_o$ values (plots are not shown here)) are revealed in Figure 3.

The overall trend observed in Figure 3 is a decline in the initial rates of adsorption with increase
in the initial MB concentrations. The decrease of $k_s$ with increasing $C_o$ was previously reported
for dye adsorption and as before is attributed to the MB dye molecules competing for accessible
surface area. This competition appears to have more impact on the adsorption initial rates than
the increasing driving force caused by higher dye concentration. Also, it was observed that the
estimation of the adsorption initial rate by $k_s$ is more consistent than $h_{0,1}$ and $h_{0,2}$. This is due to
(i) $k_s$ is calculated without any hypothetical assumptions regarding the adsorption mechanism:
it is obtained directly from the temporal change of $C_t/C_o$; (ii) $k_s$ is accurately calculated at $t = 0$, from fitting $C_t/C_o$ values for a short time at the start of adsorption, whereas $h_{0,1}$ and $h_{0,2}$ are
calculated from estimates of $q_e$ and $k$ throughout the entire adsorption period.

3.5. Diffusion parameters
The entire process of adsorption can be divided into four major steps: (i) dye molecules travel
from the bulk solution to the adsorbent surface by bulk diffusion. The transport of dye
molecules through the boundary occurs due to random molecular motion of individual
molecules [26]; (ii) film diffusion; (iii) intra-particle or pore diffusion of the dye (iv) chemical
reaction or complex formation [27]. The plots of the intraparticle diffusion (Eq. 5) display
multi-linearity, demonstrating three stages which are taking place, related to steps (ii), (iii) and (iv) (Figure 4A). Stage (i) occurs too quickly to be observed. The time needed for the second step is typically related to the changes in the system (i.e. temperature, adsorbent particle size and solute concentration, and), which make it complicated for prediction or control [28]. The calculated $k_{id}$ values are presented in Table 3. It can be seen that the rate constants for intraparticle diffusion increased with increasing MB initial concentrations which are attributable to the greater driving force at higher $C_0$ values. At the beginning of the diffusion process (short times), Fick’s law reduces to Equation 13 [29, 30]:

$$\frac{q_t}{q_e} = 6 \left( \frac{D_1}{\pi r^2} \right)^{0.5} t^{0.5}$$

(13)

where $D_1$ is the film diffusion and $r$ is the radius of the ALiCE particle.

For longer times, the relationship between weight uptake and diffusion can be described by the approximated Boyd model [31]:

$$Bt = -0.4977 - \ln(1 - F)$$

(14), where $F = q_t/q_e$,

The intraparticle (pore) diffusion, $D_2$ (cm²/s), can be determined by plotting $Bt$ against time; from the equation: $Slope = B = \pi^2 D_2 / r^2$

(15)

The rate of mass transfer is controlled by pore diffusion if the plots of $Bt$ vs $t$ at different MB concentrations linearly pass via the origin. If the plots are either linear or nonlinear and do not pass through the origin, the adsorption rate is ruled by chemical reaction or film diffusion [32].

From Figure 4B, it can be confirmed that the adsorption rate is not limited by pore diffusion during the initial period as the plots do not pass through the origin and that film diffusion or chemical reaction limits the rate of adsorption through this early stage [32]. However, for longer MB adsorption times pore diffusion is the controlling process. Results show that the film and pore diffusion decreased as the initial MB concentration increased. The decrease in $D_1$ and $D_2$ with increasing MB concentration is attributed to a reduction in the number of available open
sites for adsorption with increasing initial concentration. The results also show that the film
diffusion is faster than the pore diffusion. Diffusion coefficients in the order of $10^{-6}$ to $10^{-8}$ are
found when the rate is governed by film diffusion whereas the diffusion coefficients are in the
order of $10^{-11}$ to $10^{-13}$ when the rate is controlled by pore diffusion [33]. Based on the diffusion
coefficients magnitudes in Table 3, both film and intraparticle (pore) processes are initially
involved in the MB adsorption process and that pore diffusion is the sole controlling process
over longer periods.

3.6. Equilibrium isotherms
The equilibrium data was tested by Langmuir, Freundlich, Redlich–Peterson and Sips
isotherms and the plots are illustrated in Figure 5A. The isotherms constants are shown in
Table 4. As can be observed from Figure 5A and Table 4, apart from the Freundlich isotherm
equilibrium, the equilibrium data fitted well to all isotherm models with a correlating constant
($R^2$) higher than 0.99. The good fit of experimental equilibrium data of the adsorption of MB
onto ALiCE to the Langmuir isotherm indicates the chemisorption and monolayer adsorption
onto the ALiCE surface where adsorption sites are identical with similar affinities. The
maximum adsorption capacities calculated by Langmuir and Sips were very similar and
relatively high. High adsorption capacity is expected due to the relatively high surface area of
ALiCE. The Redlich-Peterson constant, $\beta$, was close to unity (0.965), demonstrating that the
isotherm can be reduced the Langmuir isotherm form [6]. Also, the values of $1/n$ were higher
than unity implying that the adsorption process is chemical. The $R_L$ values are shown in Figure
5B (0.448 to 0.091) and suggest that the adsorption of MB onto ALiCE is a favourable one,
and at high $C_0$ values, the adsorption is nearly irreversible. Finally, the surface area of the
ALiCE was calculated using MB adsorption capacity ($q_{max}$) and compared to that obtained by
the BET method ($N_2$ adsorption in gaseous phase). Assuming a complete adsorption of MB as
a monolayer on the ALiCE surface, the surface area can be calculated by:
Surface area ($m^2/g$) = \( \frac{q_{\text{max}} \times A_{MB} \times N_L \times 10^{-3}}{MW} \)  \hspace{1cm} (16)

where \( A_{MB} \) is the surface area taken by one molecule of MB = $130Å^2$, \( N_L \) is the Avogadro’s number = $6.023 \times 10^{23}$ molecule/mole and \( MW \) is the molecular weight of MB = 320$g/mol$.

The surface area of the ALiCE by MB adsorption was 88.69$m^2/g$. This value is close to that obtained by the BET method (80.77$m^2/g$). The slightly higher surface area accessed using the MB adsorption method is attributed to the MB aggregation and the instrumental theory of a cylindrical pore volumes which is not necessarily the case [34].

3.7. Desorption studies

The MB-loaded-ALiCE was eluted using HCl, deionized water and NaOH, and the amount of MB desorbed (%desorption) was determined using the following relationship:

\[
\text{Desorption} \ (\%) = \frac{C_{\text{des}}}{C_{\text{ads}}} \times 100
\]

(17)

where \( C_{\text{des}} \) is the concentration of dye in desorbed and \( C_{\text{ads}} \) (mg/dm$^3$) is the concentration at adsorbed phases.

The amount of MB desorbed was relatively low with the maximum MB desorbed percentage was 13.4% and it was obtained for strong acidic conditions (0.1M HCl). For higher pH values (NaOH) and deionized water, the desorption was very low and limited to 4–7%. The low desorption of MB by an acidic medium shows that the MB molecules were mainly adsorbed onto the ALiCE through strong chemisorption mechanisms. The results indicate that the regeneration of ALiCE by using acid or alkaline solutions is not feasible. However, the reduced colour leachability from the dye-loaded ALiCE is a good property bearing in mind the disposal in landfill or reuse for other purposes for instance: in polymeric composites or incorporation in construction materials.
3.8. Adsorption mechanisms: FT-IR and SEM

The FT-IR spectra in Figure 6 shows structural changes in the ALiCE adsorbents after MB adsorption, confirmed by the changes of functional group bands due to the dye adsorption. The absorption peak around 3435 \text{ cm}^{-1} is due to stretching of phenolic and aliphatic \text{–OH} and \text{–N\&H} groups. The peaks at 2918 and 2850 \text{ cm}^{-1} correspond to C\text{–H} and \text{–CH}_2\text{–} stretching modes. The peak at 1626 \text{ cm}^{-1} is due to bending of \text{–NH} in primary amine group. Symmetric bending of \text{–O–CH}_3 is detected at 1458 \text{ cm}^{-1}. The peaks around 1200 \text{ cm}^{-1} are assigned to C\text{–O–C} stretching in \text{α-O-4} and \text{β-O-4} linkages of alkali lignin \cite{35}. The peak at 671 \text{ cm}^{-1} is attributable to the out-of-plane bending of the O\text{–H} group. The changes in ALiCE surface characteristics i.e. change in all peak heights and shifts of functional group bands after MB loading indicate that groups for example hydroxyl and amine groups are involved in the adsorption process as expected. The SEM images demonstrated noteworthy changes in surface topography between the raw extruded lignin-chitosan blends, the activated extrudates and MB-loaded ALiCE. This deviation in the surface morphology is due to the activation process which has changed the quantities of available functionalities and their interactions. The development of chitosan agglomerates on the lignin surface before activation is obvious at the surface of the blends. This is due to the interactions between \text{β-1,4-glycosidic linkage}, amide and hydroxyl groups of chitosan, and ether, aromatic ring and hydroxyl groups of alkali lignin. After activation, the ALiCE surface was flattened with large, irregular intercellular spaces and well-defined edges. This indicates that the lignin-chitosan blended structure was rearranged during and after the activation. The SEM image of the MB-loaded-ALiCE showed that the edges and intercellular spaces of the microstructure disappeared after the MB adsorption and the structure is more relaxed with a paste like texture. The EDX chemical analysis of the ALiCE indicated that the unloaded ALiCE contained approx. 3.5\% S and 5.7\% Na. The EDX of the MB-loaded ALiCE showed the non-appearance of Na and that no apparent changes in elemental composition took place, signifying that ion-exchange is involved in the MB binding mechanisms. The structure
relaxation could be due to stripping of alkaline elements such as Na which is present in trace quantities originating mainly from alkali lignin. The FT-IR and SEM analysis confirm that the dye adsorption occurs possibly through electrostatic attractions/surface exchanges until the functional sites are completely unavailable; subsequently, MB molecules diffuse into ALiCE particles for more interactions possibly hydrophobic interactions and hydrogen bonding.

4. Concluding remarks
A new method involving extrusion and thermal activation was successfully employed in the production of activated lignin-chitosan extruded (ALiCE) pellets. The ALiCE materials were synthesized, structurally characterized, and used in selective cationic dye (methylene blue (MB)) adsorption. The chemisorption dye adsorption mechanisms were revealed and it was concluded that electrostatic attractions and chemical interactions were observed between amino and hydroxyl groups of the ALiCE adsorbent and amine groups of the dye. Using the Langmuir isotherm a maximum adsorption capacity of 36.25mg/g is calculated. The FTIR, electron microscopy and desorption studies confirmed that strong chemical bonds occurred between the MB molecules and the functional groups on the ALiCE.

Acknowledgement
The authors would like to thanks Mr James McClean from Queen’s University Belfast for his proofreading which has profoundly improved the quality of this article.

5. References


Table 1: Comparison of various adsorbents: sorption capacity for MB adsorption.

<table>
<thead>
<tr>
<th>Adsorbent</th>
<th>Adsorption capacity (mg/g)</th>
<th>Max C₀ (mg/dm³)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Biomass-based activated carbon by FeCl₃ activation</td>
<td>259.2 mg/g</td>
<td>450 mg/dm³</td>
<td>[7]</td>
</tr>
<tr>
<td>Propylene diamine basic activated carbons</td>
<td>182.0 mg/g</td>
<td>700 mg/dm³</td>
<td>[8]</td>
</tr>
<tr>
<td>Recyclable magnetite-loaded palm shell-waste based activated carbon</td>
<td>163.3 mg/g</td>
<td>– –</td>
<td>[9]</td>
</tr>
<tr>
<td>Activated carbon prepared from dross licorice by ultrasonic</td>
<td>82.90 mg/g</td>
<td>200 mg/dm³</td>
<td>[10]</td>
</tr>
<tr>
<td>Steam activated carbon from lantana camara stem</td>
<td>19.84 mg/g</td>
<td>200 mg/dm³</td>
<td>[11]</td>
</tr>
<tr>
<td>Activated lignin-chitosan composite extrudates</td>
<td>36.25 mg/g</td>
<td>82 mg/dm³</td>
<td>This study</td>
</tr>
<tr>
<td>Hydrophobic activated carbons</td>
<td>34.70 mg/g</td>
<td>700 mg/dm³</td>
<td>[8]</td>
</tr>
</tbody>
</table>

Table 2: Calculated parameters for kinetic models for the adsorption of MB on ALiCE at different initial MB concentrations.

<table>
<thead>
<tr>
<th>C₀</th>
<th>qₑₑₚ</th>
<th>qₑ</th>
<th>k₁</th>
<th>R²</th>
<th>qₑ</th>
<th>k₂ × 10²</th>
<th>R²</th>
</tr>
</thead>
<tbody>
<tr>
<td>10.15</td>
<td>4.601</td>
<td>4.266</td>
<td>0.016</td>
<td>0.978</td>
<td>4.448</td>
<td>0.570</td>
<td>0.995</td>
</tr>
<tr>
<td>19.37</td>
<td>8.137</td>
<td>7.377</td>
<td>0.009</td>
<td>0.978</td>
<td>7.817</td>
<td>0.170</td>
<td>0.993</td>
</tr>
<tr>
<td>33.77</td>
<td>14.08</td>
<td>12.81</td>
<td>0.007</td>
<td>0.976</td>
<td>13.89</td>
<td>0.060</td>
<td>0.993</td>
</tr>
<tr>
<td>40.02</td>
<td>16.13</td>
<td>14.58</td>
<td>0.004</td>
<td>0.979</td>
<td>16.36</td>
<td>0.030</td>
<td>0.993</td>
</tr>
<tr>
<td>59.37</td>
<td>22.98</td>
<td>20.70</td>
<td>0.003</td>
<td>0.977</td>
<td>23.43</td>
<td>0.020</td>
<td>0.994</td>
</tr>
<tr>
<td>82.01</td>
<td>27.90</td>
<td>26.44</td>
<td>0.001</td>
<td>0.978</td>
<td>29.04</td>
<td>0.010</td>
<td>0.980</td>
</tr>
</tbody>
</table>

(C₀: mg/L; qₑₑₚ: mg/g; k₁: 1/min; k₂: g/mg min).
Table 3: Intraparticle rate parameters and diffusion coefficients of MB adsorption onto ALiCE at different MB initial concentrations.

<table>
<thead>
<tr>
<th>$C_0$</th>
<th>Intraparticle diffusion, $k_{id}$</th>
<th>Film diffusion, $D_1$</th>
<th>Pore diffusion, $D_2$</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>$\times 10^{10}$</td>
<td>$\times 10^{11}$</td>
<td></td>
</tr>
<tr>
<td>10.15</td>
<td>0.264</td>
<td>1.766</td>
<td>8.371</td>
</tr>
<tr>
<td>19.37</td>
<td>0.477</td>
<td>1.420</td>
<td>5.028</td>
</tr>
<tr>
<td>33.77</td>
<td>0.720</td>
<td>1.116</td>
<td>4.220</td>
</tr>
<tr>
<td>40.02</td>
<td>0.727</td>
<td>0.712</td>
<td>1.936</td>
</tr>
<tr>
<td>59.37</td>
<td>1.046</td>
<td>0.653</td>
<td>1.498</td>
</tr>
<tr>
<td>82.01</td>
<td>1.171</td>
<td>0.581</td>
<td>1.031</td>
</tr>
</tbody>
</table>

($k_{id}: \text{mg/g.min}^{0.5}; D_1 \text{ and } D_2: \text{cm}^2/\text{s}$).

Table 4: Isotherm model parameters for adsorption of MB onto ALiCE.

<table>
<thead>
<tr>
<th>Model</th>
<th>Parameters</th>
<th>$q_{max}$</th>
<th>$b$</th>
<th>$R^2$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Langmuir isotherm</td>
<td></td>
<td>36.25</td>
<td>0.121</td>
<td>0.997</td>
</tr>
<tr>
<td>Freundlich isotherm</td>
<td>$K_F$</td>
<td>7.024</td>
<td>2.472</td>
<td>0.978</td>
</tr>
<tr>
<td>Redlich-Peterson isotherm</td>
<td>$a_R$</td>
<td>4.626</td>
<td>0.146</td>
<td>0.965</td>
</tr>
<tr>
<td>Sips isotherm</td>
<td>$q_{max}$</td>
<td>38.75</td>
<td>0.128</td>
<td>0.898</td>
</tr>
</tbody>
</table>
Figure 1: Effect of the pH values on the adsorption of MB onto ALiCE.

Volume = 25 mL
MB Concentration = 50 ppm
ALiCE Mass = 0.05 g
Figure 2: (A) Pseudo-first-order kinetics and pseudo-second-order kinetic models for the adsorption of MB onto ALiCE, and (B) Plots of the fractional uptake $f(q_t/q_e)$ for MB adsorption onto ALiCE at different concentrations.
Figure 3: The discrepancy of the initial rates of adsorption at different initial MB concentrations.
Figure 4: (A) Intraparticle diffusion model for the adsorption of MB onto ALiCE at various initial MB concentrations and (B) Boyd plots for MB adsorption onto ALiCE at different initial MB concentrations.
Figure 5: (A) Isotherm plots for MB adsorption onto ALiCE and (B) Separation factor, $R_L$, for the adsorption of MB onto ALiCE.
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Wavenumber (1/cm)

Absorbance

- OH, -NH stretching
- C-H stretching
- NH bending
- CH₃ symmetric bending
- C-O-C stretching

Fresh ALICE

MB Loaded ALICE

Unloaded raw lignin-chitosan extruded blend

Chitosan

Lignin
Unloaded activated lignin-chitosan blend

Loaded activated lignin-chitosan blend
Figure 6: FT-IR spectra, SEM images for ALiCE adsorbent before and after MB adsorption and proposed adsorption mechanisms.